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(54) Title: MORPHOGEN-INDUCED MODULATION OF INFLAMMATORY RESPONSE

The present invention is directed to methods and compositions for alleviating tissue destructive effects associated with the inflammatory response to tissue injury in a mammal. The methods and compositions include administering a therapeutically effective concentration of a morphogen or morphogen-stimulating agent sufficient to alleviate immune cell-mediated tissue destruction tion.

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MORPHOGEN-INDUCED MODULATION OF INFLAMMATORY RESPONSE

Field of the Invention

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The present invention relates generally to a method for modulating the inflammatory response induced in a mammal following tissue injury. More particularly, this invention relates to a method for alleviating immune-cell mediated tissue destruction associated with the inflammatory response.

Background of the Invention

The body's inflammatory response to tissue injury 20 can cause significant tissue destruction, leading to loss of tissue function. Damage to cells resulting from the effects of inflammatory response e.g., by immune-cell mediated tissue destruction, has been implicated as the cause of reduced tissue function or 25 loss of tissue function in diseases of the joints (e.g., rheumatoid and osteo-arthritis) and of many organs, including the kidney, pancreas, skin, lung and heart. For example, glomular nephritis, diabetes, inflammatory bowel disease, vascular diseases such as 30 atheroclerosis and vasculitis, and skin diseases such as psoriasis and dermatitis are believed to result in large part from unwanted acute inflammatory reaction A number of these diseases, including and fibrosis. arthritis, psoriasis and inflammatory bowel disease are 35 considered to be chronic inflammatory diseases.

damaged tissue also often is replaced by fibrotic tissue, e.g., scar tissue, which further reduces tissue function. Graft and transplanted organ rejection also is believed to be primarily due to the action of the body's immune/inflammatory response system.

The immune-cell mediated tissue destruction often follows an initial tissue injury or insult. The secondary damage, resulting from the inflammatory response, often is the source of significant tissue damage. Among the factors thought to mediate these damaging effects are those associated with modulating the body's inflammatory response following tissue injury, e.g., cytokines such as interleukin-1 (IL-1) and tumor necrosis factor (TNF), and oxygen-derived free radicals such as superoxide anions. These humoral agents are produced by adhering neutrophilic leukocytes or by endothelial cells and have been identified at ischemic sites upon reperfusion. Moreover, TNF concentrations are increased in humans after myocardial infarction.

A variety of lung diseases are characterized by airway inflammation, including chronic bronchitis, emphysema, idiopathic pulmonary fibrosis and asthma. Another type of lung-related inflammation disorders are inflammatory diseases characterized by a generalized, wide-spread acute inflammatory response such as adult respiratory distress syndrome. Another dysfunction associated with the inflammatory response is that mounted in response to injury caused by hyperoxia, e.g., prolonged exposure to lethally high concentrations of 02 (95-100% 02). Similarly, reduced

blood flow to a tissue (and, therefore reduced or lack of oxygen to tissues), as described below, also can induce a primary tissue injury that stimulates the inflammatory response.

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It is well known that damage occurs to cells in mammals which have been deprived of oxygen. the interruption of blood flow, whether partial (hypoxia) or complete (ischemia) and the ensuing 10 inflammatory responses may be the most important cause of coagulative necrosis or cell death in human disease. The complications of atherosclerosis, for example, are generally the result of ischemic cell injury in the brain, heart, small intestines, kidneys, and lower 15 extremities. Highly differentiated cells, such as the proximal tubular cells of the kidney, cardiac myocytes, and the neurons of the central nervous system, all depend on aerobic respiration to produce ATP, the energy necessary to carry out their specialized 20 functions. When ischemia limits the oxygen supply and ATP is depleted, the affected cells may become irreversibly injured. The ensuing inflammatory responses to this initial injury provide additional insult to the affected tissue. Examples of such 25 hypoxia or ischemia are the partial or total loss of blood supply to the body as a whole, an organ within the body, or a region within an organ, such as occurs in cardiac arrest, pulmonary embolus, renal artery occlusion, coronary occlusion or occlusive stroke.

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The tissue damage associated with ischemiareperfusion injury is believed to comprise both the initial cell damage induced by the deprivation of oxygen to the cell and its subsequent recirculation, as 35 well as the damage caused by the body's response to

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this initial damage. It is thought that reperfusion injury may result in dysfunction to the endothelium of the vasculature as well as injury to the surrounding the vasculature as well as injury to the surrounding tissue. In idiopathic pulmonary fibrosis, for example, scar tissue accumulates on the lung tissue lining, scar tissue accumulates on the lung tissue lining, inhibiting the tissue's elasticity. The tissue damage inhibiting the tissue's elasticity is believed to follow associated with hyperoxia injury is believed to follow a similar mechanism, where the initial damage is mediated primarily through the presence of toxic oxygen mediated primarily through the presence of toxic oxygen this initial injury.

Similarly, tissues and organs for transplantation also are subject to the tissue destructive effects associated with the recipient host body's inflammatory response following transplantation. It is currently believed that the initial destructive response is due in large part to reperfusion injury to the transplanted in large part to reperfusion to the organ organ after it has been transplanted to the organ recipient.

Accordingly, the success of organ or tissue transplantation depends greatly on the preservation of the tissue activity (e.g., tissue or organ viability) at the harvest of the organ, during storage of the harvested organ, and at transplantation. To date, preservation of organs such as lungs, pancreas, heart preservation of organs such as lungs, pancreas, heart and liver remains a significant stumbling block to the successful transplantation of these organs. U.S. Patent No. 4,952,409 describes a superoxide dismutase-containing liposome to inhibit reperfusion injury. U.S. Patent No. 5,002,965 describes the use of ginkolides, known platelet activating factor antagonists, to inhibit reperfusion injury. Both of these factors are described working primarily by

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inhibiting the release of and/or inhibiting the damaging effects of free oxygen radicals. A number of patents also have issued on the use of immunosuppressants for inhibiting graft rejection. A 5 representative listing includes U.S. Patent Nos. 5,104,858, 5,008,246 and 5,068,323. A significant problem with many immunosuppressants is their low therapeutic index, requiring the administration of high doses that can have significant toxic side effects.

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Rheumatoid and osteoarthritis are prevalent diseases characterized by chronic inflammation of the synovial membrane lining the afflicted joint. A major consequence of chronic inflammatory joint disease 15 (e.g., rheumatoid arthritis) and degenerative arthritis (e.g., osteoarthritis) is loss of function of those affected joints. This loss of function is due primarily to destruction of the major structural components of the joint, cartilage and bone, and 20 subsequent loss of the proper joint anatomy. As a consequence of chronic disease, joint destruction ensues and can lead to irreversible and permanent damage to the joint and loss of function. Current treatment methods for severe cases of rheumatoid 25 arthritis typically include the removal of the synovial membrane, e.g., synovectomy. Surgical synovectomy has many limitations, including the risk of the surgical procedure itself, and the fact that a surgeon often cannot remove all of the diseased membrane. The 30 diseased tissue remaining typically regenerates, causing the same symptoms which the surgery was meant to alleviate.

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Psoriasis is a chronic, recurrent, scaling skin disease of unknown etiology characterized by chronic inflammation of the skin. Erythematous eruptions, often in papules or plaques, and usually having a white 5 silvery scale, can affect any part of the skin, but most commonly affect the scalp, elbows, knees and lower back. The disease usually occurs in adults, but children may also be affected. Patients with psoriasis have a much greater incidence of arthritis (psoraitic 10 arthritis), and generalized exfoliation and even death can threaten afflicted individuals.

Current therapeutic regimens include topical or intralesional application of corticosteroids, topical 15 administration of keratolytics, and use of tar and UV light on affected areas. No single therapy is ideal, and it is rare for a patient not to be treated with several alternatives during the relapsing and remitting course of the disease. Whereas systematic treatment 20 can induce prompt resolution of psoriatic lesions, suppression often requires ever-increasing doses, sometimes with toxic side effect, and tapering of therapy may result in rebound phenomena with extensions of lesions, possibly to exfoliation.

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Inflammatory bowel disease (IBD) describes a class of clinical disorders of the gastrointestinal mucosa characterized by chronic inflammation and severe ulceration of the mucosa. The two major diseases in 30 this classification are ulcerative colitis and regional enteritis (Crohn's Disease). Like oral mucositis, the diseases classified as IBD are associated with severe mucosal ulceration (frequently penetrating the wall of the bowel and forming strictures and fistulas), severe 35 mucosal and submucosal inflammation and edema, and

fibrosis (e.g., scar tissue formation which interferes with the acid protective function of the gastrointestinal lining.) Other forms of IBD include regional ileitis and proctitis. Clinically, patients with fulminant IBD can be severely ill with massive diarrhea, blood loss, dehydration, weight loss and fever. The prognosis of the disease is not good and frequently requires resection of the diseased tissue.

Therefore, an object of the present invention is to provide a method for protecting mammalian tissue, 10 particularly human tissue, from the damage associated with the inflammatory response following a tissue injury. The inflammatory reaction may be in response 15 to an initial tissue injury or insult. The original injury may be chemically, mechanically, biologically or immunologically related. Another object is to provide methods and compositions for protecting tissue from the tissue destructive effects associated with chronic 20 inflammatory diseases, including arthritis (e.g., reheumatoid or osteoarthritis), psoriatic arthritis, psoriasis and dermatitis, inflammatory bowel disease and other autoimmune diseases. Yet another object is to provide methods and compositions for enhancing the 25 viability of mammalian tissues and organs to be transplanted, including protecting the transplanted organs from immune cell-mediated tissue destruction, such as the tissue damage associated with ischemiareperfusion injury. This tissue damage may occur 30 during donor tissue or organ harvesting and transport, as well as following initiation of blood flow after transplantation of the organ or tissue in the recipient host.

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Another object of the invention is to provide a method for alleviating tissue damage associated with ischemic-reperfusion injury in a mammal following a deprivation of oxygen to a tissue in the mammal. Other objects of the present invention include providing a method for alleviating tissue damage associated with ischemic-reperfusion injury in a human which has suffered from hypoxia or ischemia following cardiac suffered from hypoxia or ischemia following cardiac arrest, pulmonary embolus, renal artery occlusion, coronary occlusion or occlusive stroke. A further object is to provide a method for alleviating tissue damage associated with hyperoxia-induced tissue injury, e.g., lethally high oxygen concentrations.

- Still another object of the invention is to provide a method for modulating inflammatory responses in general, particularly those induced in a human following tissue injury.
- These and other objects and features of the invention will be apparent from the description, drawings and claims which follow.

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Summary of the Invention

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The present invention provides a method for alleviating the tissue destructive effects associated 5 with activation of the inflammatory response following tissue injury. The method comprises the step of providing to the affected tissue a therapeutically effective concentration of a morphogenic protein ("morphogen", as defined herein) upon tissue injury or 10 in anticipation of tissue injury, sufficient to substantially inhibit or reduce the tissue destructive effects of the inflammatory response.

In one aspect, the invention features compositions 15 and therapeutic treatment methods that comprise the step of administering to a mammal a therapeutically effective amount of a morphogenic protein ("morphogen"), as defined herein, upon injury to a tissue, or in anticipation of such injury, for a time and at a concentration sufficient to inhibit the tissue destructive effects associated with the body's inflammatory response, including repairing damaged tissue, and/or inhibiting additional damage thereto.

In another aspect, the invention features compositions and therapeutic treatment methods for 25 protecting tissues and organs from the tissue destructive effects of the inflammatory response which include administering to the mammal, upon injury to a 30 tissue or in anticipation of such injury, a compound that stimulates <u>in vivo</u> a therapeutically effective concentration of an endogenous morphogen within the body of the mammal sufficient to protect the tissue from the tissue destructive effects associated with the 35 inflammatory response, including repairing damaged

tissue and/or inhibiting additional damage thereto.

These compounds are referred to herein as morphogenstimulating agents, and are understood to include
substances which, when administered to a mammal, act on
cells of tissue(s) or organ(s) that normally are
responsible for, or capable of, producing a morphogen
and/or secreting a morphogen, and which cause the
endogenous level of the morphogen to be altered. The
agent may act, for example, by stimulating expression
and/or secretion of an endogenous morphogen.

As embodied herein, the term "ischemic-reperfusion injury" refers to the initial damage associated with oxygen deprivation of a cell and the subsequent damage 15 associated with the inflammatory response when the cell is resupplied with oxygen. As embodied herein, the term "hyperoxia-induced injury" refers to the tissue damage associated with prolonged exposure to lethally high doses of oxygen, e.g., greater than 95% 0_2 , 20 including the tissue damage associated with the inflammatory response to the toxically high oxygen dose. Accordingly, as used herein, "toxic oxygen concentrations" refers to the tissue damage associated withthe injury induced by both lethally low oxygen 25 concentrations of oxygen (including a complete lack of oxygen), and by lethally high oxygen concentrations. The expression "alleviating" means the protection from, reduction of and/or elimination of undesired tissue destruction, particularly immune cell-mediated tissue 30 destruction. The tissue destruction may be in response to an initial tissue injury, which may be mechanical, chemical or immunological in origin. The expression "enhance the viability of" living tissues or organs, as used herein, means protection from, reduction of and/or 35 elimination of reduced or lost tissue or organ function ŧ

as a result of tissue death, particularly immune cellmediated tissue death. "Transplanted" living tissue
encompasses both tissue transplants (e.g., as in the
case of bone marrow transplants) and tissue grafts.

5 Finally, a "free oxygen radical inhibiting agent" means
a molecule capable of inhibiting the release of and/or
inhibiting tissue damaging effects of free oxygen
radicals.

In one embodiment of the invention, the invention provides methods and compositions for alleviating the 10 ischemic-reperfusion injury in mammalian tissue resulting from a deprivation of, and subsequent In another reperfusion of, oxygen to the tissue. 15 embodiment, the invention provides a method for alleviating the tissue-destructive effects associated with hyperoxia. In still another embodiment of the invention, the invention provides methods and compositions for maintaining the viability of tissues 20 and organs, particularly living tissues and organs to be transplanted, including protecting them from ischemia-reperfusion injury. In still another embodiment, the invention provides methods for protecting tissues and organs from the tissue 25 destructive effects of chronic inflammatory diseases, such as arthritis, psoriasis, dermatitis, including contact dermatitis, IBD and other chronic inflammatory diseases of the gastrointestinal tract, as well as the tissue destructive effects associated with other, known 30 autoimmune diseases, such as diabetes, multiple sclerosis, amyotrophic lateral sclerosis (ALS), and other autoimmune neurodegenerative diseases.

In one aspect of the invention, the morphogen is provided to the damaged tissue following an initial injury to the tissue. The morphogen may be provided directly to the tissue, as by injection to the damaged tissue site or by topical administration, or may be provided indirectly, e.g., systemically by oral or parenteral means. Alternatively, as described above, an agent capable of stimulating endogenous morphogen expression and/or secretion may be administered to the mammal. Preferably, the agent can stimulate an endogenous morphogen in cells associated with the damaged tissue. Alternatively, morphogen expression and/or secretion may be stimulated in a distant tissue and the morphogen transported to the damaged tissue by the circulatory system.

In another aspect of the invention, the morphogen is provided to tissue at risk of damage due to immune cell-mediated tissue destruction. Examples of such tissues include tissue grafts and tissue or organ transplants, as well as any tissue or organ about to undergo a surgical procedure or other clinical procedure likely to either inhibit blood flow to the tissue or otherwise induce an inflammatory response.

Bere the morphogen or morphogen-stimulating agent preferably is provided to the patient prior to induction of the injury, e.g., as a prophylactic, to provide a cyto-protective effect to the tissue at risk.

organ to be transplanted, the tissue or organ to be transplanted preferably is exposed to a morphogen prior to transplantation. Most preferably, the tissue or organ is exposed to the morphogen prior to its removal from the donor, by providing the donor with a

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composition comprising a morphogen or morphogenstimulating agent. Alternatively or, in addition, once removed from the donor, the organ or tissue is placed in a preservation solution containing a morphogen or 5 morphogen-stimulating agent. In addition, the recipient also preferably is provided with a morphogen or morphogen-stimulating agent just prior to, or concommitant with, transplantation. In all cases, the morphogen or morphogen-stimulating agent may be 10 administered directly to the tissue at risk, as by injection or topical administration to the tissue, or it may be provided systemically, either by oral or parenteral administration.

The morphogens described herein are envisioned to be useful in enhancing viability of any organ or living 15 tissue to be transplanted. The morphogens may be used to particular advantage in lung, heart, liver, kidney or pancreas transplants, as well as in transplantation 20 and/or grafting of bone marrow, skin, gastrointestinal mucosa, and other living tissues.

Where the patient suffers from a chronic inflammatory disease, such as diabetes, arthritis, 25 psoriasis, IBD, and the like, the morphogen or morphogen-stimulating agent preferably is administered at regular intervals as a prophylactic, to prevent and/or inhibit the tissue damage normally associated with the disease during flare periods. As above, the 30 morphogen or morphogen-stimulating agent may be provided directly to the tissue at risk, for example by injection or by topical administration, or indirectly, as by systemic e.g., oral or parenteral administration.

Among the morphogens useful in this invention are proteins originally identified as osteogenic proteins, such as the OP-1, OP-2 and CBMP2 proteins, as well as amino acid sequence-related proteins such as DPP (from 5 Drosophila), Vgl (from Xenopus), Vgr-1 (from mouse, see U.S. 5,011,691 to Oppermann et al.), GDF-1 (from mouse, see Lee (1991) PNAS 88:4250-4254), all of which are presented in Table II and Seq. ID Nos.5-14), and the recently identified 60A protein (from Drosophila, Seq. 10 ID No. 24, see Wharton et al. (1991) PNAS 88:9214-9218.) The members of this family, which include members of the TGF- β super-family of proteins, share substantial amino acid sequence homology in their The proteins are translated as a C-terminal regions. 15 precursor, having an N-terminal signal peptide sequence, typically less than about 30 residues, followed by a "pro" domain that is cleaved to yield the mature sequence. The signal peptide is cleaved rapidly upon translation, at a cleavage site that can be 20 predicted in a given sequence using the method of Von Heijne ((1986) Nucleic Acids Research 14:4683-4691.) Table I, below, describes the various morphogens identified to date, including their nomenclature as used herein, their Seq. ID references, and publication 25 sources for the amino acid sequences for the full length proteins not included in the Seq. Listing. disclosure of these publications is incorporated herein by reference. TABLE I

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"OP-1" Refers generically to the group of
morphogenically active proteins expressed
from part or all of a DNA sequence
encoding OP-1 protein, including allelic
and species variants thereof, e.g., human

OP-1 ("hOP-1", Seq. ID No. 5, mature protein amino acid sequence), or mouse OP-1 ("mOP-1", Seq. ID No. 6, mature protein amino acid sequence.) conserved seven cysteine skeleton is defined by residues 38 to 139 of Seq. ID 5 The cDNA sequences and the Nos. 5 and 6. amino acids encoding the full length proteins are provided in Seq. Id Nos. 16 and 17 (hOP1) and Seq. ID Nos. 18 and 19 (mOP1.) The mature proteins are defined 10 by residues 293-431 (hOP1) and 292-430 (mOP1). The "pro" regions of the proteins, cleaved to yield the mature, morphogenically active proteins are defined essentially by residues 30-292 15 (hOP1) and residues 30-291 (mOP1). refers generically to the group of active "OP-2" proteins expressed from part or all of a DNA sequence encoding OP-2 protein, 20 including allelic and species variants thereof, e.g., human OP-2 ("hOP-2", Seq. ID No. 7, mature protein amino acid sequence) or mouse OP-2 ("mOP-2", Seq. ID 25 No. 8, mature protein amino acid sequence). The conserved seven cysteine skeleton is defined by residues 38 to 139 of Seq. ID Nos. 7 and 8. The CDNA sequences and the amino acids encoding the full length proteins are provided in Seq. 30 ID Nos. 20 and 21 (hOP2) and Seq. ID Nos. 22 and 23 (mOP2.) The mature proteins are defined essentially by residues 264-402 (hOP2) and 261-399 (mOP2). The "pro"

regions of the proteins, cleaved to yield the mature, morphogenically active proteins likely are defined essentially by residues 18-263 (hOP2) and residues 18-260 (mOP2). (Another cleavage site also occurs 21 residues upstream for both OP-2 5 proteins.) refers generically to the morphogenically active proteins expressed from a DNA "CBMP2" sequence encoding the CBMP2 proteins, 10 including allelic and species variants thereof, e.g., human CBMP2A ("CBMP2A(fx)", Seq ID No. 9) or human CBMP2B DNA ("CBMP2B(fx)", Seq. ID No. 10). The amino acid sequence for the full length 15 proteins, referred to in the literature as BMP2A and BMP2B, or BMP2 and BMP4, appear in Wozney, et al. (1988) Science 242:1528-1534. The pro domain for BMP2 (BMP2A) likely includes residues 25-248 or 25-282; 20 the mature protein, residues 249-396 or 283-396. The pro domain for BMP4 (BMP2B) likely includes residues 25-256 or 25-292; the mature protein, residues 257-408 or 25 293-408refers to protein sequences encoded by the Drosophila DPP gene and defining the "DPP(fx)" conserved seven cysteine skeleton (Seq. ID No. 11). The amino acid sequence for the 30 full length protein appears in Padgett, et al (1987) Nature 325: 81-84. The pro domain likely extends from the signal peptide cleavage site to residue 456; the mature protein likely is defined by residues 457-588.

refers to protein sequences encoded by the "Vgl(fx)" Xenopus Vgl gene and defining the conserved seven cysteine skeleton (Seq. ID No. 12). The amino acid sequence for the full length protein appears in Weeks (1987) Cell 51: 861-867. prodomain likely extends from the signal peptide cleavage site to residue 246; the mature protein likely is defined by residues 247-360.

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refers to protein sequences encoded by the "Var-1(fx)" murine Vgr-1 gene and defining the conserved seven cysteine skeleton (Seq. ID 20 No. 13). The amino acid sequence for the full length protein appears in Lyons, et al, (1989) PNAS 86: 4554-4558. prodomain likely extends from the signal peptide cleavage site to residue 299; the 25 mature protein likely is defined by residues 300-438.

refers to protein sequences encoded by the "GDF-1(fx)" human GDF-1 gene and defining the 30 conserved seven cysteine skeleton (Seq. ID No. 14). The cDNA and encoded amino sequence for the full length protein is

provided in Seq. ID. No. 32. The prodomain likely extends from the signal peptide clavage site to residue 214; the mature protein likely is defined by residues 215-372.

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refers generically to the morphogenically active proteins expressed from part or all of a DNA sequence (from the Drosophila 60A gene) encoding the 60A proteins (see Seq. ID No. 24 wherein the cDNA and encoded amino acid sequence for the full length protein is provided). "60A(fx)" refers to the protein sequences defining the conserved seven cysteine skeleton (residues 354 to 455 of Seq. ID No. 24.) The prodomain likely extends from the signal peptide cleavage site to residue 324; the mature protein likely is defined by residues 325-455.

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"BMP3(fx)" refers to protein sequences encoded by the human BMP3 gene and defining the conserved seven cysteine skeleton (Seq. ID No. 26).

The amino acid sequence for the full length protein appears in Wozney et al.

(1988) Science 242: 1528-1534. The prodomain likely extends from the signal peptide cleavage site to residue 290; the mature protein likely is defined by residues 291-472.

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refers to protein sequences encoded by the "BMP5(fx)" human BMP5 gene and defining the conserved seven cysteine skeleton (Seq. ID No. 27). The amino acid sequence for the full length protein appears in Celeste, et al. (1991) PNAS 87: 9843-9847. The pro domain likely extends from the signal peptide cleavage site to residue 316; the mature protein likely is defined by residues 317-454.

refers to protein sequences encoded by the "BMP6(fx)" human BMP6 gene and defining the conserved seven cysteine skeleton (Seq. ID No. 28). The amino acid sequence for the full length protein appears in Celeste, et al. (1990) PNAS 87: 9843-5847. The pro domain likely includes extends from the signal peptide cleavage site to residue 374; the mature sequence likely includes 20 residues 375-513.

The OP-2 proteins have an additional cysteine 25 residue in this region (e.g., see residue 41 of Seq. ID Nos. 7 and 8), in addition to the conserved cysteine skeleton in common with the other proteins in this family. The GDF-1 protein has a four amino acid insert within the conserved skeleton (residues 44-47 of Seq. ID No. 14) but this insert likely does not interfere with the relationship of the cysteines in the folded structure. In addition, the CBMP2 proteins are missing one amino acid residue within the cysteine skeleton.

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The morphogens are inactive when reduced, but are active as oxidized homodimers and when oxidized in combination with other morphogens of this invention Thus, as defined herein, a (e.g., as heterodimers). 5 morphogen is a dimeric protein comprising a pair of polypeptide chains, wherein each polypeptide chain comprises at least the C-terminal six cysteine skeleton defined by residues 43-139 of Seq. ID No. 5, including functionally equivalent arrangements of these cysteines 10 (e.g., amino acid insertions or deletions which alter the linear arrangement of the cysteines in the sequence but not their relationship in the folded structure), such that, when the polypeptide chains are folded, the dimeric protein species comprising the pair of 15 polypeptide chains has the appropriate threedimensional structure, including the appropriate intraor inter-chain disulfide bonds such that the protein is capable of acting as a morphogen as defined herein. Specifically, the morphogens generally are capable of 20 all of the following biological functions in a morphogenically permissive environment: stimulating proliferation of progenitor cells; stimulating the differentiation of progenitor cells; stimulating the proliferation of differentiated cells; and supporting 25 the growth and maintenance of differentiated cells, including the "redifferentiation" of transformed cells. In addition, it is also anticipated that these morphogens are capable of inducing redifferentiation of committed cells under appropriate environmental 30 conditions.

In one preferred aspect, the morphogens of this invention comprise one of two species of generic amino acid sequences: Generic Sequence 1 (Seq. ID No. 1) or Generic Sequence 2 (Seq. ID No. 2); where each Xaa

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indicates one of the 20 naturally-occurring L-isomer, α -amino acids or a derivative thereof. Generic Sequence 1 comprises the conserved six cysteine skeleton and Generic Sequence 2 comprises the conserved six cysteine skeleton plus the additional cysteine identified in OP-2 (see residue 36, Seq. ID No. 2). another preferred aspect, these sequences further comprise the following additional sequence at their Nterminus:

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Cys Xaa Xaa Xaa Xaa (Seq. ID No. 15) 1

Preferred amino acid sequences within the 15 foregoing generic sequences include: Generic Sequence 3 (Seq. ID No. 3), Generic Sequence 4 (Seq. ID No. 4), Generic Sequence 5 (Seq. ID No. 30) and Generic Sequence 6 (Seq. ID No. 31), listed below. Generic Sequences accommodate the homologies shared 20 among the various preferred members of this morphogen family identified in Table II, as well as the amino acid sequence variation among them. Specifically, Generic Sequences 3 and 4 are composite amino acid sequences of the following proteins presented in 25 Table II and identified in Seq. ID Nos. 5-14: human OP-1 (hOP-1, Seq. ID Nos. 5 and 16-17), mouse OP-1 (mOP-1, Seq. ID Nos. 6 and 18-19), human and mouse OP-2 (Seq. ID Nos. 7, 8, and 20-22), CBMP2A (Seq. ID No. 9), CBMP2B (Seq. ID No. 10), DPP (from Drosophila, Seq. ID 30 No. 11), Vgl, (from Xenopus, Seq. ID No. 12), Vgr-1 (from mouse, Seq. ID No. 13), and GDF-1 (from mouse, Seq. ID No. 14.) The generic sequences include both the amino acid identity shared by the sequences in Table II, as well as alternative residues for the 35 variable positions within the sequence. Note that

these generic sequences allow for an additional cysteine at position 41 or 46 in Generic Sequences 3 or 4, respectively, providing an appropriate cysteine skeleton where inter- or intramolecular disulfide bonds can form, and contain certain critical amino acids which influence the tertiary structure of the proteins.

Generic Sequence 3

Leu Tyr Val Xaa Phe

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Xaa Xaa Xaa Gly Trp Xaa Xaa Trp Xaa

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Xaa Ala Pro Xaa Gly Xaa Xaa Ala

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15 Xaa Tyr Cys Xaa Gly Xaa Cys Xaa

Xaa Pro Xaa Xaa Xaa Xaa Xaa

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Xaa Xaa Xaa Asn His Ala Xaa Xaa

20 40

Xaa Xaa Leu Xaa Xaa Xaa Xaa

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Xaa Xaa Xaa Xaa Xaa Xaa Cys

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25 Cys Xaa Pro Xaa Xaa Xaa Xaa Xaa

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Xaa Xaa Xaa Leu Xaa Xaa Xaa

70 75

Xaa Xaa Xaa Xaa Val Xaa Leu Xaa

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5 Xaa Xaa Xaa Xaa Met Xaa Val Xaa

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Xaa Cys Gly Cys Xaa

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wherein each Xaa is independently selected from a group 10 of one or more specified amino acids defined as follows: "Res." means "residue" and Xaa at res.4 = (Ser, Asp or Glu); Xaa at res.6 = (Arg, Gln, Ser or Lys); Xaa at res.7 = (Asp or Glu); Xaa at res.8 = (Leu or Val); Xaa at res.11 = (Gln, Leu, Asp, His or Asn); 15 Xaa at res.12 = (Asp, Arg or Asn); Xaa at res.14 = (Ile or Val); Xaa at res.15 = (Ile or Val); Xaa at res.18 = (Glu, Gln, Leu, Lys, Pro or Arg); Xaa at res.20 = (Tyr or Phe); Xaa at res.21 = (Ala, Ser, Asp, Met, His, Leu or Gln); Xaa at res.23 = (Tyr, Asn or Phe); Xaa at 20 res.26 = (Glu, His, Tyr, Asp or Gln); Xaa at res.28 = (Glu, Lys, Asp or Gln); Xaa at res.30 = (Ala, Ser, Pro or Gln); Xaa at res.31 = (Phe, Leu or Tyr); Xaa at res.33 = (Leu or Val); Xaa at res.34 = (Asn, Asp, Ala or Thr); Xaa at res.35 = (Ser, Asp, Glu, Leu or Ala); 25 Xaa at res.36 = (Tyr, Cys, His, Ser or Ile); Xaa at res.37 = (Met, Phe, Gly or Leu); Xaa at res.38 = (Asn or Ser); Xaa at res.39 = (Ala, Ser or Gly); Xaa at res.40 = (Thr, Leu or Ser); Xaa at res.44 = (Ile or Val); Xaa at res.45 = (Val or Leu); Xaa at res.46 = (Gln or Arg); Xaa at res.47 = (Thr, Ala or Ser); Xaa at res.49 = (Val or Met); Xaa at res.50 = (His or Asn); Xaa at res.51 = (Phe, Leu, Asn, Ser, Ala or Val); Xaa

at res.52 = (Ile, Met, Asn, Ala or Val); Xaa at res.53 = (Asn, Lys, Ala or Glu); Xaa at res.54 = (Pro or Ser); Xaa at res.55 = (Glu, Asp, Asn, or Gly); Xaa at res.56 = (Thr, Ala, Val, Lys, Asp, Tyr, Ser or Ala); Xaa at 5 res.57 = (Val, Ala or Ile); Xaa at res.58 = (Pro or Asp); Xaa at res.59 = (Lys or Leu); Xaa at res.60 = (Pro or Ala); Xaa at res.63 = (Ala or Val); Xaa at res.65 = (Thr or Ala); Xaa at res.66 = (Gln, Lys, Arg or Glu); Xaa at res.67 = (Leu, Met or Val); Xaa at 10 res.68 = (Asn, Ser or Asp); Xaa at res.69 = (Ala, Pro or Ser); Xaa at res.70 = (Ile, Thr or Val); Xaa at res.71 = (Ser or Ala); Xaa at res.72 = (Val or Met); Xaa at res.74 = (Tyr or Phe); Xaa at res.75 = (Phe, Tyr or Leu); Xaa at res.76 = (Asp or Asn); Xaa at res.77 = 15 (Asp, Glu, Asn or Ser); Xaa at res.78 = (Ser, Gln, Asn or Tyr); Xaa at res.79 = (Ser, Asn, Asp or Glu); Xaa at res.80 = (Asn, Thr or Lys); Xaa at res.82 = (Ile or Val); Xaa at res.84 = (Lys or Arg); Xaa at res.85 = (Lys, Asn, Gln or His); Xaa at res.86 = (Tyr or His); 20 Xaa at res.87 = (Arg, Gln or Glu); Xaa at res.88 = (Asn, Glu or Asp); Xaa at res.90 = (Val, Thr or Ala); Xaa at res.92 = (Arg, Lys, Val, Asp or Glu); Xaa at res.93 = (Ala, Gly or Glu); and Xaa at res.97 = (His or Arg);

Generic Sequence 4

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 Xaa
 Pro
 Xaa
 Cys

 Xaa
 Xaa
 Xaa
 Xaa
 Xaa
 Xaa
 Xaa
 Cys

 Cys
 Xaa
 Pro
 Xaa
 X

Xaa Cys Gly Cys Xaa 100 wherein each Xaa is independently selected from a group 20 of one or more specified amino acids as defined by the following: "Res." means "residue" and Xaa at res.2 = (Lys or Arg); Xaa at res.3 = (Lys or Arg); Xaa at res.4 = (His or Arg); Xaa at res.5 = (Glu, Ser, His, Gly, Arg or Pro); Xaa at res.9 = (Ser, Asp or Glu); Xaa at 25 res.11 = (Arg, Gln, Ser or Lys); Xaa at res.12 = (Asp or Glu); Xaa at res.13 = (Leu or Val); Xaa at res.16 = (Gln, Leu, Asp, His or Asn); Xaa at res.17 = (Asp, Arg, or Asn); Xaa at res.19 = (Ile or Val); Xaa at res.20 = (Ile or Val); Xaa at res.23 = (Glu, Gln, Leu, Lys, Pro or Arg); Xaa at res.25 = (Tyr or Phe); Xaa at res.26 = (Ala, Ser, Asp, Met, His, Leu, or Gln); Xaa at res.28 = (Tyr, Asn or Phe); Xaa at res.31 = (Glu, His, Tyr, Asp or Gln); Xaa at res.33 = Glu, Lys, Asp or Gln); Xaa at res.35 = (Ala, Ser or Pro); Xaa at res.36 = (Phe, Leu 35 or Tyr); Xaa at res.38 = (Leu or Val); Xaa at res.39 =

(Asn, Asp, Ala or Thr); Xaa at res.40 = (Ser, Asp, Glu, Leu or Ala); Xaa at res.41 = (Tyr, Cys, His, Ser or Ile); Xaa at res.42 = (Met, Phe, Gly or Leu); Xaa at res.44 = (Ala, Ser or Gly); Xaa at res.45 = (Thr, Leu 5 or Ser); Xaa at res.49 = (Ile or Val); Xaa at res.50 = (Val or Leu); Xaa at res.51 = (Gln or Arg); Xaa at res.52 = (Thr, Ala or Ser); Xaa at res.54 = (Val or Met); Xaa at res.55 = (His or Asn); Xaa at res.56 = (Phe, Leu, Asn, Ser, Ala or Val); Xaa at res.57 = (Ile, 10 Met, Asn, Ala or Val); Xaa at res.58 = (Asn, Lys, Ala or Glu); Xaa at res.59 = (Pro or Ser); Xaa at res.60 = (Glu, Asp, or Gly); Xaa at res.61 = (Thr, Ala, Val, Lys, Asp, Tyr, Ser or Ala); Xaa at res.62 = (Val, Ala or Ile); Xaa at res.63 = (Pro or Asp); Xaa at res.64 = 15 (Lys or Leu); Xaa at res.65 = (Pro or Ala); Xaa at res.68 = (Ala or Val); Xaa at res.70 = (Thr or Ala); Xaa at res.71 = (Gln, Lys, Arg or Glu); Xaa at res.72 = (Leu, Met or Val); Xaa at res.73 = (Asn, Ser or Asp); Xaa at res.74 = (Ala, Pro or Ser); Xaa at res.75 = 20 (Ile, Thr or Val); Xaa at res.76 = (Ser or Ala); Xaa at res.77 = (Val or Met); Xaa at res.79 = (Tyr or Phe); Xaa at res.80 = (Phe, Tyr or Leu); Xaa at res.81 = (Asp or Asn); Xaa at res.82 = (Asp, Glu, Asn or Ser); Xaa at res.83 = (Ser, Gln, Asn or Tyr); Xaa at res.84 = (Ser, 25 Asn, Asp or Glu); Xaa at res.85 = (Asn, Thr or Lys); Xaa at res.87 = (Ile or Val); Xaa at res.89 = (Lys or Arg); Xaa at res.90 = (Lys, Asn, Gln or His); Xaa at res.91 = (Tyr or His); Xaa at res.92 = (Arg, Gln or Glu); Xaa at res.93 = (Asn, Glu or Asp); Xaa at res.95 30 = (Val, Thr or Ala); Xaa at res.97 = (Arg, Lys, Val, Asp or Glu); Xaa at res.98 = (Ala, Gly or Glu); and Xaa at res.102 = (His or Arg).

Similarly, Generic Sequence 5 (Seq. ID No. 30) and Generic Sequence 6 (Seq. ID No. 31) accommodate the homologies shared among all the morphogen protein Specifically, family members identified in Table II. 5 Generic Sequences 5 and 6 are composite amino acid sequences of human OP-1 (hOP-1, Seq. ID Nos. 5 and 16-17), mouse OP-1 (mOP-1, Seq. ID Nos. 6 and 18-19), human and mouse OP-2 (Seq. ID Nos. 7, 8, and 20-22), CBMP2A (Seq. ID No. 9), CBMP2B (Seq. ID No. 10), DPP 10 (from Drosophila, Seq. ID No. 11), Vgl, (from Xenopus, Seq. ID No. 12), Vgr-1 (from mouse, Seq. ID No. 13), and GDF-1 (from mouse, Seq. ID No. 14), human BMP3 (Seq. ID No. 26), human BMP5 (Seq. ID No. 27), human BMP6 (Seq. ID No. 28) and 60(A) (from Drosophila, Seq. 15 ID Nos. 24-25). The generic sequences include both the amino acid identity shared by these sequences in the C-terminal domain, defined by the six and seven cysteine skeletons (Generic Sequences 5 and 6, respectively), as well as alternative residues for the 20 variable positions within the sequence. As for Generic Sequences 3 and 4, Generic Sequences 5 and 6 allow for an additional cysteine at position 41 (Generic Sequence 5) or position 46 (Generic Sequence 6), providing an appropriate cysteine skeleton where inter- or 25 intramolecular disulfide bonds can form, and containing certain critical amino acids which influence the tertiary structure of the proteins.

Generic Sequence 5

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3.

Leu Xaa Xaa Xaa Phe

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Xaa Xaa Xaa Gly Trp Xaa Xaa Trp Xaa

	ves ves Vas Ala						
	Xaa Xaa Pro Xaa Xaa Xaa Ala						
	15 20						
	Xaa Tyr Cys Xaa Gly Xaa Cys Xaa						
	2 5						
5	Xaa Pro Xaa Xaa Xaa Xaa						
•	35						
	Xaa Xaa Xaa Asn His Ala Xaa Xaa						
	40 45						
	Xaa Xaa Xaa Xaa Xaa Xaa Xaa						
	Xaa						
10							
	Xaa Xaa Xaa Xaa Xaa Xaa Cys						
	55						
	Cys Xaa Pro Xaa Xaa Xaa Xaa Xaa						
	65						
15	Xaa Xaa Xaa Leu Xaa Xaa Xaa						
	70 ⁷⁵						
	Xaa Xaa Xaa Val Xaa Leu Xaa						
	80						
	Xaa Xaa Xaa Met Xaa Val Xaa						
	90						
20	85						
	Xaa Cys Xaa Cys Xaa						
	95						
	wherein each Xaa is independently selected from a group of one or more specified amino acids defined as						
	" "recinie" and acc						
25	VAL UL ##\\!						
	= (Ser, Asp or Glu); Xaa at res.8 or Ala); Xaa at res.7 = (Asp, Glu or Lys); Xaa at res.8						

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= (Leu, Val or Ile); Xaa at res.11 = (Gln, Leu, Asp, His, Asn or Ser); Xaa at res.12 = (Asp, Arg, Asn or Glu); Xaa at res.14 = (Ile or Val); Xaa at res.15 = (Ile or Val); Xaa at res.16 (Ala or Ser); Xaa at res.18 5 = (Glu, Gln, Leu, Lys, Pro or Arg); Xaa at res.19 = (Gly or Ser); Xaa at res.20 = (Tyr or Phe); Xaa at res.21 = (Ala, Ser, Asp, Met, His, Gln, Leu or Gly); Xaa at res.23 = (Tyr, Asn or Phe); Xaa at res.26 = (Glu, His, Tyr, Asp, Gln or Ser); Xaa at res.28 = (Glu, 10 Lys, Asp, Gln or Ala); Xaa at res.30 = (Ala, Ser, Pro, Gln or Asn); Xaa at res.31 = (Phe, Leu or Tyr); Xaa at res.33 = (Leu, Val or Met); Xaa at res.34 = (Asn, Asp, Ala, Thr or Pro); Xaa at res.35 = (Ser, Asp, Glu, Leu, Ala or Lys); Xaa at res.36 = (Tyr, Cys, His, Ser or 15 Ile); Xaa at res.37 = (Met, Phe, Gly or Leu); Xaa at res.38 = (Asn, Ser or Lys); Xaa at res.39 = (Ala, Ser, Gly or Pro); Xaa at res.40 = (Thr, Leu or Ser); Xaa at res.44 = (Ile, Val or Thr); Xaa at res.45 = (Val, Leu or Ile); Xaa at res.46 = (Gln or Arg); Xaa at res.47 = (Thr, Ala or Ser); Xaa at res.48 = (Leu or Ile); Xaa at res.49 = (Val or Met); Xaa at res.50 = (His, Asn or Arg); Xaa at res.51 = (Phe, Leu, Asn, Ser, Ala or Val); Xaa at res.52 = (Ile, Met, Asn, Ala, Val or Leu); Xaa at res.53 = (Asn, Lys, Ala, Glu, Gly or Phe); Xaa at 25 res.54 = (Pro, Ser or Val); Xaa at res.55 = (Glu, Asp, Asn, Gly, Val or Lys); Xaa at res.56 = (Thr, Ala, Val, Lys, Asp, Tyr, Ser, Ala, Pro or His); Xaa at res.57 = (Val, Ala or Ile); Xaa at res.58 = (Pro or Asp); Xaa at res.59 = (Lys, Leu or Glu); Xaa at res.60 = (Pro or 30 Ala); Xaa at res.63 = (Ala or Val); Xaa at res.65 = (Thr, Ala or Glu); Xaa at res.66 = (Gln, Lys, Arg or Glu); Xaa at res.67 = (Leu, Met or Val); Xaa at res.68 = (Asn, Ser, Asp or Gly); Xaa at res.69 = (Ala, Pro or Ser); Xaa at res.70 = (Ile, Thr, Val or Leu); Xaa at 35 res.71 = (Ser, Ala or Pro); Xaa at res.72 = (Val, Met

or Ile); Xaa at res.74 = (Tyr or Phe); Xaa at res.75 = (Phe, Tyr, Leu or His); Xaa at res.76 = (Asp, Asn or Leu); Xaa at res.77 = (Asp, Glu, Asn or Ser); Xaa at res.78 = (Ser, Gln, Asn, Tyr or Asp); Xaa at res.79 = (Ser, Asn, Asp, Glu or Lys); Xaa at res.80 = (Asn, Thr or Lys); Xaa at res.82 = (Ile, Val or Asn); Xaa at res.84 = (Lys or Arg); Xaa at res.85 = (Lys, Asn, Gln, His or Val); Xaa at res.86 = (Tyr or His); Xaa at res.87 = (Arg, Gln, Glu or Pro); Xaa at res.88 = (Asn, Glu or Asp); Xaa at res.90 = (Val, Thr, Ala or Ile); Xaa at res.92 = (Arg, Lys, Val, Asp or Glu); Xaa at res.93 = (Ala, Gly, Glu or Ser); Xaa at res.95 = (Gly or Ala) and Xaa at res.97 = (His or Arg).

15 <u>Generic Sequence 6</u>

Cys Xaa Xaa Xaa Xaa Leu Xaa Xaa Phe 5 1 Xaa Xaa Xaa Gly Trp Xaa Xaa Trp Xaa 15 20 Xaa Xaa Pro Xaa Xaa Xaa Xaa Ala 25 20 Xaa Tyr Cys Xaa Gly Xaa Cys Xaa 30 Xaa Pro Xaa Xaa Xaa Xaa Xaa 25 40 Xaa Xaa Xaa Asn His Ala Xaa Xaa 50 Xaa Xaa Xaa Xaa Xaa Xaa Xaa 55 30

Xaa Xaa Xaa Xaa Xaa Xaa Cys 60

Cys Xaa Pro Xaa Xaa Xaa Xaa Xaa 70

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Z,

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Xaa Xaa Xaa Leu Xaa Xaa Xaa 80 Xaa Xaa Xaa Xaa Val Xaa Leu Xaa 85 Xaa Xaa Xaa Xaa Met Xaa Val Xaa 95 90 Xaa Cys Xaa Cys Xaa

10 wherein each Xaa is independently selected from a group of one or more specified amino acids as defined by the following: "Res." means "residue" and Xaa at res.2 = (Lys, Arg, Ala or Gln); Xaa at res.3 = (Lys, Arg or Met); Xaa at res.4 = (His, Arg or Gln); Xaa at res.5 = 15 (Glu, Ser, His, Gly, Arg, Pro, Thr, or Tyr); Xaa at res.7 = (Tyr or Lys); Xaa at res.8 = (Val or Ile); Xaa at res.9 = (Ser, Asp or Glu); Xaa at res.11 = (Arg, Gln, Ser, Lys or Ala); Xaa at res.12 = (Asp, Glu, or Lys); Xaa at res.13 = (Leu, Val or Ile); Xaa at res.16 20 = (Gln, Leu, Asp, His, Asn or Ser); Xaa at res.17 = (Asp, Arg, Asn or Glu); Xaa at res.19 = (Ile or Val); Xaa at res.20 = (Ile or Val); Xaa at res.21 = (Ala or Ser); Xaa at res.23 = (Glu, Gln, Leu, Lys, Pro or Arg); Xaa at res.24 = (Gly or Ser); Xaa at res.25 = (Tyr or 25 Phe); Xaa at res.26 = (Ala, Ser, Asp, Met, His, Gln, Leu, or Gly); Xaa at res.28 = (Tyr, Asn or Phe); Xaa at res.31 = (Glu, His, Tyr, Asp, Gln or Ser); Xaa at res.33 = Glu, Lys, Asp, Gln or Ala); Xaa at res.35 = (Ala, Ser, Pro, Gln or Asn); Xaa at res.36 = (Phe, Leu 30 or Tyr); Xaa at res.38 = (Leu, Val or Met); Xaa at res.39 = (Asn, Asp, Ala, Thr or Pro); Xaa at res.40 = (Ser, Asp, Glu, Leu, Ala or Lys); Xaa at res.41 = (Tyr, Cys, His, Ser or Ile); Xaa at res.42 = (Met, Phe, Gly or Leu); Xaa at res.43 = (Asn, Ser or Lys); Xaa at 35 res.44 = (Ala, Ser, Gly or Pro); Xaa at res.45 = (Thr,

Leu or Ser); Xaa at res.49 = (Ile, Val or Thr); Xaa at res.50 = (Val, Leu or Ile); Xaa at res.51 = (Gln or Arg); Xaa at res.52 = (Thr, Ala or Ser); Xaa at res.53 = (Leu or Ile); Xaa at res.54 = (Val or Met); Xaa at 5 res.55 = (His, Asn or Arg); Xaa at res.56 = (Phe, Leu, Asn, Ser, Ala or Val); Xaa at res.57 = (Ile, Met, Asn, Ala, Val or Leu); Xaa at res.58 = (Asn, Lys, Ala, Glu, Gly or Phe); Xaa at res.59 = (Pro, Ser or Val); Xaa at res.60 = (Glu, Asp, Gly, Val or Lys); Xaa at res.61 = 10 (Thr, Ala, Val, Lys, Asp, Tyr, Ser, Ala, Pro or His); Xaa at res.62 = (Val, Ala or Ile); Xaa at res.63 = (Pro or Asp); Xaa at res.64 = (Lys, Leu or Glu); Xaa at res.65 = (Pro or Ala); Xaa at res.68 = (Ala or Val); Xaa at res.70 = (Thr, Ala or Glu); Xaa at res.71 = 15 (Gln, Lys, Arg or Glu); Xaa at res.72 = (Leu, Met or Val); Xaa at res.73 = (Asn, Ser, Asp or Gly); Xaa at res.74 = (Ala, Pro or Ser); Xaa at res.75 = (Ile, Thr, Val or Leu); Xaa at res.76 = (Ser, Ala or Pro); Xaa at res.77 = (Val, Met or Ile); Xaa at res.79 = (Tyr or 20 Phe); Xaa at res.80 = (Phe, Tyr, Leu or His); Xaa at res.81 = (Asp, Asn or Leu); Xaa at res.82 = (Asp, Glu, Asn or Ser); Xaa at res.83 = (Ser, Gln, Asn, Tyr or Asp); Xaa at res.84 = (Ser, Asn, Asp, Glu or Lys); Xaa at res.85 = (Asn, Thr or Lys); Xaa at res.87 = (Ile, 25 Val or Asn); Xaa at res.89 = (Lys or Arg); Xaa at res.90 = (Lys, Asn, Gln, His or Val); Xaa at res.91 = (Tyr or His); Xaa at res.92 = (Arg, Gln, Glu or Pro); Xaa at res.93 = (Asn, Glu or Asp); Xaa at res.95 = (Val, Thr, Ala or Ile); Xaa at res.97 = (Arg, Lys, Val, 30 Asp or Glu); Xaa at res.98 = (Ala, Gly, Glu or Ser); Xaa at res.100 = (Gly or Ala); and Xaa at res.102 = (His or Arg).

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Particularly useful sequences for use as morphogens in this invention include the C-terminal domains, e.g., the C-terminal 96-102 amino acid residues of Vgl, Vgr-1, DPP, OP-1, OP-2, CBMP-2A, CBMP-2B, GDF-1 (see 5 Table II, below, and Seq. ID Nos. 5-14), as well as proteins comprising the C-terminal domains of 60A, BMP3, BMP5 and BMP6 (see Seq. ID Nos. 24-28), all of which include at least the conserved six or seven cysteine skeleton. In addition, biosynthetic 10 constructs designed from the generic sequences, such as COP-1, 3-5, 7, 16, disclosed in U.S. Pat. No. 5,011,691, also are useful. Other sequences include the inhibins/activin proteins (see, for example, U.S. Pat. Nos. 4,968,590 and 5,011,691). Accordingly, other 15 useful sequences are those sharing at least 70% amino acid sequence homology or "similarity", and preferably 80% homology or similarity with any of the sequences above. These are anticipated to include allelic and species variants and mutants, and biosynthetic muteins, 20 as well as novel members of this morphogenic family of proteins. Particularly envisioned in the family of related proteins are those proteins exhibiting morphogenic activity and wherein the amino acid changes from the preferred sequences include conservative 25 changes, e.g., those as defined by Dayoff et al., Atlas of Protein Sequence and Structure; vol. 5, Suppl. 3, pp. 345-362, (M.O. Dayoff, ed., Nat'l BioMed. Research Fdn., Washington, D.C. 1979). As used herein, potentially useful sequences are aligned with a known 30 morphogen sequence using the method of Needleman et al. ((1970) <u>J.Mol.Biol.</u> <u>48</u>:443-453) and identities calculated by the Align program (DNAstar, Inc.). "Homology" or "similarity" as used herein includes allowed conservative changes as defined by Dayoff et 35 al.

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The currently most preferred protein sequences useful as morphogens in this invention include those having greater than 60% identity, preferably greater than 65% identity, with the amino acid sequence defining the conserved six cysteine skeleton of hOP1 (e.g., residues 43-139 of Seq. ID No. 5). These most preferred sequences include both allelic and species variants of the OP-1 and OP-2 proteins, including the Drosophila 60A protein. Accordingly, in another preferred aspect of the invention, useful morphogens include active proteins comprising species of polypeptide chains having the generic amino acid sequence herein referred to as "OPX", which accommodates the homologies between the various identified species of OP1 and OP2 (Seq. ID No. 29).

The morphogens useful in the methods, composition and devices of this invention include proteins comprising any of the polypeptide chains described 20 above, whether isolated from naturally-occurring sources, or produced by recombinant DNA or other synthetic techniques, and includes allelic and species variants of these proteins, naturally-occurring or biosynthetic mutants thereof, as well as various 25 truncated and fusion constructs. Deletion or addition mutants also are envisioned to be active, including those which may alter the conserved C-terminal cysteine skeleton, provided that the alteration does not functionally disrupt the relationship of these 30 cysteines in the folded structure. Accordingly, such active forms are considered the equivalent of the specifically described constructs disclosed herein.

The proteins may include forms having varying glycosylation patterns, varying N-termini, a family of related proteins having regions of amino acid sequence homology, and active truncated or mutated forms of

5 native or biosynthetic proteins, produced by expression of recombinant DNA in host cells.

The morphogenic proteins can be expressed from intact or truncated cDNA or from synthetic DNAs in procaryotic or eucaryotic host cells, and purified, cleaved, refolded, and dimerized to form morphogenically active compositions. Currently preferred host cells include E. coli or mammalian cells, such as CHO, COS or BSC cells. A detailed description of the morphogens useful in the methods, compositions and devices of this invention is disclosed in copending US patent application Serial Nos. 752,764, filed August 30, 1991, and 667,274, filed March 11, 1991, the disclosure of which are incorporated herein by reference.

Thus, in view of this disclosure, skilled genetic engineers can isolate genes from cDNA or genomic libraries of various different species which encode appropriate amino acid sequences, or construct DNAs from oligonucleotides, and then can express them in various types of host cells, including both procaryotes and eucaryotes, to produce large quantities of active proteins capable of protecting tissues and organs from immune cell-mediated tissue destruction, including substantially inhibiting such damage and/or regenerating the damaged tissue in a variety of mammals, including humans.

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The foregoing and other objects, features and advantages of the present invention will be made more apparent from the following detailed description of the invention.

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Brief Description of the Drawings

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- FIG 1 shows the cardioprotective effects of morphogen (hOP1) in a rat myocardial ischemia-reperfusion model, as evidenced by the smaller loss of myocardial creatine kinase in hOP1-treated rats;
- shows the effects of 20 µg of morphogen (hOP1)

 given 24 hours prior to isolation of rat heart

 on endothelial-dependent vasorelaxation to

 acetycholine following induced ischemiareperfusion injury;
- 15 FIG 3 shows the effect of morphogen (hOP1) on neutrophil adherence to LTB₄-stimulated mesenteric artery endothelium in neutrophilactivated rats;
- 20 FIG 4 (A and B) are schematic representations of morphogen inhibition of early mononuclear phagocytic multinuclearization in vivo;
- FIG 5 graphs the effect of a morphogen (e.g., OP-1)
 and a placebo control on mucositic lesion
 formation; and
- FIG 6 (A-D) graphs the effects of a morphogen (eg., OP-1, Figs. 6A and 6C) and TGF-β (Fig. 6B and 6D) on collagen (6A and 6B) and hyaluronic acid (6C and 6D) production in primary fibroblast cultures.

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Detailed Description of the Invention

It now has been surprisingly discovered that the morphogens defined herein are effective agents in alleviating the tissue destructive effects associated with the body's inflammatory response to tissue injury. In particular, as disclosed herein, the morphogens are capable of alleviating the necrotic tissue effects associated with the ensuing inflammatory responses that occur following an initial tissue injury.

When tissue injury occurs, whether caused by bacteria, trauma, chemicals, heat, or any other phenomenon, the body's inflammatory response is 15 stimulated. In response to signals released from the damaged cells (e.g., cytokines), extravascularization of immune effector cells is induced. Under ordinary circumstances these invading immune effector cells kill the infectious agent and/or infected or damaged cells 20 (through the release of killing substances such as superoxides, perforins, and other antimicrobial agents stored in granules), remove the dead tissues and organisms (through phagocytosis), release various biological response modifiers that promote rapid 25 healing and covering of the wound (quite often resulting in the formation of fibrotic scar tissue), and then, after the area is successfully healed, exit from the site of the initial insult. Once the site is perceived to be normal, the local release of 30 inflammatory cytokines ceases and the display of adhesion molecules on the vessel endothelium returns to basal levels. In some cases, however, the zeal of these interacting signals and cellular systems, which are designed to capture and contain very rapidly 35 multiplying infectious agents, act to the detriment of

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the body, killing additional, otherwise healthy, surrounding tissue. This additional unnecessary tissue death further compromises organ function and sometimes results in death of the individual. In addition, the resulting scar tissue that often forms can interfere with normal tissue function as occurs, for example, in idiopathic pulmonary fibrosis, IBD and organ cirrhosis.

The vascular endothelium constitutes the first 10 barrier between circulating immune effector cells and extravascular tissues. Extravasation of these circulating cells requires that they bind to the vascular endothelial cells, cross the basement membrane, and enter insulted tissues e.g, by 15 phagocytosis or protease-mediated extracellular matrix degradation. Without being limited to a particular theory, it is believed that the morphogens of this invention may modulate the inflammatory response in part by modulating the attachment of immune effector cells to the luminal side of the endothelium of blood vessels at or near sites of tissue damage and/or inflammatory lesions. Because the method reduces or prevents the attachment of immune effector cells at these sites, it also prevents the subsequent release of 25 tissue destructive agents by these same immune effector cells at sites of tissue damage and/or inflammatory lesions. Because attachment of immune effector cells to the endothelium must precede their extravascularization, the method also prevents the 30 initial or continued entry of these cells into extravascular sites of tissue destruction or ongoing inflammatory lesions. Therefore, the invention not only relates to a method to reduce or prevent the immune cell-mediated cellular destruction at 35 extravascular sites of recent tissue destruction, but

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also relates to a method to prevent or reduce the continued entry of immune effector cells into extravascular sites of ongoing inflammatory cascades. As will be appreciated by those skilled in the art, the 5 morphogens of this invention also may be contemplated in mechanisms for disrupting the functional interaction of immune effector cells with endothelium where the adhesion molecules are induced by means other than in response to tissue injury.

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One source of tissue injury is induced by cell exposure to toxic oxygen concentrations, such as ischemic-reperfusion tissue injury (oxygen deprivation), and following hyperoxia injury (lethally 15 high oxygen concentrations). Accordingly, the process of the present invention provides a method for alleviating the tissue damage induced by ischemicreperfusion injury or hyperoxia-induced injury comprising the step of administering to the afflicted 20 individual a therapeutic amount of a morphogen prior to, during, or after damage to the affected tissue. Where the toxic oxygen concentrations may be deliberately induced, as by a surgical or clinical procedure, the morphogen preferably is administered 25 prior to induction.

In addition, the morphogens described herein, in contrast to fibrogenic growth factors such as $TGF-\beta$, stimulate tissue morphogenesis and do not stimulate 30 fibrosis or scar tissue formation (see Example 9, below.) Accordingly, in addition to inhibiting the tissue destructive effects associated with the inflammatory response, the morphogens further enhance the viability of damaged tissue and/or organs by 35 stimulating the regeneration of the damaged tissue and preventing fibrogenesis.

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The morphogens described herein also can inhibit epithelial cell proliferation (see Example 10, below.) This activity of the morphogens also may be particularly useful in the treatment of psoriasis and other inflammatory diseases that involve epithelial cell populations.

Provided below are detailed descriptions of suitable morphogens useful in the methods and compositions of this invention, as well as methods for their administration and application, and numerous, nonlimiting examples which 1) illustrate the suitability of the morphogens and morphogen-stimulating agents described herein as therapeutic agents for protecting tissue from the tissue destructive effects associated with the body's inflammatory response; and 2) provide assays with which to test candidate morphogens and morphogen-stimulating agents for their efficacy.

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I. Useful Morphogens

As defined herein a protein is morphogenic if it is capable of inducing the developmental cascade of cellular and molecular events that culminate in the formation of new, organ-specific tissue and comprises at least the conserved C-terminal six cysteine skeleton or its functional equivalent (see supra).

30 Specifically, the morphogens generally are capable of all of the following biological functions in a morphogenically permissive environment: stimulating proliferation of progenitor cells; stimulating the differentiation of progenitor cells; stimulating the proliferation of differentiated cells; and supporting

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the growth and maintenance of differentiated cells, including the "redifferentiation" of transformed cells. Details of how the morphogens useful in the method of this invention first were identified, as well as a description on how to make, use and test them for morphogenic activity are disclosed in USSN 667,274, filed March 11, 1991 and USSN 752,764, filed August 30, 1991, the disclosures of which are hereinabove incorporated by reference. As disclosed therein, the morphogens may be purified from naturally-sourced material or recombinantly produced from procaryotic or eucaryotic host cells, using the genetic sequences disclosed therein. Alternatively, novel morphogenic sequences may be identified following the procedures disclosed therein.

Particularly useful proteins include those which comprise the naturally derived sequences disclosed in Table II. Other useful sequences include biosynthetic constructs such as those disclosed in U.S. Pat. 5,011,691, the disclosure of which is incorporated herein by reference (e.g., COP-1, COP-3, COP-4, COP-5, COP-7, and COP-16).

25 Accordingly, the morphogens useful in the methods and compositions of this invention also may be described by morphogenically active proteins having amino acid sequences sharing 70% or, preferably, 80% homology (similarity) with any of the sequences described above, where "homology" is as defined herein above.

The morphogens useful in the method of this invention also can be described by any of the 6 generic sequences described herein (Generic Sequences 1, 2, 3, 4, 5 and 6). Generic sequences 1 and 2 also may include, at their N-terminus, the sequence

Cys Xaa Xaa Xaa Xaa (Seq. ID No. 15)

Table II, set forth below, compares the amino acid sequences of the active regions of native proteins that 10 have been identified as morphogens, including human OP-1 (hOP-1, Seq. ID Nos. 5 and 16-17), mouse OP-1 (mOP-1, Seq. ID Nos. 6 and 18-19), human and mouse OP-2 15 (Seq. ID Nos. 7, 8, and 20-23), CBMP2A (Seq. ID No. 9), CBMP2B (Seq. ID No. 10), BMP3 (Seq. ID No. 26), DPP (from Drosophila, Seq. ID No. 11), Vgl, (from Xenopus, Seq. ID No. 12), Vgr-1 (from mouse, Seq. ID No. 13), GDF-1 (from mouse, Seq. ID Nos. 14, 32 and 33), 60A 20 protein (from Drosophila, Seq. ID Nos. 24 and 25), BMP5 (Seq. ID No. 27) and BMP6 (Seq. ID No. 28). The sequences are aligned essentially following the method of Needleman et al. (1970) J. Mol. Biol., 48:443-453, calculated using the Align Program (DNAstar, Inc.) 25 the table, three dots indicates that the amino acid in that position is the same as the amino acid in hOP-1. Three dashes indicates that no amino acid is present in that position, and are included for purposes of illustrating homologies. For example, amino acid 30 residue 60 of CBMP-2A and CBMP-2B is "missing". Of course, both these amino acid sequences in this region comprise Asn-Ser (residues 58, 59), with CBMP-2A then comprising Lys and Ile, whereas CBMP-2B comprises Ser and Ile.

TABLE II

			Lys	Lys	His	Glu	Leu	Tyr	Val		ક
	hOP-1	Cys	•	•••		•••	• • •	***	• • •		, *
	mOP-1	• • •	4 + + Amore	AIG	•••	•••	•••	* • •	* * *		.
5	h0F-2	•••	Arg	Arg	• • •	• • •	•,••	•••	•••		
	mOP-2	• • •	Arg	Arg	•••	Ser		• • •	• • •		
	DPP	• • •	Arg	Lys	Arg	His	•••	***			
	Vgl		•••		•••	Gly	•••	• • •	. •••		
	Vgr-1	• • •	• • •	Arg	• • •	Pro	•••	•••	• • •		
10	CBHP-2A	• • •	* * *	Arg		Ser	• • •	•••	• • •		
	CBMP-2B	• • •	Arg	Arg	Arg	Tyr		Lys	• • •		•
	BMP3	•••	Ala	Ala	Arg	Arg		•••	• • •		
	GDF-1	•••	Arg	Met	Glu	Thr		•••	• • •		
	60A	• • •	Gln				•••	• • •	• • •		
15	BMP5	. • • •					•. • •	•••	• • •	•	
	BMP6	• • •	Arg	•••		5					
		1		•							
									6 15	Asp	
		0	Phe	Arg	Asp	Leu	Gly	Tr		-	
20	hOP-1	Ser	-	•				• •		•••	
	mOP-1	•••				•••	• • •			• • •	
	h0P-2	•••		•			• • •	• • •			
	mOP-2	Se		ČO		Val	•••	• ••		Asn	
	DPP	As		` T++		Val	٠٠.				
25	Vgl	G1		C1.	•	77.4				•••	
	Vgr-1	••		°0°		. Va	1	• •	Asn	• • •	
	CBHP-2A	As	_	 Co		. Va	1	•	Asn	Glu	
	CBHP-2B	As		A 7.		; Il	.e •	•	Ser		
	BHP3				~3	_	1 .	•	His	Arg	•
30	GDF-1		•	14	•			••	His		æ,
	40a	A	sp ·					•		•••	
	BMP5	•	••	C.	_,		•	••	· · · · · · ·	•••	غي
	BMP6	•	••	• • •					15		
				10							

						Pro	Glu	Gly	Tyr	Ala
	h0P-1	Trp	Ile	Ile				•••	•••	•••
-	mOP-1	• • •	•••	•••	•••	• • •	Gln	•••	•••	Ser
	hOP-2	• • •	Val	• • •	•••	• • •	Gln		•••	Ser
	mOP-2	• • •	Val	• • •	• • •	•••	•	• • •	•••	Asp
5	DPP	•••	• • •	Val	• • •	• • •	Leu	•••	•••	Het
	♥ gl	• • •	Val	• • •	•••	•••	Gln	• • •	•••	
	Vgr-1	• • •	• • •	•••	• • •	• • •	Lys	•••	•••	His
	CBMP-2A	•••	•••	Val	•••	• • •	Pro	•••	• • •	Gln
	CBMP-2B	• • •	•••	Val	•••	•••	Pro	···	Phe	Asp
10	вирз	• • •	• • •	•••	Ser	• • •	Lys	Ser	Phe	Leu
••	GDF-1	•••	Val	•••	• • •	• • •	Arg	• • •		Gly
	60A	•••	•••	• • •	•••	•••	•••	***	• • •	•••
	BMP5		• • •	•••	•••	•••	•••	• • •	•••	
	BMP6		•••	•••	•••	•••	Lys	• • •	•••	• • •
15	2			20	. •				25	
13										
							•		~	Ala
	hOP-1	Ala	Tyr	Tyr	Cys	Glu	Gly	Glu	Cys	
	mOP-1	•••	• • •		•••	•••	•••	•••	•••	
20	hOP-2	•••		•••	•••	•••	• • •	• • •	• • •	Ser
20	mOP-2	•••			•••	• • •	• • •	•••	• • •	 D
	DPP			•••	•••	His	•••	Lys	• • •	Pro
		•••	Asn	•••	•••	Tyr	•••	• • •	•••	Pro
	Vgl		Asn	•••		Asp	•••	• • •	***	Ser
	Vgr-1 CBMP-2A	•••	Phe	•••		His		Glu	• • •	Pro
25		•••	Phe	•••		His	• • •	Asp	•••	Pro
	CBMP-2B	•••	•••			Ser	•••	Ala		Gln
	BMP3	•••	Asn	•••	•••	Gln		Gln	• • •	
	GDF-1		Phe		•••	Ser	•••	•••	• • •	Asn
	· 60A	• • •	Pho			Asp	• • • •		•••	Ser
30	BMP5	• • •	Asn			Asp				Ser
	BMP6	• • •	, 201		30					35
		71 1.	. D=/	Lev	ı Asr	ı Se	r Ty	r Net	t Ası	1 Ala
	hOP-1	Phe		ا عالم			_			
35	mOP-1	- ••	• •••	, •••	,					

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										•			
					Asp		Cys	•••	•••				
	h0P-2		•••	•••			Cys	•••	• • •	•••		_	
	mOP-2		•••	• • •	Asp	Asp	His	Phe	• • •	Ser		ė.	
	DPP		•••	•••	Ala	Glu	Ile	Leu	• • •	Gly			
	Vgl	Tyr	•••	• • •	Thr	Ala	His	•••		•••		7	
_	Vgr-1	- • • •	•••	•••	• • •		His	Leu		Ser			
5	CBHP-2A		•••	• • •	Ala	Asp	His	Leu		Ser			
	CBMP-2B	•••		•••	Ala	Asp	Ser	Gly	Ser*				
		Leu		Val	Ala	Leu		Leu	Lys	Pro			
	GDF-1		•••	Met	Pro	Lys	Ser		•••	•••			
	BMP3	•••		• • •	• • •	Ala	His		•••	• • •			
10	A00			• • •	•••	Ala	His	Het					
	BHP5	• • •	•••		•••	Ala	His	Het		• .		•	
	BHP6	•••	,			40	·						
									mt	Leu			
			Asn	His	Ala	Ile	Val	Gln	Thr				
15	hOP-1	Thr		•••		•••	• • •	•••	•••	• • •			
	mOP-1	•••	***	•••			Leu	•••	Ser	•••			
	hop-2	• • •	***			• • •	Leu	•••	Ser	.9. • •	•		
	mOP-2	• • •	•••	•••	•••	Va]	٠						
	DPP	•••	•••	•••			. Let	L •••	• • •	• • •			
20	Vgl	Ser	•••	• • •						•••			
20	Vgr-1			• • •	. • • •								
	CBMP-2A									•••			
	CBMP-2B	• • •			• •••	Th			. Se	r Ile			
	BMP3	Se	· · ·			. Va	_	•	g Al	a •••			
05	GDF-1	Le	u		• ••	-		_	_				
25	60A	••			• ••	• • •							
	BHP5	••				• •	• • • •	•	_				
					••	•	• •	. • · 50	,				
	BHP6		5				;	ου					
		·	-									3	
30								~	1., T	hr Val		v	
		17	al H	is P	he I	le A	lsn P						
	hor-1						•••	• •		 la		<u>ş</u>	
	mOP-1		, n			let !	Lys	* -					
	hoP-2	•:		_			Lys	1	Asp V	al ···			
35	mOP-2	•		lis I					•				

10	DPF Vg1 Vgr-1 CBMP-2A CBMP-2B BMP3 GDF-1 60A BMP5 BMP6			Ser Val Ser	Het Val Val	Glu ·	Ser -	A I I S Pro Gly Lys Asp	sp II yr . ys I Ger I Gly I Ala A Lys	le le le le la
15 20	hOP-1 mOP-1 hOP-2 mOP-2 DPP Vgl Vgr-1	Pro	Lys	Pro Ala Ala Ala Ala	Cys	Cys	Ala Val Val Val	Pro	Thr	Gln Lys Lys Lys Glu Glu
25	CBMP-2A CBMP-2B BMP3 GDF-1 60A BMP5 BMP6	Asp	Glu Leu	•••		•••	Val Val Val 	•••	Glu Ala 70	Lys Arg Arg Lys Lys
30 35	hOP-1 mOP-1 hOP-2 mOP-2 Vg1 Vgr-1	He	. Se . Se et Se	 ar	. Th	 r	val	•••	Tyr Phe	Phe Tyr Tyr Tyr

5	DPP CBMP-2A CBMP-2B BMP3 GDF-1 60A BMP5 BMP6	• • •	F	Ser Pro	Leu		Het Het Het Ile		•••	Leu Leu Tyr His		8) **	
10			4 ==	Ser	Ser	Asn	Val	Ile	Leu	Lys			
	hOP-1	Asp	Asp		•••	•••	• • •		• • •				
	mOP-1	• • •	Ser		Asn	• •, •	• • •	•••	• • •	Arg Arg			
	hop-2	• • •	Ser	•••	Asn	•••	•••	•••	•••				
	mOP-2 DPP	Asn	* • •	Gln	• • •	Thr	•••	Val Val	•••	Arg			
15	Vgl	•••	Asn	Asn	Asp	• • •	•••	••••					
	Vgr-1	•••	•••	Asn	 77	Lys	•••	Val	•••	•••			
	CBHP-2A	•••	Glu	Asn	Glu Asp	Lys	• • •	Val	• • •	•••			
	CBMP-2B	•••	Glu	Tyr Asn	Lys	-,-	• • •	Val	•••	•••			
20	BMP3		Glu		Asp	• • •	• • •	Val	•••	Arg			
	GDF-1	Ten	Asn Asn	Asp	Glu		•••	Asn	• * •				
	60A	Leu			•••	• • •	•••	• • •	•••	•••	•		
	BHP5 BHP6	•••		Asn		•••			***			• .	•
05						85							
25													
		Lys	; Tyr	Arg	Asn	Het	•	l Va					
	hor-1		-			• •		• ••	. Ly:				
	mOP-1	•••	. 114 6			. ••	• ••		Tee		•		
30	h0P-2	••	111				• ••		17.0			5	
	mOP-2	As		. GL	n Glu	ı						a	
	DPP Vgl	Hi		. Gl	u		. Al					ŝ	•
	vgr Vgr-1	• •					• • •	, -	ر م				
35	CBHP-2A	As	in	. Gl	n As	р	• • •	•					

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5	CBMP-2B BMP3 GDF-1 60A BMP5 BMP6	Val Gln	•••	Gln Pro Glu 	Glu Asp Trp	•••	Thr Ile	•••	Glu Glu Asp Lys
		90							
10	hOP-1	Ala	Cys	Gly	Cys	His			
•	mOP-1	•••	• • •	•••	•••	* • •			
	h0P-2	•••	• • •	• • •	•••	• • •			
	mOP-2	• • •	•••	•••	• • •				
	DPP	Gly	• • •	•••	• • •	Arg			
15	Vgl	Glu	• • •	•••	• • •	Arg			
	Vgr-1	•••	• • •	• • •	•••				
	CBMP-2A	Gly	• • •		• • •	Arg			
	CBMP-2B	Gly	• • •	• • •	•••	Arg			
	BHP3	Ser	•••	Ala	•••	Arg			
20	GDF-1	Glu	• • •	• • •	•••	Arg			
	60A	Ser	• • •	• • •	• • •	•••			
	BMP5	Ser	•••	• • •	• • •	• • •			
	BMP6	• • •	• • •	•••	• • •	•••			
				100					•••
25	++Retveen	residue	s 56	and 57	of BME	?3 is	a Val	residu	76 }

25 **Between residues 56 and 57 of BMP3 is a Val residue; between residues 43 and 44 of GDF-1 lies the amino acid sequence Gly-Gly-Pro-Pro.

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30 As is apparent from the foregoing amino acid sequence comparisons, significant amino acid changes can be made within the generic sequences while retaining the morphogenic activity. For example, while the GDF-1 protein sequence depicted in Table II shares only about 50% amino acid identity with the hOP1

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sequence described therein, the GDF-1 sequence shares greater than 70% amino acid sequence homology (or "similarity") with the hOP1 sequence, where "homology" or "similarity" includes allowed conservative amino acid changes within the sequence as defined by Dayoff, et al., Atlas of Protein Sequence and Structure vol.5, supp.3, pp.345-362, (M.O. Dayoff, ed., Nat'l BioMed. Res. Fd'n, Washington D.C. 1979.)

The currently most preferred protein sequences useful as morphogens in this invention include those 10 having greater than 60% identity, preferably greater than 65% identity, with the amino acid sequence defining the conserved six cysteine skeleton of hOP1 15 (e.g., residues 43-139 of Seq. ID No. 5). These most preferred sequences include both allelic and species variants of the OP-1 and OP-2 proteins, including the Drosophila 60A protein. Accordingly, in still another preferred aspect, the invention includes morphogens 20 comprising species of polypeptide chains having the generic amino acid sequence referred to herein as "OPX", which defines the seven cysteine skeleton and accommodates the identities between the various identified mouse and human OP1 and OP2 proteins. OPX 25 is presented in Seq. ID No. 29. As described therein, each Xaa at a given position independently is selected from the residues occurring at the corresponding position in the C-terminal sequence of mouse or human OP1 or OP2 (see Seq. ID Nos. 5-8 and/or Seq. ID Nos. 30 16-23).

II. Formulations and Methods for Administering Therapeutic Agents

The morphogens may be provided to an individual by 5 any suitable means, preferably directly (e.g., locally, as by injection or topical administration to a tissue locus) or systemically (e.g., parenterally or orally). Where the morphogen is to be provided parenterally, such as by intravenous, subcutaneous, intramuscular, 10 intraorbital, ophthalmic, intraventricular, intracranial, intracapsular, intraspinal, intracisternal, intraperitoneal, buccal, rectal, vaginal, intranasal or by aerosol administration, the morphogen preferably comprises part of an aqueous 15 solution. The solution is physiologically acceptable so that in addition to delivery of the desired morphogen to the patient, the solution does not otherwise adversely affect the patient's electrolyte and volume balance. The aqueous medium for the 20 morphogen thus may comprise normal physiologic saline (9.85% NaCl, 0.15M), pH 7-7.4. The aqueous solution containing the morphogen can be made, for example, by dissolving the protein in 50% ethanol containing acetonitrile in 0.1% trifluoroacetic acid (TFA) or 0.1% 25 HCl, or equivalent solvents. One volume of the resultant solution then is added, for example, to ten volumes of phosphate buffered saline (PBS), which further may include 0.1-0.2% human serum albumin (HSA). The resultant solution preferably is vortexed extensively. If desired, a given morphogen may be made more soluble by association with a suitable molecule. For example, association of the mature dimer with the pro domain of the morphogen keeps the morphogen soluble in physiological buffers. In fact, the endogenous 35 protein is thought to be transported in this form.

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Another molecule capable of enhancing solubility and particularly useful for oral administrations, is casein. For example, addition of 0.2% casein increases solubility of the mature active form of OP-1 by 80%. 5 Other components found in milk and/or various serum proteins also may be useful.

Useful solutions for parenteral administration may be prepared by any of the methods well known in the 10 pharmaceutical art, described, for example, in Remington's Pharmaceutical Sciences (Gennaro, A., ed.), Mack Pub., 1990. Formulations may include, for example, polyalkylene glycols such as polyethylene glycol, oils of vegetable origin, hydrogenated 15 naphthalenes, and the like. Formulations for direct administration, in particular, may include glycerol and other compositions of high viscosity to help maintain the morphogen at the desired locus. Biocompatible, preferably bioresorbable, polymers, including, for 20 example, hyaluronic acid, collagen, tricalcium phosphate, polybutyrate, lactide and glycolide polymers, and lactide/glycolide copolymers, may be useful excipients to control the release of the morphogen in vivo. Other potentially useful parenteral 25 delivery systems for these morphogens include ethylenevinyl acetate copolymer particles, osmotic pumps, implantable infusion systems, and liposomes. Formulations for inhalation administration contain as excipients, for example, lactose, or may be aqueous 30 solutions containing, for example, polyoxyethylene-9lauryl ether, glycocholate and deoxycholate, or oily solutions for administration in the form of nasal drops, or as a gel to be applied intranasally.

Formulations for parenteral administration may also include glycocholate for buccal administration, methoxysalicylate for rectal administration, or cutric acid for vaginal administration.

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Suppositories for rectal administration also may be prepared by mixing the morphogen or morphogenstimulating agent with a non-irritating excipient such as cocoa butter or other compositions which are solid 10 at room temperature and liquid at body temperatures.

Formulations for topical administration to the skin surface may be prepared by dispersing the morphogen or morphogen-stimulating agent with a dermally acceptable 15 carrier such as a lotion, cream, ointment or soap. Particularly useful are carriers capable of forming a film or layer over the skin to localize application and inhibit removal. For topical administration to internal tissue surfaces, the morphogen may be 20 dispersed in a liquid tissue adhesive or other substance known to enhance adsorption to a tissue surface. For example, hydroxypropylcellulose or fibrinogen/thrombin solutions may be used to advantage. Alternatively, tissue-coating solutions, such as 25 pectin-containing formulations, may be used.

Alternatively, the morphogens described herein may be administered orally. Oral administration of proteins as therapeutics generally is not practiced as 30 most proteins are readily degraded by digestive enzymes and acids in the mammalian digestive system before they can be absorbed into the bloodstream. However, the morphogens described herein typically are acid stable and protease-resistant (see, for example, U.S. Pat.No. 35 4,968,590.) In addition, at least one morphogen, OP-1,

has been identified in mammary gland extract, colostrum and 57-day milk. Moreover, the OP-1 purified from mammary gland extract is morphogenically active. Specifically, this protein induces endochondral bone 5 formation in mammals when implanted subcutaneously in association with a suitable matrix material, using a standard in vivo bone assay, such as is disclosed in U.S. Pat.No. 4,968,590. Moreover, the morphogen also is detected in the bloodstream. Finally, soluble form 10 morphogen, e.g., mature morphogen associated with the pro domain, is morphogenically active. These findings indicate that oral and parenteral administration are viable means for administering morphogens to an individual. In addition, while the mature forms of 15 certain morphogens described herein typically are sparingly soluble, the morphogen form found in milk (and mammary gland extract and colostrum) is readily soluble, probably by association of the mature, morphogenically active form with part or all of the pro 20 domain of the intact sequence and/or by association with one or more milk components. Accordingly, the compounds provided herein also may be associated with molecules capable of enhancing their solubility in vitro or in vivo.

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Where the morphogen or morphogen-stimulating agent comprises part of a tissue or organ preservation solution, any commercially available preservation solution may be used to advantage. For example, useful 30 solutions known in the art include Collins solution, Wisconsin solution, Belzer solution, Eurocollins solution and lactated Ringer's solution. Generally, an organ preservation solution usually possesses one or more of the following properties: (a) an osmotic 35 pressure substantially equal to that of the inside of a

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mammalian cell, (solutions typically are hyperosmolar and have K+ and/or Mg++ ions present in an amount sufficient to produce an osmotic pressure slightly higher than the inside of a mammalian cell; (b) the 5 solution typically is capable of maintaining substantially normal ATP levels in the cells; and (c) the solution usually allows optimum maintenance of glucose metabolism in the cells. Organ preservation solutions also may contain anticoagulants, energy 10 sources such as glucose, fructose and other sugars, metabolites, heavy metal chelators, glycerol and other materials of high viscosity to enhance survival at low temperatures, free oxygen radical inhibiting agents and A detailed description of a pH indicator. 15 preservation solutions and useful components may be found, for example, in US Patent No. 5,002,965, the disclosure of which is incorporated herein by reference.

The compounds provided herein also may be associated with molecules capable of targeting the morphogen or morphogen-stimulating agent to the desired tissue. For example, an antibody, antibody fragment, or other binding protein that interacts specifically with a surface molecule on cells of the desired tissue, may be used. Useful targeting molecules may be designed, for example, using the single chain binding site technology disclosed, for example, in U.S. Pat. No. 5,091,513.

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As described above, the morphogens provided herein share significant sequence homology in the C-terminal active domains. By contrast, the sequences typically diverge significantly in the sequences which define the pro domain. Accordingly, the pro domain is thought to

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be morphogen-specific. As described above, it is also known that the various morphogens identified to date are differentially expressed in the different tissues. Accordingly, without being limited to any given theory, 5 it is likely that, under natural conditions in the body, selected morphogens typically act on a given tissue. Accordingly, part or all of the pro domains which have been identified associated with the active form of the morphogen in solution, may serve as 10 targeting molecules for the morphogens described herein. For example, the pro domains may interact specifically with one or more molecules at the target tissue to direct the morphogen associated with the pro domain to that tissue. Accordingly, another useful 15 targeting molecule for targeting morphogen to a tissue of interest is part or all of a morphogen pro domain. For example, part or all of the pro domain of GDF-1 may be used to target a morphogen to nerve tissue. Alternatively, part or all of the pro domain of OP-1 or 20 CBMP2 may be used to target a morphogen to bone tissue, both of which proteins are found naturally associated with bone tissue.

The morphogens described herein are useful for 25 providing neuroprotective effects to alleviate neural pathway damage associated with the body's immune/inflammatory response to an initial injury to nerve tissue. As used herein, a "neural pathway" describes a nerve circuit for the passage of electric 30 signals from a source to a target cell site and includes both the central nervous system (CNS) and peripheral nervous system (PNS). The pathway includes the neurons through which the electric impulse is transported, including groups of interconnecting 35 neurons, the nerve fibers formed by bundled neuronal

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axons, and the glial cells surrounding and associated with the neurons. An inflammatory response to nerve tissue injury may follow trauma to nerve tissue, caused, for example, by an autoimmune (including 5 autoantibody) dysfunction, neoplastic lesion, infection, chemical or mechanical trauma, or other disease. An exemplary nerve-related inflammatory disease is multiple sclerosis. Neural pathway damage also can result from a reduction or interruption, e.g., 10 occlusion, of a neural blood supply, as in an embolic stroke, (e.g, ischemia or hypoxia-induced injury), or by other trauma to the nerve or surrounding material. In addition, at least part of the damage associated with a number of primary brain tumors also appears to 15 be immunologically related. Application of the morphogen directly to the cells to be treated, or providing the morphogen to the mammal systemically, for example, intravenously or indirectly by oral administration, may be used to alleviate and/or inhibit 20 the immunologically related response to a neural injury. Alternatively, administration of an agent capable of stimulating morphogen expression and/or secretion in vivo, preferably at the site of injury, also may be used. Where the injury is to be induced, 25 as during surgery or other aggressive clinical treatment, the morphogen or agent may be provided prior to induction of the injury to provide a neuroprotective effect to the nerve tissue at risk.

Where the morphogen is intended for use as a therapeutic to alleviate tissue damage associated with an immune/inflammatory condition of the CNS, an additional problem must be addressed: overcoming the so-called "blood-brain barrier", the brain capillary wall structure that effectively screens out all but

selected categories of molecules present in the blood, preventing their passage into the brain. The blood-brain barrier may be bypassed effectively by direct infusion of the morphogen or morphogen-5 stimulating agent into the brain. Alternatively, the morphogen or morphogen-stimulating agent may be modified to enhance its transport across the blood-brain barrier. For example, truncated forms of the morphogen or a morphogen-stimulating agent may be 10 most successful. Alternatively, the morphogen or morphogen-stimulating agent may be modified to render it more lipophilic, or it may be conjugated to another molecule which is naturally transported across the barrier, using standard means known to those skilled in 15 the art, as, for example, described in Pardridge, Endocrine Reviews 7:314-330 (1986) and U.S. Pat. No. 4,801,575.

Finally, the morphogens or morphogen-stimulating
agents provided herein may be administered alone or in
combination with other molecules known to be beneficial
in the treatment compositions and methods described
herein, including, but not limited to anticoagulants,
free oxygen radical inhibiting agents, salicylic acid,
free oxygen radical inhibiting agents, provided the salicylic acid,
treatments also may include ultra-violet light
treatment, zinc oxide and retinoids.

The compounds provided herein can be formulated
into pharmaceutical compositions by admixture with
pharmaceutically acceptable nontoxic excipients and
carriers. As noted above, such compositions may be
prepared for parenteral administration, particularly in

the form of liquid solutions or suspensions; for oral administration, particularly in the form of tablets or capsules; or intranasally, particularly in the form of powders, nasal drops, or aerosols.

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The compositions can be formulated for parenteral or oral administration to humans or other mammals in therapeutically effective amounts, e.g., amounts which provide appropriate concentrations for a time 10 sufficient to alleivate the tissue destructive effects associated with the inflammatory response, including protecting tissue in anticipation of tissue damage.

As will be appreciated by those skilled in the art, 15 the concentration of the compounds described in a therapeutic composition will vary depending upon a number of factors, including the dosage of the drug to be administered, the chemical characteristics (e.g., hydrophobicity) of the compounds employed, and the 20 route of administration. The preferred dosage of drug to be administered also is likely to depend on such variables as the type and extent of progression of the tissue damage, the overall health status of the particular patient, the relative biological efficacy of 25 the compound selected, the formulation of the compound excipients, and its route of administration. In general terms, the compounds of this invention may be provided in an aqueous physiological buffer solution containing about 0.001% to 10% w/v compound for parenteral administration. Typical dose ranges are from about 10 ng/kg to about 1 g/kg of body weight per day; a preferred dose range is from about 0.1 μ g/kg to 100 mg/kg of body weight per day. Optimally, the morphogen dosage given is between 0.1-100 μg of protein 35 per kilogram weight of the patient. No obvious

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morphogen induced pathological lesions are induced when mature morphogen (e.g., OP-1, 20 μ g) is administered daily to normal growing rats for 21 consecutive days. Moreover, 10 μ g systemic injections of morphogen (e.g., 5 OP-1) injected daily for 10 days into normal newborn mice does not produce any gross abnormalities.

In administering morphogens systemically in the methods of the present invention, preferably a large 10 volume loading dose is used at the start of the treatment. The treatment then is continued with a maintenance dose. Further administration then can be determined by monitoring at intervals the levels of the morphogen in the blood.

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Where tissue injury is induced deliberately as part of, for example, a surgical procedure, the morphogen preferably is provided just prior to, or concomitant with induction of the trauma. Preferably, the morphogen 20 is administered prophylactically in a surgical setting.

Alternatively, an effective amount of an agent capable of stimulating endogenous morphogen levels may be administered by any of the routes described above. For example, an agent capable of stimulating morphogen 25 production and/or secretion from cells of affected tissue and/or transplant tissue may be provided to a mammal, e.g., by direct administration of the agent to the tissue to be treated. A method for identifying and testing agents capable of modulating the levels of 30 endogenous morphogens in a given tissue is described generally herein in Example 15, and in detail in copending USSN 752,859, filed August 30, 1991, the disclosure of which is incorporated herein by reference. Briefly, candidate compounds can be

identified and tested by incubating the compound $\underline{\text{in}}$ vitro with a test tissue or cells thereof, for a time sufficient to allow the compound to affect the production, i.e., the expression and/or secretion, of a 5 morphogen produced by the cells of that tissue.

For purposes of the present invention, the abovedescribed morphogens effective in alleviating tissue damage associated with ischemic-reperfusion injury (or 10 the agents that stimulate them, referred to collectively herein as "therapeutic agent") are administered prior to or during the restoration of oxygen (e.g., restoration of blood flow, reperfusion.) Where treatment is to follow an existing injury, the 15 therapeutic agent preferably is administered as an intravenous infusion provided acutely after the hypoxic or ischemic condition occurs. For example, the therapeutic agent can be administered by intravenous infusion immediately after a cerebral infarction, a 20 myocardial infarction, asphyxia, or a cardiopulmonary arrest. Where ischemia or hypoxia injury is deliberately and/or unavoidably induced as part of, for example, a surgical procedure where circulation to an organ or organ system is deliberately and/or 25 transiently interrupted, e.g., in carotid enterectomy, coronary artery bypass, grafting, organ transplanting, fibrinolytic therapy, etc., the therapeutic agent preferably is provided just prior to, or concomitant with, reduction of oxygen to the tissue. Preferably, 30 the morphogen is administered prophylactically in a surgical setting.

Similarly, where hyperoxia-induced injury already has occurred, the morphogen is administered upon 35 diagnosis. Where hyperoxia injury may be induced as,

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for example, during treatment of prematurely newborn babies, or patients suffering from pulmonary diseases such as emphysema, the therapeutic agent preferably is administered prior to administration of oxygen e.g., prophylactically.

III. Examples

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10 Example 1. Identification of Morphogen-Expressing Tissue

Determining the tissue distribution of morphogens may be used to identify different morphogens expressed 15 in a given tissue, as well as to identify new, related morphogens. Tissue distribution also may be used to identify useful morphogen-producing tissue for use in screening and identifying candidate morphogenstimulating agents. The morphogens (or their mRNA 20 transcripts) readily are identified in different tissues using standard methodologies and minor modifications thereof in tissues where expression may be low. For example, protein distribution may be determined using standard Western blot analysis or 25 immunofluorescent techniques, and antibodies specific to the morphogen or morphogens of interest. Similarly, the distribution of morphogen transcripts may be determined using standard Northern hybridization protocols and transcript-specific probes.

Any probe capable of hybridizing specifically to a transcript, and distinguishing the transcript of interest from other, related transcripts may be used. Because the morphogens described herein share such high sequence homology in their active, C-terminal domains,

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the tissue distribution of a specific morphogen transcript may best be determined using a probe specific for the pro region of the immature protein and/or the N-terminal region of the mature protein. 5 Another useful sequence is the 3' non-coding region flanking and immediately following the stop codon. These portions of the sequence vary substantially among the morphogens of this invention, and accordingly, are specific for each protein. For example, a particularly 10 useful Vgr-1-specific probe sequence is the PvuII-SacI fragment, a 265 bp fragment encoding both a portion of the untranslated pro region and the N-terminus of the mature sequence (see Lyons et al. (1989) PNAS 86:4554-4558 for a description of the cDNA sequence). 15 Similarly, particularly useful mOP-1-specific probe sequences are the BstX1-BglI fragment, a 0.68 Kb sequence that covers approximately two-thirds of the mOP-1 pro region; a StuI-StuI fragment, a 0.2 Kb sequence immediately upstream of the 7-cysteine domain; 20 and the Earl-Pstl fragment, an 0.3 Kb fragment containing a portion of the 3'untranslated sequence (See Seq. ID No. 18, where the pro region is defined essentially by residues 30-291.) Similar approaches may be used, for example, with hOP-1 (Seq. ID No. 16) 25 or human or mouse OP-2 (Seq. ID Nos. 20 and 22.)

Using these morphogen-specific probes, which may be synthetically engineered or obtained from cloned sequences, morphogen transcripts can be identified in mammalian tissue, using standard methodologies well known to those having ordinary skill in the art.

Briefly, total RNA is prepared from various adult murine tissues (e.g., liver, kidney, testis, heart, brain, thymus and stomach) by a standard methodology such as by the method of Chomczyaski et al. ((1987)

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Anal. Biochem 162:156-159) and described below. Poly (A)+ RNA is prepared by using oligo (dT)-cellulose chromatography (e.g., Type 7, from Pharmacia LKB Poly (A)+ RNA (generally 15 μg) Biotechnology, Inc.). 5 from each tissue is fractionated on a 1% agarose/formaldehyde gel and transferred onto a Nytran membrane (Schleicher & Schuell). Following the transfer, the membrane is baked at 80°C and the RNA is cross-linked under UV light (generally 30 seconds at 1 10 mW/cm²). Prior to hybridization, the appropriate probe is denatured by heating. The hybridization is carried out in a lucite cylinder rotating in a roller bottle apparatus at approximately 1 rev/min for approximately 15 hours at 37°C using a hybridization mix of 40% 15 formamide, 5 x Denhardts, 5 x SSPE, and 0.1% SDS. Following hybridization, the non-specific counts are washed off the filters in 0.1 x SSPE, 0.1% SDS at 50°C.

Examples demonstrating the tissue distribution of 20 various morphogens, including Vgr-1, OP-1, BMP2, BMP3, BMP4, BMP5, GDF-1, and OP-2 in developing and adult tissue are disclosed in co-pending USSN 752,764, and in Ozkaynak, et al., (1991) Biochem. Biophys. Res. Commn. 179:116-123, and Ozkaynak, et al. (1992) (JBC, in 25 press), the disclosures of which are incorporated herein by reference. Using the general probing methodology described herein, northern blot hybridizations using probes specific for these morphogens to probe brain, spleen, lung, heart, liver 30 and kidney tissue indicate that kidney-related tissue appears to be the primary expression source for OP-1, with brain, heart and lung tissues being secondary sources. OP-1 mRNA also was identified in salivary glands, specifically rat parotid glands, using this 35 probing methodology. Lung tissue appears to be the

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primary tissue expression source for Vgr-1, BMP5, BMP4 and BMP3. Lower levels of Vgr-1 also are seen in kidney and heart tissue, while the liver appears to be a secondary expression source for BMP5, and the spleen 5 appears to be a secondary expression source for BMP4. GDF-1 appears to be expressed primarily in brain tissue. To date, OP-2 appears to be expressed primarily in early embryonic tissue. Specifically, northern blots of murine embryos and 6-day post-natal 10 animals shows abundant OP2 expression in 8-day embryos. Expression is reduced significantly in 17-day embryos and is not detected in post-natal animals.

Active Morphogens in Body Fluids Example 2.

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OP-1 expression has been identified in saliva (specifically, the rat parotid gland, see Example 1), human blood serum, and various milk forms, including mammary gland extract, colostrum, and 57-day bovine 20 milk. Moreover, and as described in USSN 923,780, the disclosure of which is incorporated herein by reference, the body fluid-extracted protein is morphogenically active. The discovery that the morphogen naturally is present in milk and saliva, 25 together with the known observation that mature, active OP-1 is acid-stable and protease-resistant, indicate that oral administration is a useful route for therapeutic administration of morphogen to a mammal. Oral administration typically is the preferred mode of 30 delivery for extended or prophylactic therapies. addition, the identification of morphogen in all milk forms, including colostrum, suggests that the protein may play a significant role in tissue development, including skeletal development, of juveniles.

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2.1 Morphogen Detection in Milk

op-1 was partially purified from rat mammary gland extract and bovine colostrum and 57 day milk by passing these fluids over a series of chromatography columns:

(e.g., cation-exchange, affinity and reverse phase). At each step the eluant was collected in fractions and these were tested for the presence of OP-1 by standard immunoblot. Immunoreactive fractions then were combined and purified further. The final, partially purified product then was examined for the presence of OP-1 by Western blot analysis using OP-1-specific antisera, and tested for in vivo and in vitro activity.

15 OP-1 purified from the different milk sources were characterized by Western blotting using antibodies raised against OP-1 and BMP2. Antibodies were prepared using standard immunology protocols well known in the art, and as described generally in Example 15, below, using full-length <u>E. coli-produced OP-1</u> and BMP2 as the immunogens. In all cases, the purified OP-1 reacted only with the anti-OP-1 antibody, and not with anti-BMP2 antibody.

The morphogenic activity of OP-1 purified from mammary gland extract was evaluated in vivo essentially following the rat model assay described in U.S. Pat. No. 4,968,590, hereby incorporated by reference. Briefly, a sample was prepared from each OP-1 immunoreactive fraction of the mammary gland extract-derived OP-1 final product by lyophilizing a portion (33%) of the fraction and resuspending the protein in 220µl of 50% acetonitrile/0.1% TFA. After vortexing, 25 mg of collagen matrix was added. The samples were lyophilized overnight, and implanted in

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Long Evans rats (Charles River Laboratories,
Wilmington, MA, 28-35 days old). Each fraction was
implanted in duplicate. For details of the collagen
matrix implantation procedure, see, for example, U.S.
Pat. No. 4,968,590, hereby incorporated by reference.
After 12 days, the implants were removed and evaluated
for new bone formation by histological observation as
described in U.S. Patent No. 4,968,590. In all cases,
the immunoreactive fractions were osteogenically
active.

2.2 Morphogen Detection in Serum

Morphogen may be detected in serum using morphogen-15 specific antibodies. The assay may be performed using any standard immunoassay, such as Western blot (immunoblot) and the like. Preferably, the assay is performed using an affinity column to which the morphogen-specific antibody is bound and through which 20 the sample serum then is poured, to selectively extract the morphogen of interest. The morphogen then is A suitable elution buffer may be determined empirically by determining appropriate binding and elution conditions first with a control (e.g., 25 purified, recombinantly-produced morphogen.) Fractions then are tested for the presence of the morphogen by standard immunoblot, and the results confirmed by N-terminal sequencing. Preferably, the affinity column is prepared using monoclonal antibodies. Morphogen 30 concentrations in serum or other fluid samples then may be determined using standard protein quantification techniques, including by spectrophotometric absorbance or by quantitation of conjugated antibody.

Presented below is a sample protocol for identifying OP-1 in serum. Following this general methodology other morphogens may be detected in body fluids, including serum. The identification of 5 morphogen in serum further indicates that systemic administration is a suitable means for providing therapeutic concentrations of a morphogen to an individual, and that morphogens likely behave systemically as endocrine-like factors. Finally, using 10 this protocol, fluctuations in endogenous morphogen levels can be detected, and these altered levels may be used as an indicator of tissue dysfunction. Alternatively, fluctuations in morphogen levels may be assessed by monitoring morphogen transcription levels, 15 either by standard northern blot analysis as described in Example 1, or by in situ hybridization, using a labelled probe capable of hybridizing specifically to morphogen mRNA, and standard RNA hybridization protocols well described in the art and described 20 generally in Example 1.

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OP-1 was detected in human serum using the
following assay. A monoclonal antibody raised against
mammalian, recombinantly produced OP-1 using standard
immunology techniques well described in the art and
described generally in Example 15, was immobilized by
passing the antibody over an agarose-activated gel
(e.g., Affi-GelTM, from Bio-Rad Laboratories, Richmond,
(A, prepared following manufacturer's instructions) and
used to purify OP-1 from serum. Human serum then was
passed over the column and eluted with 3M
K-thiocyanate. K-thiocyanante fractions then were
dialyzed in 6M urea, 20mM PO₄, pH 7.0, applied to a C8
HPLC column, and eluted with a 20 minute, 25-50%
sectonitrile/0.1% TFA gradient. Mature, recombinantly

produced OP-1 homodimers elute between 20-22 minutes. Fractions then were collected and tested for the presence of OP-1 by standard immunoblot using an OP-1 specific antibody as for Example 2.A.

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Administered or endogenous morphogen levels may be monitored in the therapies described herein by comparing the quantity of morphogen present in a body fluid sample with a predetermined reference value, for 10 example, to evaluate the efficiency of a therapeutic protocol, and the like. In addition, fluctuations in the level of endogenous morphogen antibodies may be detected by this method, most likely in serum, using an antibody or other binding protein capable of 15 interacting specifically with the endogenous morphogen antibody. Detected fluctuations in the levels of the morphogen or endogenous antibody may be used, for example, as indicators of a change in tissue status. For example, as damaged tissue is regenerated and the 20 tissue or organ's function returns to "normal" and, in the absence of additional tissue damage, lower doses of morphogen may be required, and a higher level of circulating morphogen antibody may be measured.

Effect of Morphogen after the Onset of 25 Example 3. the Ischemic Process

The cardioprotective effect of morphogens following ischemic-reperfusion injury in a mammal can readily be 30 assessed in a rat model. In this example, morphogen (e.g., OP-1) is administered just prior to the onset of the ischemic process in experimentally-induced myocardial infracted rats, essentially following the method of Lefer, et al. (1990) Science 249:61-64 and 35 (1992) <u>J. Mol. Cell. Cardiol.</u> <u>24</u>: 385-393, the

disclosures of which are hereby incorporated by reference. Briefly, loss of myocardial tissue function following ischemia and reperfusion is assayed by measuring loss of myocardial creatine kinease activity 5 (CK) and loss of endothelium-dependent vasorelaxation function (see Example 4, below).

In a first group of ether-anesthetized rats, the left coronary artery was occluded just proximal to the 10 first main branch with a silk ligature to induce a myocardial infarction (MI). The ligature was removed 10 minutes after occlusion to allow for coronary reperfusion. This first group is referred to herein as the "myocardial infarcted" (MI) group. A second group 15 of rats underwent the same procedure except that the coronary artery was not occluded, and thus no myocardial infarction occurred. The second group of rats is referred to herein as the "sham myocardial infarcted group" (SHAM MI).

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The first group of rats, the MI group of rats, further was divided into three sup-groups. $2\mu g$ of morphogen (OP-1) were injected intravenously into the first sub-group of MI rats 10 minutes after ligature, 25 immediately before reperfusion; into the second subgroup of MI rats 20 μg of OP-1 were injected intravenously 10 minutes after ligature and immediately before reperfusion; and into the third sub-group of MI rats (control) was injected vehicle only, e.g., 0.9% 30 NaCl, as for the OP-1 treated rats.

Twenty-four hours later, the hearts were removed from all of the rats and the levels of creatine kinase (CK) from the left ventricle (the infarcted region) and 35 from the interventricular septum (the control

nonischemic region) were determined by standard means. By comparing the difference in CK activities in both regions, the amount of CK activity lost from the infarcted region was used as an index of cardiac cellular injury to the infarcted region.

As shown in Figure 1, the data indicate that morphogens (e.g., OP-1) can provide significant cardioprotective effect when provided to ischemic tissue. In the figure, CK loss is graphed as the difference in specific CK activity between the interventricular septum and the left ventricle.

The loss of CK activity by the subgroup of MI rats which received 2 µg of OP-1 just before reperfusion showed some protection as compared with the control MI rats which received injections of vehicle alone, when the levels from both subgroups are measured against, and compared to, the levels obtained for the SHAM MI control. Significant cardioprotection was observed in the subgroup of MI rats which received 20 µg of OP-1 immediately before reperfusion as compared with the control MI rats which received injections of vehicle alone, when the levels from both subgroups are measured against, and compared to, the levels contained within the SHAM MI control.

These data indicate that OP-1 offers significant cardiac protection when administered after ischemia and 30 before reperfusion.

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A variation of this example also may be performed providing morphogen to the animal prior to induction of ischemia. The experiments may be performed both in normal and immune-compromised rats to assess the 5 cardioprotective effects of morphogen administered prior to ischemia.

Vasodilation of Myocardial Infarcted Example 4. Cardiac Tissue Treated with Morphogen

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Certain vasodilators like acetylcholine (ACh) and adenosine diphosphate (ADP, an immune mediator) exert their vasodilation activity only in the presence of intact endothelium, which is stimulated to release a 15 substance termed endothelium-derived relaxing factor (EDRF). If the endothelium is injured so that EDRF is not released, no vasodilation occurs in response to these endothelium-dependent agents. In contrast, several other vasodilators including nitroglycerine 20 (NTG) and nitroprusside, are endothelium-independent dilators, as they dilate blood vessels directly.

The present example demonstrates the ability of OP-1 to prevent the loss of cardioendothelium-dependent 25 relaxation (EDR) activity in the coronary microvasculature following reperfusion of ischemic myocardium, and their ability to reduce myocardial injury 24 hours after morphogen treatment. Briefly, 2 or 24 hours after morphogen treatment ischemia-30 reperfusion injury is induced in isolated rat hearts, the reperfused hearts are are vasodilated with either ACh or NTG. In the absence of morphogen treatment, injured tissue should inhibit ACh-induced vasodilation, but not NTG-induced vasodilation. Morphogen treatment in expected to enhance ACh-induced vasodilation in the reperfused hearts.

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Accordingly, 48 adult male Sprague-Dawley rats (250-330 g) were divided into eight groups of 6 rats each. Twelve rats were subjected to sham myocardial infarcts (SHAM MI) as described in Example 3. 5 hearts of the remaining 36 rats were isolated as follows: one set of twelve rats was injected intravenously with OP-1 24 hours prior to isolation of the heart; another set of rats was injected intravenously with $20\mu g$ of OP-1 2 hours prior to 10 isolation of the heart; the final group of rats was injected with vehicle only (e.g., 0.9% NaCl.). The rats then were anesthetized with pentobarbital sodium (35 mg/kg, intraperitonial); their hearts were isolated and perfused by the Langendorff method at a constant 15 flow (15 ml/min) with oxygenated Krebs-Henseleit solution (Aoki et al. (1988) J. Pharmacol. 95:35).

Each group of rats then were divided into two subgroups of six rats each. Twenty minutes before reperfusion, coronary vasodilator response was measured 20 by inducing constriction with 0.05 μ mol U-44619 (9,11methanoepoxyprostaglandin H2) followed by a vasodilating agent 3 minutes later: subgroup one -15 nmol ACh; subgroup 2 - 15 nmol NTG and the increase in coronary perfusion pressure (CPP) level measured as 25 an indication of vasodilation. When CPP levels returned to normal, the hearts were subjected to ischemia by reducing coronary infusion to 15% of control flow for 30 minutes, then reestablishing normal flow, i.e., reperfusion, for an additional 20 minutes.

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The vasodilator reponse then was remeasured by constriction and administration of vasodilating agent as described above.

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The results of these experiments are shown in FIG 2. Before the ischemic event, both Ach and NTG gave normal vasorelaxant results in all events. The hearts which received OP-1 24 hours prior to ischemia showed 5 an approximately 70% response to ACh while the hearts which received OP-1 2 hours prior to ischemia showed a 55% response to ACh. The group which received vehicle alone showed a 40% response to ACh. Finally, the control group which was not subjected to ischemia 10 showed an ACh response of approximately 95%. This shows that endothelium-dependent vasodilators exert a reduced vasodilator response following ischemia and reperfusion in the rat heart. Moreover, OP-1 significantly preserved endothelium-dependent dilation 15 when provided 24 hours prior to induction of myocardial ischemia. No defect in vasodilation occurred in response to the direct vasodilator (NTG); NTG-induced vasodilation activities were 95% of initial in hearts subject to ischemia and 100% of initial nonischemic 20 hearts.

Example 5. Effect of Morphogen on Neutrophil Adherence

The role of neutrophil adherence in endothelium 25 dysfunction and the cardioprotective effects of morphogens in modulating this activity can be assessed using a standard polymorphonuclear neutrophil (PMN) adherence assay such as described in Lefer et al., (1992) J. Mol. Cell. Cardiol. 24: 385-393, disclosed 30 hereinabove by reference. Briefly, segments of superior mesenteric artery were isolated from rats which had either been treated with morphogen (OP-1, 20 μ g) or 0.9% NaCl, 24 h prior to isolation of the artery. The segments were cleaned, cut into transverse 35 rings of 1-2mm in length, and these were subsequently

cut open and incubated in K-H solution at 37°C, pH 7.4. Neutrophils were prepared and fluorescently labelled using standard procedures (e.g., leukocytes were isolated from rats essentially following the procedure 5 of Pertroft et. al. (1968) Exp Cell Res 50: 355-368, washed in phosphate buffered saline (PBS), purified by gradient centrifugation; and labelled by the method of Yuan et. al. (1990) Microvasc Res 40: 218-229...

Labelled neutrophils then were added to open ring 10 baths and activated with 100nM leukotriene B_4 (LTB₄). Rings were incubated for 20 minutes and the number of neutrophils adhering to the endothelial surface then determined visually by fluorescent microscopy.

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As shown in Figure 3, unstimulated PMNs (i.e., PMNs alone) added to the baths did not significantly adhere to the vascular endothelium. In rings taken from rats injected with 0.9% NaCl, activation of neutrophils with 20 LTB $_4$ (100 nM) greatly increased the number of PMNs adherent to the endothelium (P<0.001). OP-1 (20 μ g administered 24 h prior) significantly inhibited adherence of PMNs activated by LTB₄ (P<0.01 from control).

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In Vivo Models for Ischemic-Reperfusion Example 6. Protection in Lung, Nerve and Renal Tissue.

Other tissues seriously affected by ischemicreperfusion injury include neural tissue, renal tissue 30 and lung tissue. The effect of morphogens on alleviating the ischemic-reperfusion injury in these tissues may be assessed using methodologies and models 35 known to those skilled in the art, and disclosed below.

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Similarly, a methodology also is provided for assessing the tissue-protective effects of a morphogen on damaged lung tissue following hyperoxia injury.

For example, the rabbit embolic stroke model provides a useful method for assessing the effect of 5 morphogens on tissue injury following cerebral ischemia-reperfusion. The protocol disclosed below is essentially that of Phillips et al. (1989) Annals of 10 Neurology 25:281-285, the disclosure of which is herein incorporated by reference. Briefly, white New England rabbits (2-3kg) are anesthesized and placed on a respirator. The intracranial circulation then is selectively catheterized by the Seldinger technique. 15 Baseline cerebral angiography then is performed, employing a digital substration unit. The distal internal carotid artery or its branches then is selectively embolized with 0.035 ml of 18-hour-aged autologous thrombus. Arterial occlusion is documented 20 by repeat angiography immediately after embolization. After a time sufficient to induce cerebral infarcts (15 minutes or 90 minutes), reperfusion is induced by administering a bolus of a reperfusion agent such as the TPA analogue Fb-FB-CF (e.g., 0.8 mg/kg over 2 25 minutes).

The effect of morphogen on cerebral infarcts can be assessed by administering varying concentrations of morphogens, e.g., OP1, at different times preceding or following embolization and/or reperfusion. The rabbits are sacrificed 3-14 days post embolization and their brains prepared for neuropathological examination by fixing by immersion in 10% neutral buffered formalin

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for at least 2 weeks. The brains then are sectioned in a coronal plane at 2-3 mm intervals, numbered and submitted for standard histological processing in paraffin, and the degree of neutral tissue necrosis determined visually.

The renal-protective effects of morphogens on renal ischemia-reperfusion injury readily can be assessed using the mouse model disclosed by Oueliette, et al.

10 (1990), J. Clin. Invest. 85:766-771, the disclosure of which is hereby incorporated by reference. Briefly, renal ischemia is induced surgically in 35-45 days old out-bred Swiss male mice by performing a standard right nephrectomy, and occluding the artery to the left

15 kidney with a microaneurism clamp for 10-30 minutes. Morphogen then may be provided parentally, at various times prior to or following occulsion and/or reperfusion. The effects of morphogen then may be assessed by biological evaluation and histological evaluation using standard techniques well known in the art.

The tissue protective effects of morphogen on tissue exposed to lethally high oxygen concentrations

25 may be assessed by the following procedure. Adult rats (275-300 gms) first are provided with morphogen (e.g., hOP1) or vehicle only, and then are exposed to 96-98% oxygen essentially as described by Rinaldo et al (1983)

Am. Rev. Respir. Dis. 130:1065, to induce hyperoxia.

30 Animals are housed in plastic cages (38 cm x 48 xm x 21 cm). A cage containing 4-5 animals is placed in a 75 liter water-sealed plexiglass chamber. An atmosphere of 96-98% oxygen then is maintained by delivery of 02 gas (liquid 02). Gas flow through the chamber is adjusted to maintain at least 10 air changes/hr.,

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temperature at 22 ± 1°C, minimal levels of condensation within the cage, and carbon dioxide concentration of < 0.5% as measured with a mass spetrophotometric medical gas analyzer.

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At the end of 72 hours all survivors are observed at room air for 1.5 hours and at longer time periods to assess degree of respiratory distress and cyanosis induced by the initial insult and subsequent immune 10 cell-mediated damage. The number of survivors at the end of the challenge is recorded and the treated groups compared with the untreated control group by chi-square test of proportions. Several of the surviving animals for each group are randomly chosen for histological 15 processing of lung tissue.

Lung tissue for histological processing is fixed by infusion of 10% buffered formalin through a tracheal cannula at a constant pressure of 20 cm H20. After 20 fixation for 24-48 hours, sections from each lobe are cut and subsequently stained with hematoxylin and eosin. Coded slides then are examined, preferably in a double-blind fashion for evidence of pathological changes such as edema, interstitial cellularity, and 25 inflammatory response.

Morphogen Inhibition of Cellular and Example 7. Humoral Inflammatory Response

Morphogens described herein inhibit multinucleation of mononuclear phagocytic cells under conditions where 30 these cells normally would be activated, e.g., in response to a tissue injury or the presence of a foreign substance. For example, in the absence of 35 morphogen, an implanted substrate material (e.g.,

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implanted subcutaneously) composed of, for example, mineralized bone, a ceramic such as titanium oxide or any other substrate that provokes multinucleated giant cell formation, rapidly becomes surrounded by multinucleated giant cells, e.g., activated phagocytes

- 5 multinucleated giant cells, e.g., activated phagocytes stimulated to respond and destroy the foreign object. In the presence of morphogen however, the recruited cells remain in their mononuclear precursor form and the matrix material is undisturbed. Figure 4
- illustrates this effect of morphogens, in a schematic representation of histology results of a titanium oxide substrate implanted subcutaneously. In the figure, "mg" means mononuclear giant cells and "ob" means osteoblasts. The substrate represented in Fig. 4B was
 - osteoblasts. The substrate represented in Fig. 4B was implanted together with morphogen (OP-1) and newly formed osteoblasts are evident surrounding the substrate. By contrast, the substrate represented in substrate. By contrast, the substrate represented in Fig. 4A was implanted without morphogen and extensive multinucleated giant cell formation is evident
 - 20 surrounding the substrate. Accordingly, the morphogens' effect in inhibiting excessive bone mass loss in a mammal also may include inhibiting activation of these cells.
 - suppress antibody production stimulated in response to a foreign antigen in a mammal. Specifically, when a bovine bone collagen matrix alone was implanted in a bony site in a rat, a standard antibody response to the collagen is stimulated in the rat as determined by standard anti-bovine collagen ELISA experiments performed on blood samples taken at four week intervals following implantation (e.g., between 12 and 20 weeks.) Serum anti-collagen antibody titers, measured by ELISA essentially following the procedure described by

Nagler-Anderson et al, (1986) PNAS 83:7443-7446, the disclosure of which is incorporated herein by reference, increased consistently throughout the experiment. However, when the matrix was implanted together with a morphogen (e.g., OP-1, dispersed in the matrix and adsorbed thereto, essentially as described in U.S. Pat. No. 4,968,590) anti-bovine collagen antibody production was suppressed significantly. This ability of morphogen to suppress the humoral response is further evidence of morphogen utility in alleviating tissue damage associated with autoimmune diseases, including autoantibody diseases, such as rheumatoid arthritis.

15 Example 8. Morphogen protection of Gastrointestinal
Tract Mucosa from Ulceration and
Inflammation

inflammatory disease which involves ulcerations of the mouth mucosa as a consequence of, e.g., radiation therapy or chemotherapy. While not typically a chronic disease, the tissue destructive effects of oral mucositis mirror those of chronic inflammatory diseases such as IBD. The example below demonstrates morphogen efficacy in protecting the oral mucosa from oral mucositis in a hamster model, including both inhibiting inflammatory ulceration and enhancing regeneration of ulcerated tissue. Details of the protocol can be found in Sonis, et al., (1990) Oral Surg. Oral Med. Oral Pathol 69: 437-443, the disclosure of which is incorporated herein by reference. Based on these data,

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the morphogens described herein should be efficacious in treating chronic inflammatory diseases including IBD, arthritis, psoriasis and psoriatic arthritis, multiple sclerosis, and the like.

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Golden syrian hamsters (6-8 wks old, Charles River Laboratories, Wilmington, MA) were divided into 3 test groups: Group 1, a placebo (e.g., saline) control, and a morphogen low dose group (100 ng) and a morphogen 10 high dose group (1 μ g), Groups 2 and 3, respectively. Morphogen dosages were provided in 30% ethanol. group contained 12 animals.

Beginning on day 0 and continuing through day 5, 15 Groups 2 and 3 received twice daily morphogen applications. On day 3, all groups began the mucositis-induction procedure. 5-fluorouracil (60 mg/kg) was injected intraperitoneally on days 3 and 5. On day 7, the right buccal pouch mucosa was 20 superficially irritated with a calibrated 18 gauge needle. In untreated animals, severe ulcerative mucositis was induced in at least 80% of the animals by day 10.

For each administration of the vehicle control (placebo) or morphogen, administration was performed by 25 first gently drying the cheek pouch mucosa, then providing an even application over the mucosal surface of the vehicle or morphogen material. A 30 hydroxypropylcellulose-based coating was used to maintain contact of the morphogen with the mucosa. This coating provided at least 4 hours of contact time.

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On day 12, two animals in each group were sacrificed for histological studies. The right buccal pouch mucosa and underlying connective tissue were dissected and fixed in 10% formalin using standard 5 dissection and histology procedures. The specimens were mounted in paraffin and prepared for histologic examination. Sections then were stained with hematoxylin and eosin and were examined blindly by three oral pathologists with expertise in hamster 10 histology and scored blind against a standard mucositis panel. The extent of atrophy, cellular infiltration, connective tissue breakdown, degree of ulceration and epithelialization were assessed.

The mean mucositis score for each group was determined daily for each experimental group for a 15 period of 21 days by photography and visual examination of the right buccal cheek pouch. Differences between groups were determined using a standard 't' test, e.g., 20 the Students' 't' test. In addition, data was evaluated between groups by comparing the numbers of animals with severe mucositis using Chi Square The significance of differences statistical analysis. in mean daily weights also was determined.

The experimental results are presented in Fig. 5, which graphs the effect of morphogen (high dose, squares; low dose, diamonds) and placebo (circles) on mean mucositis scores. Both low and high morphogen 30 doses inhibit lesion formation significantly in a dosedependent manner. In addition, histology results consistently showed significantly reduced amounts of

tissue atrophy, cellular debris, and immune effector cells, including macrophages and activated neutrophils, in the morphogen-treated animals, as compared with the untreated, control animals.

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Morphogen Effect on Fibrogenesis and Scar Example 9. Tissue Formation .

The morphogens described herein induce tissue 10 morphogenesis of damaged or lost tissue. The ability of these proteins to regenerate new tissue enhances the anti-inflammatory effect of these proteins. Provided below are a series of in vitro experiments demonstrating the ability of morphogens to induce 15 migration and accumulation of mesenchymal cells. addition, the experiments demonstrate that morphogens, TGF- β , do not stimulate fibrogenesis or scar tissue formation. Specifically, morphogens do not stimulate production of collagen, hyaluronic acid (HA) 20 or metalloproteinases in primary fibroblasts, all of which are required for fibrogenesis or scar tissue formation. By contrast, $TGF-\beta$, a known inducer of fibrosis, but not of tissue morphogenesis, does stimulate production of these fibrosis markers.

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Chemotaxis and migration of mesenchymal progenitor cells were measured in modified Boyden chambers essentially as described by Fava, R.A. et al (1991) J. Exp. Med. 173: 1121-1132, the disclosure of which is 30 incorporated herein by reference, using polycarbonate filters of 2, 3 and 8 micron ports to measure migration of progenitor neutrophils, monocytes and fibroblasts. Chemotaxis was measured over a range of morphogen concentrations, e.g., 10^{-20} M to 10^{-12} M OP-1. For 35 progenitor neutrophils and monocytes, $10^{-18}-10^{-17} \text{M OP-1}$

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consistently induced maximal migration, and 10⁻¹⁴ to 10⁻¹³M OP-1 maximally induced migration of progenitor fibroblasts. In all cases the chemotactic activity could be inhibited with anti-OP-1 antibody. Similar migration activities also were measured and observed with TGF-β.

The effect of morphogen on fibrogenesis was determined by evaluating fibroblast production of hyaluronic acid (HA), collagen, collagenese and tissue inhibitor of metalloproteinases (TIMP).

Human fibroblasts were established from explants of infant foreskins and maintained in monolayer culture 15 using standard culturing procedures. (See, for example, (1976) J. Exp. Med. 144: 1188-1203.) Briefly, fibroblasts were grown in maintenance medium consisting of Eagle's MEM, supplemented with nonessential amino acids, ascorbic acid (50 μ g/ml), NaHCO₃ and HEPES 20 buffers (pH 7.2), penicillin (100 U/ml), streptomycin (100 μ g/ml), amphotericin B (1 μ g/ml) and 9% heat inactivated FCS. Fibroblasts used as target cells to measure chemotaxis were maintained in 150 mm diameter glass petri dishes. Fibroblasts used in assays to 25 measure synthesis of collagen, hyaluronic acid, collagenase and tissue inhibitors of metalloproteinases (TIMP) were grown in 100 mm diameter plastic tissue culture petri dishes.

of hyaluronic acid, collagens, collagenase and TIMP were determined by standard assays (See, for example, Posttethwaite et al. (1989) J. Clin. Invest. 83: 629-636, Posttethwaithe (1988) J./ Cell Biol. 106: 311-318 and Clark et al (1985) Arch. Bio-chem Biophys. 241: 36-

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44, the disclosures of which are incorporated by reference.) For these assays, fibroblasts were transferred to 24-well tissue culture plates at a density of 8 \times 10⁴ cells per well. Fibroblasts were 5 grown confluency in maintenance medium containing 9% FCS for 72 h and then grown in serum-free maintenance medium for 24 h. Medium was then removed from each well and various concentrations of OP-1 (recombinantly produced mature or soluble form) or TGF- β -1 (R&D 10 Systems, Minneapolis) in 50 μ 1 PBS were added to triplicate wells containing the confluent fibroblast monolayers. For experiments that measured production of collagenase and TIMP, maintenance medium (450 μ l) containing 5% FCS was added to each well, and culture 15 supernatants were harvested from each well 48 h later and stored at -70°C until assayed. For experiments that assessed HA production, maintenance medium (450 μ l) containing 2.5% FCS was added to each well, and cultures grown for 48 h. For experiments that measured 20 fibroblast production of collagens, serum-free maintenance medium (450 μ l) without non-essential amino acids was added to each well and cultures grown for 72 h. Fibroblast production of HA was measured by labeling newly synthesized glycosaminoglycans (GAG) 25 with [3H]-acetate the last 24 h of culture and quantitating released radioactivity after incubation with hyaluronidase from Streptomyces hyalurolyticus (ICN Biochemicals, Cleveland, OH) which specifically degrades hyaluronic acid. Production of total collagen 30 by fibroblasts was measured using a collagenasesensitive protein assay that reflects [3H]-proline incorporation the last 24 h of culture into newly synthesized collagens. Collagenase and TIMP protein levels in fibroblast cultures supernatants was measured 35 by specific ELISAs.

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As shown in Fig. 6, OPI does not stimulate significant collagen or HA production, as compared with TGF-β. In the figure, panel A shows OP-1 efect on collagen production, panel B shows TGF-β effect on collagen production, and panels C and D show OP-1 (panel C) and TGF-β (panel D) effect on HA production. (panel C) and TGF-β (panel D) effect on HA production. The morphogen results were the same whether the soluble or mature form of OPI was used. By contrast, the latent form of TGF-β (e.g., pro domain-associated form of TGF-β) was not active.

Example 10. Morphogen Inhibition of Epithelial Cell Proliferation

This example demonstrates the ability of morphogens to inhibit epithelial cell proliferation in vitro, as 15 determined by ³H-thymidine uptake using culture cells from a mink lung epithelial cell line (ATCC No. CCL 64), and standard mammalian cell culturing procedures. 20 Briefly, cells were grown to confluency in Eagle's minimum essential medium (EMEM) supplemented with 10% fetal bovine serum (FBS), 200 units/ml penicillin, and 200 μ g/ml streptomycin, and used to seed a 48-well cell culture plate at a cell density of 200,000 cells per 25 well. When this culture became confluent, the media was replaced with 0.5 ml of EMEM containing 1% FBS and penicillin/streptomycin and the culture incubated for 24 hours at 37 C. Morphogen test samples in EMEM containing 5% FBS then were added to the wells, and the 30 cells incubated for another 18 hours. After incubation, 1.0 μ Ci of 3 H-thymidine in 10 μ l was added to each well, and the cells incubated for four hours at 37 C. The media then was removed and the cells washed once with ice-cold phosphate-buffer saline and DNA 35 precipitated by adding 0.5 ml of 10% TCA to each well

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and incubating at room temperature of 15 minutes. The cells then were washed three times with ice-cold distilled water, lysed with 0.5 ml 0.4 M NaOH, and the lysate from each well then transferred to a scintillation vial and the radioactivity recorded using a scintillation counter (Smith-Kline Beckman).

The results are presented in Table III, below. The anti-proliferative effect of the various morphogens 10 tested was expressed as the counts of 3H-thymidine (x 1000) integrated into DNA, and were compared with untreated cells (negative control) and TGF- β (1 ng), a local-acting factor also known to inhibit epithelial cell proliferation. COP-5 and COP-7 are biosynthetic 15 constructs that previously have been shown to have osteogenic activity, capable of inducing the complete cascade resulting in endochondral bone formation in a standard rat bone assay (see U.S. Pat. No. 5,011,691.) The morphogens significantly inhibit epithelial cell 20 proliferation. Similar experiments, performed with the morphogens COP-16, bOP (bone-purified osteogenic protein, a dimeric protein comprising CBMP2 and OP-1), and recombinant OP-1, also inhibit cell proliferation. bOP and COP-16 also induce endochondral bone formation 25 (see US Pat. No. 4,968,590 and 5,011,691.)

TABLE III

		Thymidine uptake (x 1000)
		50.048, 53.692
30	control	11.874
	COP-7-1 (10 ng)	11.136
	COP-7-2 (3 ng)	16.094
	COP-5-1 (66 ng)	14.43
	COP-5-2 (164 ng)	1.86, 1.478
35	TGF-β (1 ng)	

Example 11. Morphogen Treatment of a Systemic Inflammatory Disease

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The following example provides a rat adjuvantinduced arthritis model for demonstrating morphogen efficacy in treating arthritis and other systemic inflammatory diseases. Rat adjuvant-induced arthritis 10 induces a systemic inflammatory disease with bone and cartilage changes similar to those observed in rhematoid arthritis, but in an accelerated time span (see, for example, Pearson (1964) Arth. Rheum. 7:80). A detailed description of the protocol is provided in 15 Walz, et al., (1971) J. Pharmac. Exp. Ther. 178: 223-231, the disclosure of which is incorporated herein by reference.

Briefly, Sprague-Dawley female rats (e.g., Charles 20 River Laboratories, Wilmington, MA) are randomized into 3 groups: control; morphogen, low dose (e.g., 1-10 μ g/kg weight per day) and morphogen, high dose (e.g., 10-20 μ g/kg weight per day), referred to as Groups 1, 2, and 3, respectively.

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Adjuvant arthritis is induced in all three groups by injection of 0.05 ml of a suspension of 1.5% dead Mycobacterium butyricum in mineral oil into the subplantar surface of the right hand paw. On Day 18 30 after adjuvant injection, the limb volumes of both hind limb are determined. In the absence of morphogen treatment, a systemic arthritic condition is induced in adjuvant-injected rats by this time, as determined by significant swelling of the uninjected hind limbs (< 2.3 ml, volume measured by mercury displacement).

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Subsequent determinations of paw edema and x-ray scores are made on the uninjected hind limb. Rats in Group 2 and 3 also are dosed orally daily, beginning on Day 1, with morphogen. Limb volumes are recorded on Days 29 and 50 after adjuvant injection and edema determined by volume difference compared to Day 18. The uninjected hind limb on each rat is x-rayed on Day 50 and the joint damage assayed on an arbitrary scale of 1 to 10 (1=no damage, 10=maximum damage). Data on differences between control and treated groups (Day 29 edema, Day 50 edema and Day 50 x-ray scores) are analyzed by using a standard "t-test. Morphogen-treated rats show consistently reduced joint damage (e.g., decreased in edema and in x-ray scores) as compared with untreated control rats.

As another, alternative example, Groups 2 and 3 are dosed daily with morphogen beginning on Day 18 and continuing through Day 50 to demonstrate the efficacy of morphogens in arthritic animals.

Example 12. Morphogen Inhibition of Localized Edema

The following example demonstrates morphogen
25 efficacy in inhibiting a localized inflammatory
response in a standard rat edema model. Experimental
rats (e.g., Long-Evans from Charles River Laboratories,
Wilmington, MA) are divided into three groups: Group
1, a negative control, which receives vehicle alone;
30 Group 2, a positive control, to which is administered a
Well-known characterized anti-inflammatory agent
(e.g., indomethacin), and Group 3, to which morphogen
is provided.

Groups 2 and 3 may be further subdivided to test
low, medium and high doses (e.g., Group 2: 1.0 mg/kg,
3.0 mg/kg and 9.0 mg/kg indomethacin; Group 3: 0.1-5µg;
5-20µg, and 20-50µg of morphogen). Sixty minutes after
indomethacin or morphogen is provided to the rats of
Group 2 or 3 (e.g., as by injection into the tail vein,
or by oral gavage) inflammation is induced in all rats
by a sub-plantar injection of a 1% carrageenin solution
(50µl) into the right hind paw. Three hours after
carrageenin administration paw thickness is measured as
an indication of edema (e.g., swelling) and induced
inflammatory response to the injected carrageenin
solution.

by three hours after carrageenin injection.

Inflammation also is measured by histology by standard means, following euthanasia e.g.: the right hind paw from each animal is removed at the ankle joint and weighed and foot pad tissue is fixed in 10% neutral buffered formalin, and slides prepared for visual examination by staining the prepared tissue with hematoxylin and eosin.

The morphogen-treated rats show substantially reduced edema induction following carrageenin injection as compared with the untreated rats.

Example 13. Morphogen Treatment of Allergic Encephalomyelitis

The following example demonstrates morphogen

efficacy in treating experimental allergic
encephalomyelitis (EAE) in a rat. EAE is a
well-characterized animal model for multiple sclerosis,
an autoimmune disease. A detailed description of the
protocol is disclosed in Kuruvilla, et al., (1991) PNAS

88:2918-2921, the disclosure of which is incorporated
herein by reference.

Charles River Laboratories, Wilmington, MA) by

15 injection of a CNS tissue (e.g., spinal cord)

16 homogenate in complete Freund's adjuvant (CFA) on days

17 -44, -30 and 0 (last day of immunization), by

18 subcutaneous injection to three sites on the animal's

19 back. Morphogen is administered daily by

20 interperitoneal injection beginning on day -31.

21 Preferably, a series of morphogen dose ranges is

22 evaluated (e.g., low, medium and high) as for

23 Example 12, above.) Control rats receive morphogen

24 vehicle only (e.g. 0.9% NaCl or buffered saline). Rats

25 are examined daily for signs of disease and graded on

an increasing severity scale of 0-4.

In the absence of morphogen treatment, significant neurological dysfunction (e.g., hind and fore limb weakness, progressing to total hind limb paralysis) is evident by day +7 to +10. Hematology, serum chemistry profiles and histology are performed to evaluate the

degree of tissue necropsy using standard procedures.

Morphogen treatment significantly inhibits the
neurological dysfunction normally evident in an EAE
animal. In addition, the histopathological markers
typically associated with EAE are absent in the
morphogen-treated animals.

Example 14. Morphogen Treatment of Collagen-Induced Arthritis

The following example demonstrates the efficacy of 10 morphogens in inhibiting the inflammatory response in a collagen-induced arthritis (CIA) in a rat. CIA is a well-characterized animal model for rheumatoid 15 arthritis, an autoimmune disease. The protocol disclosed is essentially that disclosed in Kuruvilla et al., (1991) PNAS 88:2918-2921, incorporated by reference hereinabove. Briefly, CIA is induced in experimental rats (e.g., Long-Evans, Charles River 20 Laboratories, Wilmington), by multiple intradermal injection of bovine Type II collagen (e.g., $100\mu g$) in CFA (0.2 ml) on Day 1. Animals are divided into two groups: Group 1, control animals, which receive vehicle alone, and Group 2: morphogen-treated animals, which, 25 preferably, are subdivided into low, medium and high dose ranges, as described for Example 13, above. Morphogen is administered daily (e.g., by tail vein injection) beginning at different times following collagen injection, e.g., beginning on day 7, 14, 28, 30 35 and 42. Animals are evaluated visually and paw thickness and body weight is monitored throughout the experiment. Animals are sacrificed on day 60 and the proximal and distal limb joints, and ear, tail and spinal cord prepared for histological evaluation as described for Examples 12 and 13, above. In a

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variation of the experiment, morphogen may be administered for prescribed periods, e.g., five day periods, beginning at different times following collagen injection (e.g., on days 0-4, 7-11, 14-18, 28-32.)

In the absence of morphogen treatment, an arthritic condition typically is induced by 30 days post collagen injection. In morphogen-treated animals, CIA is suppressed and the histopathological changes typically evidenced in control CIA-induced animals are absent:

e.g., accumulations of activated mononuclear inflammatory cells and fibrous connective tissue. In addition, consistent with the results in Example 7, above, serum anti-collagen antibody titers are suppressed significantly in the morphogen-treated animals.

Example 15. Screening Assay for Candidate Compounds which Alter Endogenous Morphogen Levels

Candidate compound(s) which may be administered to affect the level of a given morphogen may be found using the following screening assay, in which the level of morphogen production by a cell type which produces measurable levels of the morphogen is determined with and without incubating the cell in culture with the compound, in order to assess the effects of the compound on the cell. This can be accomplished by detection of the morphogen either at the protein or RNA level. A more detailed description also may be found in USSN 752,861, incorporated hereinabove by reference.

15.1 Growth of Cells in Culture

Cell cultures of kidney, adrenals, urinary bladder, brain, or other organs, may be prepared as described 5 widely in the literature. For example, kidneys may be explanted from neonatal or new born or young or adult rodents (mouse or rat) and used in organ culture as whole or sliced (1-4 mm) tissues. Primary tissue cultures and established cell lines, also derived from 10 kidney, adrenals, urinary, bladder, brain, mammary, or other tissues may be established in multiwell plates (6 well or 24 well) according to conventional cell culture techniques, and are cultured in the absence or presence of serum for a period of time (1-7 days). Cells may be 15 cultured, for example, in Dulbecco's Modified Eagle medium (Gibco, Long Island, NY) containing serum (e.g., fetal calf serum at 1%-10%, Gibco) or in serum-deprived medium, as desired, or in defined medium (e.g., containing insulin, transferrin, glucose, albumin, or 20 other growth factors).

Samples for testing the level of morphogen
production includes culture supernatants or cell
lysates, collected periodically and evaluated for OP-1
production by immunoblot analysis (Sambrook et al.,
eds., 1989, Molecular Cloning, Cold Spring Harbor
Press, Cold Spring Harbor, NY), or a portion of the
cell culture itself, collected periodically and used to
prepare polyA+ RNA for RNA analysis. To monitor de
prepare polyA+ RNA for RNA analysis. To monitor de
according to conventional procedures with an
according to conventional procedures with an
then evaluated to OP-1 synthesis by conventional
immunoprecipitation methods.

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15.2 Determination of Level of Morphogenic Protein

In order to quantitate the production of a morphogenic protein by a cell type, an immunoassay may be performed to detect the morphogen using a polyclonal or monoclonal antibody specific for that protein. For example, OP-1 may be detected using a polyclonal antibody specific for OP-1 in an ELISA, as follows.

1 μ g/100 μ l of affinity-purified polyclonal rabbit IgG specific for OP-1 is added to each well of a 10 96-well plate and incubated at 37°C for an hour. wells are washed four times with 0.167M sodium borate buffer with 0.15 M NaCl (BSB), pH 8.2, containing 0.1% 15 Tween 20. To minimize non-specific binding, the wells are blocked by filling completely with 1% bovine serum albumin (BSA) in BSB and incubating for 1 hour at 37°C. The wells are then washed four times with BSB containing 0.1% Tween 20. A 100 μ l aliquot of an 20 appropriate dilution of each of the test samples of cell culture supernatant is added to each well in triplicate and incubated at 37°C for 30 min. After incubation, 100 μ l biotinylated rabbit anti-OP-1 serum (stock solution is about 1 mg/ml and diluted 1:400 in 25 BSB containing 1% BSA before use) is added to each well and incubated at 37°C for 30 min. The wells are then washed four times with BSB containing 0.1% Tween 20. 100 μ l strepavidin-alkaline (Southern Biotechnology Associates, Inc. Birmingham, Alabama, diluted 1:2000 in 30 BSB containing 0.1% Tween 20 before use) is added to The plates each well and incubated at 37°C for 30 min. are washed four times with 0.5M Tris buffered Saline (TBS), pH 7.2. 50μ l substrate (ELISA Amplification System Kit, Life Technologies, Inc., Bethesda, MD) is 35 added to each well incubated at room temperature for 15

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min. Then, 50 μ l amplifier (from the same amplification system kit) is added and incubated for another 15 min at room temperature. The reaction is stopped by the addition of 50 μ I 0.3 M sulphuric acid. 5 The OD at 490 nm of the solution in each well is recorded. To quantitate OP-1 in culture media, a OP-1 standard curve is performed in parallel with the test samples.

Polyclonal antibody may be prepared as follows. .10 Each rabbit is given a primary immunization of 100 ug/500 μ l E. coli produced OP-1 monomer (amino acids 328-431 in SEQ ID NO:5) in 0.1% SDS mixed with 500 μ l Complete Freund's Adjuvant. The antigen is injected 15 subcutaneously at multiple sites on the back and flanks of the animal. The rabbit is boosted after a month in the same manner using incomplete Freund's Adjuvant. Test bleeds are taken from the ear vein seven days later. Two additional boosts and test bleeds are 20 performed at monthly intervals until antibody against OP-1 is detected in the serum using an ELISA assay. Then, the rabbit is boosted monthly with 100 μg of antigen and bled (15 ml per bleed) at days seven and ten after boosting.

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Monoclonal antibody specific for a given morphogen may be prepared as follows. A mouse is given two injections of E. coli produced OP-1 monomer. The first injection contains $100\mu g$ of OP-1 in complete Freund's 30 adjuvant and is given subcutaneously. The second injection contains 50 μg of OP-1 in incomplete adjuvant and is given intraperitoneally. The mouse then receives a total of 230 μg of OP-1 (amino acids 307-431 in SEQ ID NO:5) in four intraperitoneal injections at 35 various times over an eight month period. One week

prior to fusion, both mice are boosted intraperitoneally with 100 μg of OP-1 (307-431) and 30 μ g of the N-terminal peptide (Ser₂₉₃-Asn₃₀₉-Cys) conjugated through the added cysteine to bovine serum 5 albumin with SMCC crosslinking agent. This boost was repeated five days (IP), four days (IP), three days (IP) and one day (IV) prior to fusion. The mouse spleen cells are then fused to myeloma (e.g., 653) cells at a ratio of 1:1 using PEG 1500 (Boeringer 10 Mannheim), and the cell fusion is plated and screened for OP-1-specific antibodies using OP-1 (307-431) as antigen. The cell fusion and monoclonal screening then are according to standard procedures well described in standard texts widely available in the art.

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The invention may be embodied in other specific forms without departing from the spirit or essential characteristics thereof. The present embodiments are therefore to be considered in all respects as 20 illustrative and not restrictive, the scope of the invention being indicated by the appended claims rather than by the foregoing description, and all changes which come within the meaning and range of equivalency of the claims are therefore intended to be embraced 25 therein.

SEQUENCE LISTING

	(1)	GENERAL	INFOR	fation:
5		(i)APPLIC	ANT:	KUBERASAHPATH, THANGAVEL PANG, ROY H.L. OPPERMANN, HERMANN
10				RUEGER, DAVID C. COHEN, CHARLES H. OZKAYNAK, ENGIN SHART, JOHN
15		(11)	TITL INFL	E OF INVENTION: MORPHOGEN-INDUCED MODULATION OF AMMATORY RESPONSE
		(iii)	NUHB	ER OF SEQUENCES: 33
20		(iv)	(A) (B) (C)	STREET: 35 SOUTH STREET CITY: HOPKINTON STATE: HASSACHUSETTS
25			(E) (F)	COUNTRY: U.S.A. ZIP:
30		(♥)	(A)	UTER READABLE FORM: MEDIUM TYPE: Floppy disk COMPUTER: IBH PC compatible OPERATING SYSTEM: PC-DOS/MS-DOS SOFTWARE: Patent In Release #1.0, Version #1.25
35		(vii)	(A) (B)	R APPLICATION DATA: APPLICATION NUMBER: US 667,274 FILING DATE: 11-MAR-1991
		(viì)		R APPLICATION DATA: APPLICATION NUMBER: US 753,059 FILING DATE: 30-AUG-1991
40		(vii)	743	R APPLICATION DATA: APPLICATION NUMBER: US 752,764 FILING DATE: 30-AUG-1991
		(2)		INFORMATION FOR SEQ ID NO:1:
45	•	(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 97 amino acids (B) TYPE: amino acids (C) TOPOLOGY: linear
50)	(ii)	MOLECULE TYPE: protein

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			į			TRIE	מאסרטי			auu	AGG	allv.
	•		. •		-44	****	. One	DE.		20 11		ally-
4.0				_	A	rino	1 L-3	LSOME	er, o	(-ami	no a	cids
40				_	r a	deri	vati	lve t	nere	OT.		
		141	١ 5	EOUE	ENCE	DESC	RIPI	: NOI	SE	Q ID	NO	2:
		(2 2	-, -	, 								W
							Xaa	Xaa	Xaa	Xaa	Xaa	Xaa
							1				5	
45		V = 2	Vas	Yas	Xaa	a Xaa	xaa	a Xaa	ı Xaa	a Xaa	Xaa	xaa
	•											
		77		. Yaa	yaa	. Хаа	a Xaa	a Xaa	a Cys	s Xaa	ı Xaa	хаа
		_		. V	y s Vs	a Yas	Xaa	a Xaa	a Cvs	xaa	ı Xaa	xaa
50												
			30) - V-:		. Y.s.	a Xas	a Xaa	a Xaa	a Xaa	a Xaa	a Xaa 50
			a Xaa	a xaa	1 Add	a Adi	4	ς				50
		40						_				

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02/0/602	1 CH days	
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	Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Cys	
	Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xa	
	Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xa	
5	Xaa	ģ
•	75 Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Cys	÷.
	85	
	Xaa Cys Xaa	3
10	95	
(2)	INFORMATION FOR SEQ ID NO:3:	
(2)		
	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 97 amino acids	
15	(R) TYPE: amino acids	
	(c) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: protein	
_	(ix) FEATURE: (A) NAME: Generic Sequence 3 (A) NAME: Generic Sequence 3	
20		
	Xaa is independently selectified	
	amino acids as defined in the	
0.5	specification.	
25	PRESENTANTON: SEO ID NO:3:	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:	
	Leu Tyr Val Xaa Phe	
30	9 3	
30	Xaa Xaa Xaa Gly Trp Xaa Xaa Trp Xaa	
	Xaa Ala Pro Gly Xaa Xaa Ala	
	4 <i>5</i>	
35	YAA TYT CYS XAA GIY XAA CYS XAA	
0 4	Xaa Pro Xaa Xaa Xaa Xaa Xaa	
	• • • • • • • • • • • • • • • • • • • •	
	Xaa Xaa Asn His Ala Xaa Xaa 45	
40	Xaa Xaa Leu Xaa Xaa Xaa Xaa	
	. 30	
	Xaa Xaa Xaa Xaa Xaa Xaa Cys	
	EE .	
45	Cys Xaa Pro Xaa Xaa Xaa Xaa Xaa Xaa	
	Yaa Xaa Xaa Leu Xaa Xaa Xaa	
	マー・マー・アン・アン・アン・アン・アン・アン・アン・アン・アン・アン・アン・アン・アン・	Ġ
	Xaa Xaa Xaa Val Xaa Leu Xaa 80	
50	Xaa Xaa Xaa Met Xaa Val Xaa	S
	85	
0	xaa Cys Gly Cys Xaa	
	95	

(2)	INFORMATION FOR SEQ ID NO:4:	
5		(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 102 amino acids (B) TYPE: amino acids (C) TOPOLOGY: linear	
10		<pre>(ii) MOLECULE TYPE: protein (ix) FEATURE: (A) NAME: Generic Sequence 4 (D) OTHER INFORMATION: wherein each</pre>	ı
15		_	
		(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:	
20		Cys Xaa Xaa Xaa Leu Tyr Val Xaa Phe	
20		Xaa Xaa Xaa Gly Trp Xaa Xaa Trp Xaa	
		Xaa Ala Pro Xaa Gly Xaa Xaa Ala	
25		Xaa Tyr Cys Xaa Gly Xaa Cys Xaa 35	•
		Xaa Pro Xaa Xaa Xaa Xaa 40	
20		Asn Xaa Xaa Asn His Ala Xaa Xaa 45 50	
30		Xaa Xaa Leu Xaa Xaa Xaa Xaa Xaa 55	
		Xaa Xaa Xaa Xaa Xaa Xaa Cys 60 65	
35		Cys Xaa Pro Xaa Xaa Xaa Xaa Xaa 70	
		Xaa Xaa Xaa Leu Xaa Xaa Xaa 75	
40		Xaa Xaa Xaa Val Xaa Leu Xaa 85	
40		Xaa Xaa Xaa Met Xaa Val Xaa 90 95	
		Xaa Cys Gly Cys Xaa 100	
45	(2)	INFORMATION FOR SEQ ID NO:5: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 139 amino acids (B) TYPE: amino acids	
50		(C) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (ix) FEATURE: (A) NAME: hOP-1 (mature form)	

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		SEQUENC	R DESCI	RIPTIC	: NC	SEQ II	NO:	:
	(xi)	2500500	سابار ي				Ser	Gln
	Ser T	hr Gly	Ser	Lys	Gln	Arg	261	91
	1	_		Thr	Pro	Lys	Asn	Gln
5		rg Ser	груз		15		77-1	Ala
	10 Glu A	la Lev	1 Arg	Met	Ala	Asn	Val	NIG.
	יי חדה	20		a	Nen	25 Gln	Arg	Gln
	Glu A	lsn Sei		Ser	Asp		35	
10	_	3(_	His	Glu	Leu	Tyr	Val
	Ala C	ys Ly	40				C1-	45 Asp
	Ser E	he Ar		Leu	Gly	Trp	Gln	vañ
	261 +		_	50	Glu	Gly	Tyr	Ala
15	Trp 1	Ile Il	e Ala	Pro	60		_	
73	55	m	r Cys	Glu	Gly	Glu	Cys	Ala
	Ala 7	Tyr Ty 65	r cys		_	70	2	Ala
	Phe I	bro re	u Asn	Ser	TYI	Met	Asn 80	A-G
20		7		TIC	Val	Gln	Thr	Leu
20	Thr	Asn Hi	s Ala 85	Ile	7. (2.4.		•	90 .
		His Ph		Asn	Pro	Glu	Thr	Val
	Val	His Ph		9.5		D	Thr	Gln
3 E	Pro	Lys Pr	o Cys	Cys	Ala 105	Pro	T +1-	
25	100		_ +1-	Ser	Val	Leu	Tyr	Phe
		Asn Al	a Ile	367	,	115	_	T
		110 Asp Se	r Ser	Asn	Val	Ile	Leu	Lys
	Asp	Asp Se			1	Val	125 Arg	Ala
30	Lys	Tyr A	g Asn	Met	Val	Val	Hr. A	135
		•	130		•			
	- 2	Gly C						
_	******	RMATION	FOR SE	Q ID	NO:6:			
35 (2)	(i)	SEQUE	ACE COW				a .	
	(+)	(A) L	ength:	137			.	
		(B) T	YPE: a	mino	near	1		
		(C) T	OPOLOGY ULE TYP	· 上上 注: D	rotei	n .	•	
40	(ii)	MOLEC! FEATU	RE:	F		- -	 \	
	(ix)			10P-1	(matu	re to	TTU NO	:6:
	(xi)	SEQUE	AME: II NCE DES	CRIPT	: ION;	357		
	, /							
45	Ser	Thr G	ly Gly	, raine		_		C1 n
	1	2-4	er Lys		Pro		Asr	Gln
	Asn 10	Arg S	_		1		· Val	Ala
	Glu	Ala I	eu Arg	g Met	Ala	a Ser		
50	324	20		- 80:	r As			Gln
טכ	Glu	Asn S	er Se	r Se	r və		. 3	5
			30					

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	Ala	Cys	Lys	Lys	His	Glu	Leu	Tyr	Val 45	
	Ser	Phe	Arg	40 Asp	Leu 50	Gly	Trp	Gln	Asp	•
5	Trp	Ile	Ile	Ala	Pro	Glu 60	Gly	Tyr	Ala	
	55 Ala	Tyr	Tyr	Cys	Glu	Gly	Glu 70	Cys	Ala	
	Phe	65 Pro	Leu 75	Asn	Ser	Tyr	Met	Asn 80	Ala	
10	Thr	Asn	His	Ala 85	Ile	Val	Gln	Thr	Leu 90	
·	Val	His	Phe	Ile	Asn 95	Pro	Asp	Thr	Val	
15	Pro 100	Lys	Pro	Cys	Cys	Ala 105	Pro	Thr	Gln	
	Leu	Asn 110	Ala	Ile	Ser	Val	Leu 115	Ţyr	Phe	
20	Asp	Asp	Ser 120	Ser	Asn	Val	Ile	Leu 125	Lys	
	Lys	Tyr	Arg	Asn 130	Met	Val	Val	Arg	Ala 135	
	Cys	Gly	Cys	His					•	
25 (2)	INFO	(A) (B)	UENCE LENG TYPE TOPO	CHAR TH: am LOGY:	ACTER 139 a ino a lin	ISTIC mino cids ear	acias			
30	(ii) (ix)	MOL FEA	ECULE TURE:	TYPE	: pr	otein		-m \		
	(xi)	• •	name UENCE	DESC	RIPTI	Macui ON:	e for SEQ I	D'NO:	7:	
35	Ala	Val	Arg	Pro	Leu 5	Arg	Arg	Arg	Gln	
	Pro 10	Lys	Lys	Ser	Asn	Glu 15	Leu	Pro	Gln	
40	Ala	Asn 20	Arg	Leu	Pro	Gly	11e 25	Phe	Asp	
40	Asp	Val	His 30	Gly	Ser	His	Gly	Arg 35	Gln	
	Val	Cys	Arg	Arg 40	His	Glu	Leu	Tyr	Val 45	
45	Ser	Phe	Gln	Asp	Leu 50	Gly	Trp	Leu	Asp	
	Trp 55	Val	Ile	Ala	Pro	Gln 60	Gly	Tyr	Ser	
50	Ala	Tyr 65	Tyr	Cys	Glu	Gly	Glu 70	Cys	Ser	
50	Phe	Pro	Leu 75	Asp	Ser	Cys	Met	Asn 80	Ala	

			•		- 10)4 -						
•									PCT/US92	2/07358		
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110 75/04072									Leu			
	Thr	Asn	His	Ala 85	Ile	Leu	Gln	ser	90			
	Val	His	Leu	Met	Lys 95	Pro	Asn	Ala	Val			
5	Pro	Lys	Ala	Cys	Cys	Ala 105	Pro	Thr	Lys			
	100 Leu	Ser	Ala	Thr	Ser	Val	Leu 115	Tyr	Tyr		<u>\$</u>	
	Asp	110 Ser		Asn	Asn	Val	Ile	Leu 125	Arg		•	f
10	Lys	His	120 Arg	Asn	Met	Val	Val	Lys	Ala 135			
	Cys	Gly	Cys	130 His								
15 (2)	-	(A) (B)	UENCE LENG TYPE	TH: am	139 amino am	mino cids ear	4444	•				
20	(ii) (ix)	MOĹ FEA	ECULE TURE: NAME	TYPE : mO	: pr P-2 (:	matur		m) D NO:	8:			
•	(xi)	SEQ	UENCE	DESC	RIPTI	OM:			_			
25	Ala	Ala	Arg	Pro	Leu 5	Lys	Arg	Arg	Gln			
	Pro	Lys	Lys	Thr	Asn	Glu 15	Leu	Pro	His			
	10 Pro	Asn	Lys	Leu	Pro	Gly	11e 25	Phe	Asp			
30	Asp	Gly	His 30	Gly	Ser	Arg	Gly	Arg 35	Glu			
	Val	Cys	Arg	Arg 40	His	Glu	Leu -	Tyr	Val 45			
35	Ser	Phe	Arg	Asp	Leu 50	Gly	Trp	Leu	Asp			
	Trp	Val	Ile	Ala	Pro	Gln 60	Gly	Tyr	Ser			
	55 Ala	Tyr	Tyr	Cys	Glu	Gly	Glu 70	Cys	Ala			
40	Phe	65 Pro	Leu 75	Asp	Ser	Cys	Met	Asn 80	Ala			
	Thr	Asn	His	Ala 85	Ile	Leu	Gln	Ser	Leu 90			
45	Val	His	Leu	Met	Lys 95	Pro	Asp	Val	Val			
	Pro		Ala	Cys	Cys	Ala 105		Thr	Lys			
	100 Leu	Ser	Ala	Thr	Ser	Val	Leu 115	Tyr	Tyr			3
50	Asp	110 Ser	Ser 120	Asn	Asn	Val	_	Leu 125	Arg			S

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	Lys His Arg Asn Met Val Val Lys Ala 130
	Cys Gly Cys His
5 (2)	INFORMATION FOR SEQ ID NO:9: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 96 amino acids (B) TYPE: amino acids (C) TOPOLOGY: linear
10	(ii) MOLECULE TYPE: protein (ix) FEATURE: (A) NAME: CBMP-2A(fx) (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:
15	Cys Lys Arg His Pro Leu Tyr Val Asp Phe Ser
15	Asp Val Gly Trp Asn Asp Trp Ile Val Ala Pro
	Pro Gly Tyr His Ala Phe Tyr Cys His Gly Glu
20	Cys Pro Phe Pro Leu Ala Asp His Leu Asn Ser
	Thr Asn His Ala Ile Val Gln Thr Leu Val Asn
25	Ser Val Asn Ser Lys Ile Pro Lys Ala Cys Cys
20	Val Pro Thr Glu Leu Ser Ala Ile Ser Met Leu
	Tyr Leu Asp Glu Asn Glu Lys Val Val Leu Lys
30	80 Asn Tyr Gln Asp Met Val Val Glu Gly Cys Gly 90 95
	Cys Arg 100
35 (2)	INFORMATION FOR SEQ ID NO:10: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 101 amino acids (B) TYPE: amino acids
40	(c) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (ix) FEATURE: (ix) NAME: CBMP-2B(fx)
45	(xi) SEQUENCE DESCRIPTION: SEQ 15 HOUSE
45	Cys Arg Arg His Ser
	Leu Tyr Val Asp Phe Ser Asp Val Gly Trp Asn 10 15 10 The Val Ala Pro Pro Gly Tyr Gln Ala
50	Asp Trp Ile Val Ala Pro Pro Gly Tyr Gln Ala 20 25

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	Phe Tyr Cys His Gly Asp Cys Pro Phe Pro Leu	
	Ala Asp His Leu Asn Ser Thr Asn His Ala Ile	
5	Val Gln Thr Leu Val Asn Ser Val Asn Ser Ser 60	
	Ile Pro Lys Ala Cys Cys Val Pro Thr Glu Leu 70	
	Ser Ala Ile Ser Met Leu Tyr Leu Asp Glu 171	
10	Asp Lys Val Val Leu Lys Asn Tyr Gln Glu Met 85 90	
	Val Val Glu Gly Cys Gly Cys Arg 95 100	
15 (2)	INFORMATION FOR SEQ ID NO:11: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 102 amino acids (B) TYPE: amino acids	
20	(C) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (ix) FEATURE: (A) NAME: DPP(fx) (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:	
25 .	Cys Arg Arg His Ser Leu Tyr Val Asp Phe Ser	
	Asp Val Gly Trp Asp Asp Trp Ile Val Ala Pro	
30	Leu Gly Tyr Asp Ala Tyr Tyr Cys His Gly Lys	
	Cys Pro Phe Pro Leu Ala Asp His Phe Asn Ser	
25	Thr Asn His Ala Val Val Gln Thr Leu Val Asn 55 45 50 The Asn Even Ala Cys	
35	Asn Asn Pro Gly Lys Val Pro Lys Ala Cys 65 60 Asn Ser Val Ala Met	
	Cys Val Pro Thr Gln Leu Asp Ser Val Ala Met 75 The Tyr Leu Ash Asp Gln Ser Thr Val Val Leu 85	
4.0	Leu Tyr Leu Ash Asp Gin Sch 85 80 Lys Asn Tyr Gln Glu Met Thr Val Val Gly Cys	
	90	
45	Gly Cys Arg 100	
(2)	INFORMATION FOR SEQ ID NO:12: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 102 amino acids (B) TYPE: amino acids	â
50	(C) TOPOLOGY: linear	2)

```
(ix)
                   FEATURE:
                   (A) NAME:
                             Vgl(fx)
                   SEQUENCE DESCRIPTION:
                                          SEQ ID NO:12:
             (Xi)
            Cys Lys Lys Arg His Leu Tyr Val Glu Phe Lys
 5
            Asp Val Gly Trp Gln Asn Trp Val Ile Ala Pro
            Gln Gly Tyr Met Ala Asn Tyr Cys Tyr Gly Glu
25 30
10
            Cys Pro Tyr Pro Leu Thr Glu Ile Leu Asn Gly
            Ser Asn His Ala Ile Leu Gln Thr Leu Val His
                                  50
            Ser Ile Glu Pro Glu Asp Ile Pro Leu Pro Cys
15
                              60
            Cys Val Pro Thr Lys Met Ser Pro Ile Ser Met
                          70
            Leu Phe Tyr Asp Asn Asn Asp Asn Val Val Leu
                      80
20
            Arg His Tyr Glu Asn Met Ala Val Asp Glu Cys
                                      95
            Gly Cys Arg
            100
25
            INFORMATION FOR SEQ ID NO:13:
    (2)
                  SEQUENCE CHARACTERISTICS:
                   (A) LENGTH: 102 amino acids
                   (B)
                      TYPE: amino acids
                   (C) TOPOLOGY: linear
30
            (ii)
                  MOLECULE TYPE: protein
                  FEATURE:
            (ix)
                   (A) NAME: Vgr-1(fx)
                  SEQUENCE DESCRIPTION: SEQ ID NO:13:
35
            Cys Lys Lys His Glu Leu Tyr Val Ser Phe Gln
            Asp Val Gly Trp Gln Asp Trp Ile Ile Ala Pro
15 20
            Xaa Gly Tyr Ala Ala Asn Tyr Cys Asp Gly Glu
25 30
40
            Cys Ser Phe Pro Leu Asn Ala His Met Asn Ala
                                      40
            Thr Asn His Ala Ile Val Gln Thr Leu Val His
                                  50
45
            Val Met Asn Pro Glu Tyr Val Pro Lys Pro Cys
                                                   65
            Cys Ala Pro Thr Lys Val Asn Ala Ile Ser Val
```

PCT/US92/07358 WO 93/04692 Leu Tyr Phe Asp Asp Asn Ser Asn Val Ile Leu Lys Tyr Arg Asn Met Val Val Arg Ala Cys 90 Gly Cys His 5 100 (2) INFORMATION FOR SEQ ID NO:14: SEQUENCE CHARACTERISTICS: 10 LENGTH: 106 amino acids (A) TYPE: protein -(B) STRANDEDNESS: single (C) TOPOLOGY: linear (D) (ii) MOLECULE TYPE: protein 15 (vi) ORIGINAL SOURCE: (A) ORGANISH: human (F) TISSUE TYPE: BRAIN 20 (ix) FEATURE: (D) OTHER INFORMATION: /product= "GDF-1 (fx)" SEQUENCE DESCRIPTION: SEQ ID NO:14: 25 Cys Arg Ala Arg Arg Leu Tyr Val Ser Phe Arg Glu Val Gly
1 10 Trp His Arg Trp Val Ile Ala Pro Arg Gly Phe Leu Ala Asn Tyr 30 Cys Gln Gly Gln Cys Ala Leu Pro Val Ala Leu Ser Gly Ser Gly Gly Pro Pro Ala Leu Asn His Ala Val Leu Arg Ala Leu Met His
55 35 Ala Ala Ala Pro Gly Ala Ala Asp Leu Pro Cys Cys Val Pro Ala 60 65 70 40 Arg Leu Ser Pro Ile Ser Val Leu Phe Phe Asp Asn Ser Asp Asn Val Val Leu Arg Gln Tyr Glu Asp Het Val Val Asp Glu Cys Gly 45 Cys Arg 105 50

	(2) INFORMATION FOR SEQ ID NO:15:	
5	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 5 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
10	(ii) MOLECULE TYPE: peptide	
10	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:	
15	Cys Xaa Xaa Xaa 1 5	
15	(2) INFORMATION FOR SEQ ID NO:16:	
20	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1822 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
25	(ii) MOLECULE TYPE: cDNA	
	(VI) ORIGINAL SOURCE: (A) ORGANISH: HOHO SAPIENS (F) TISSUE TYPE: HIPPOCAMPUS	
30	<pre>(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 491341 (D) OTHER INFORMATION:/standard_name= "hOP1"</pre>	
35	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:	
	GGTGCGGGCC CGGAGCCCGG AGCCCGGGTA GCGCGTAGAG CCGGCGCG ATG CAC GTG Het His Val	57
40	CGC TCA CTG CGA GCT GCG GCG CCG CAC AGC TTC GTG GCG CTC TGG GCA Arg Ser Leu Arg Ala Ala Pro His Ser Phe Val Ala Leu Trp Ala 5	105
45	CCC CTG TTC CTG CTG CGC TCC GCC CTG GCC GAC TTC AGC CTG GAC AAC Pro Leu Phe Leu Leu Arg Ser Ala Leu Ala Asp Phe Ser Leu Asp Asn 20 30	153
50	GAG GTG CAC TCG AGC TTC ATC CAC CGG CGC CTC CGC AGC CAG GAG CGG Glu Val His Ser Ser Phe Ile His Arg Arg Leu Arg Ser Gln Glu Arg 40 45 50	201

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							• .				•		•			F	CI	/US92/	07358	
	O 93/ CGG Arg			ב פ	بسن	CGC Arg	GAG Glu	ATC Ile	CTC Leu	TCC Ser	ATT Ile	TTG Leu	GGC Gly	TTG Leu	CCC Pro 65	CAC His	CG AI	C g	249	
			CC	G C	,,							mac	GCA Ala	ecc.	ATG	TTC	ΓA	:G	297	
LO	Leu	ASP	Le	G I	LYE	ASII	VIG	90					GGC Gly 95						345	ş
	Gly	CAG Gln	GI	y 1	rne	Ser	105	110	-,-	-•		110	TTC				1.	LJ	393	
13	Pro	Pro	Le	u .	VTS	120	Ten	U 2			125		CTC Leu			130	,		441	
20	ATG Het	GTC Val	: AI He	t	AGC Ser 35	TTC Phe	GTC Val	AAC Asn	CTC	GTG Val 140	GAA Glu	CAT His	GAC Asp	Lys	GAA Glu 145	TIO Phe	P	TC he	489	
25	CAC His	CCA	Al			CAC His	CAT His	CGA	GAG Glu	TTC Phe	CGG	TI:	C GAT e Asi	CTI Let 160	TC(L Set	Ly:	G A	TC le	537	
30	CCA Pro	GA/ Glu	. G(GAA Glu	GCT Ala	GTC Val	ACG Thi		GCC Ala	GA/	TTO	C CGG e Arg	G ATO	C TAC	C AA	G G s: A	AC Asp	585	
25	Tyr	AT	C C	rg	GIU	ALE	185	j j				19	C CG e Ar O				•	190	633	
35	Gli	GT Va	ŀĻ	eu	GIN	200)	3. 20.			20	5	G GA			21	·		681	
40	Ası) Se	T A	rg	1111 215	CTO	IG u Tr	P	_	22	0		C TO		22	.5			729	
45	IL	e In	I A	CC la	ACC Thi		•		23	5			T CC	24	+0				7.77	
	GG(C CI				TC 1 Se	G GI r Va	G GA 1 G1 25		G CI	G GA	T G p G	GG CA	AG AG In Se	GC A'	IC A. Le A:	AC sn	Pro CCC	825	÷

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	AAG Lys 260	Leu	GCG Ala	GGC	CTG Leu	ATT Ile 265	GGG Gly	CGG Arg	CAC His	GGG Gly	CCC Pro 270	CAG Gln	AAC Asn	AAG Lys	CAG Gln	CCC Pro 275	873
5	TTC Phe	ATG Het	GTG Val	GCT Ala	TTC Phe 280	TTC Phe	AAG Lys	GCC Ala	ACG Thr	GAG Glu 285	GTC Val	CAC His	TTC Phe	CGC	AGC Ser 290	ATC. Ile	921
10	CGG Arg	TCC Ser	ACG Thr	GGG Gly 295	AGC Ser	AAA Lys	CAG Gln	CGC Arg	AGC Ser 300	CAG Gln	AAC Asn	CGC Arg	TCC Ser	AAG Lys 305	ACG Thr	CCC Pro	969
15	AAG Lys	AAC Asn	CAG Gln 310	GAA Glu	GCC Ala	CTG Leu	CGG Arg	ATG Met 315	GCC Ala	AAC Asn	GTG Val	GCA Ala	GAG Glu 320	AAC Asn	AGC Ser	AGC Ser	1017
20	AGC Ser	GAC Asp 325	CAG Gln	AGG Arg	CAG Gln	GCC Ala	TGT Cys 330	AAG Lys	AAG Lys	CAC His	GAG Glu	CTG Leu 335	TAT Tyr	GTC Val	AGC Ser	TTC Phe	1065
20	CGA Arg 340	GAC Asp	CTG Leu	GGC Gly	TGG Trp	CAG Gln 345	GAC Asp	TGG Trp	ATC Ile	ATC Ile	GCG Ala 350	CCT Pro	GAA Glu	GGC Gly	TAC Tyr	GCC Ala 355	1113
25	GCC Ala	TAC Tyr	TAC Tyr	TGT Cys	GAG Glu 360	GGG Gly	GAG Glu	TGT Cys	GCC Ala	TTC Phe 365	CCT Pro	CTG Leu	AAC Asn	TCC Ser	TAC Tyr 370	ATG Met	1161
30	AAC Asn	GCC Ala	ACC Thr	AAC Asn 375	CAC His	GCC Ala	ATC Ile	GTG Val	CAG Gln 380	ACG Thr	CTG Leu	GTC Val	CAC His	TTC Phe 385	ATC Ile	AAC Asn	1209
35	CCG Pro	GAA Glu	ACG Thr 390	GTG Val	CCC Pro	AAG Lys	CCC Pro	TGC Cys 395	TGT Cys	GCG Ala	CCC Pro	ACG Thr	CAG Gln 400	CTC Leu	AAT Asn	GCC Ala	1257
	ATC Ile	TCC Ser 405	GTC Val	CTC Leu	TAC Tyr	TTC Phe	GAT Asp 410	GAC Asp	AGC Ser	TCC Ser	AAC Asn	GTC Val 415	ATC Ile	CTG Leu	AAG Lys	AAA Lys	1305
40	TAC Tyr 420	AGA Arg	AAC Asn	ATG Met	Val	GTC Val 425	CGG Arg	GCC Ala	TGT Cys	GGC Gly	TGC Cys 430	CAC His	TAGO	TCCI	CC		1351
45	GAGA	ATTC	AG A	CCCI	TTGG	G GC	CAAG	TTTT	TCI	GGAT	CCT	CCAI	TGCI	CG C	CTT	GCCAG	1411
	GAAC	CAGC	AG A	CCAA	CTGC	C TT	TTGÍ	GAGA	CCI	TCCC	CTC	CCTA	TCCC	CA A	CTTI	AAAGG	1471
	TGTG	AGAG	TA T	TAGG	AAAC	A TG	AGCA	GCAT	ATG	GCTT	TTG	ATCA	GTTI	TT C	AGTG	GCAGC	1531
50	ATCC	AATG	AA C	AAGA	TCCT	A CA	AGCT	GTGC	AGG	CAAA	ACC	TAGO	AGGA	AA A	AAAA	ACAAC	1591

WO 93/04692 GCATAAAGAA AAATGGCCGG GCCAGGTCAT TGGCTGGGAA GTCTCAGCCA TGCACGGACT 1651 CGTITCCAGA GGTAATTATG AGCGCCTACC AGCCAGGCCA CCCAGCCGTG GGAGGAAGGG 1711 5 GGCGTGGCAA GGGGTGGGCA CATTGGTGTC TGTGCGAAAG GAAAATTGAC CCGGAAGTTC 1771 1822 INFORMATION FOR SEQ ID NO:17: SEQUENCE CHARACTERISTICS: 10 (i) LENGTH: 431 amino acids (A) TYPE: amino acid TOPOLOGY: linear (D) (ii) MOLECULE TYPE: protein 15 (D) OTHER INFORMATION: /Product="0P1-PP" (ix) FEATURE: (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17: 20 Het His Val Arg Ser Leu Arg Ala Ala Ala Pro His Ser Phe Val Ala Leu Trp Ala Pro Leu Phe Leu Leu Arg Ser Ala Leu Ala Asp Phe Ser 25 Leu Asp Asn Glu Val His Ser Ser Phe Ile His Arg Arg Leu Arg Ser 30 Gln Glu Arg Arg Glu Het Gln Arg Glu Ile Leu Ser Ile Leu Gly Leu Pro His Arg Pro Arg Pro His Leu Gln Gly Lys His Asn Ser Ala Pro Met Phe Met Leu Asp Leu Tyr Asn Ala Het Ala Val Glu Glu Gly Gly Pro Gly Gly Gln Gly Phe Ser Tyr Pro Tyr Lys Ala Val Phe Ser Thr Gln Gly Pro Pro Leu Ala Ser Leu Gln Asp Ser His Phe Leu Thr 45 Asp Ala Asp Het Val Het Ser Phe Val Asn Leu Val Glu His Asp Lys 135 Glu Phe Phe His Pro Arg Tyr His His Arg Glu Phe Arg Phe Asp Leu 150

	Ser	Lys	Ile	Pro	Glu 165	Gly	Glu	Ala	Val	Thr 170	Ala	Ala	Glu	Phe	Arg 175	Ile
5	Tyr	Lys	Asp	Tyr 180	Ile	Arg	Glu	Arg	Phe 185	Asp	Asn	Glu	Thr	Phe 190	Arg	Ile
	Ser	Val	Tyr 195	Gln	Val	Leu	Gln	Glu 200	His	Leu	Gly	Arg	Glu 205	Ser	Asp	Leu
10	Phe	Leu 210	Leu	Asp	Ser	Arg	Thr 215	Leu	Trp	Ala	Ser	Glu 220	Glu	Gly	Trp	Leu
	Val 225	Phe	Asp	Ile	Thr	Ala 230	Thr	Ser	Asn	His	Trp 235	Val	Val	Asn	Pro	Arg 240
15	His	Asn	Leu	Gly	Leu 245	Gln	Leu	Ser	Val	Glu 250	Thr	Leu	Asp	Gly	Gln 255	Ser
20	Ile	Asn	Pro	Lys 260	Leu	Ala	Gly	Leu	Ile 265	Gly	Arg	His	Gly	Pro 270	Gln	Asn
	Lys	Gln	Pro 275	Phe	Met	Val	Ala	Phe 280	Phe	Lys	Ala	Thr	Glu 285	Val	His	Phe
25	Arg	Ser 290	Ile	Arg	Ser	Thr	Gly 295	Ser	Lys	Gln	Arg	Ser 300	Gln	Asn	Arg	Ser
	Lys 305	Thr	Pro	Lys	Asn	Gln 310	Glu	Ala	Leu	Arg	Met 315	Ala	Asn	Val	Ala	Glu 320
30	Asn	Ser	Ser	Ser	Asp 325	Gln	Arg	Gln	Ala	Cys 330	Lys	Lys	His	Glu	Leu 335	Tyr
35	Val	Ser		Arg 340	Asp	Leu	Gly	Trp	Gln 345	Asp	Trp	Ile	Ile	Ala 350	Pro	Glu
	Gly	Tyr	Ala 355	Ala	Tyr	Tyr	Cys	Glu 360	Gly	Glu	Cys	Ala	Phe 365	Pro	Leu	Asn
40	Ser	Tyr 370	Het	Asn	Ala	Thr	Asn 375	His	Ala	Ile	Val	Gln 380	Thr	Leu	Val	His
	Phe 385	Ile	Asn	Pro	Glu	Thr 390	Val	Pro	Lys	Pro	Cys 395	Cys	Ala	Pro	Thr	Gln 400
45	Leu	Asn	Ala	Ile	Ser 405	Val	Leu	Tyr	Phe	Asp 410	Asp	Ser	Ser	Asn	Val 415	Ile
50	Leu	Lys	Lys	Tyr 420	Arg	Asn	Het	Val	Val 425	Arg	Ala	Cys	Gly	Cys 430	His	

	(2) INFORMATION FOR SEQ ID NO:18:		
5	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1873 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear		∵
10	(ii) MOLECULE TYPE: cDNA (vi) ORIGINAL SOURCE: (A) ORGANISH: HURIDAE (F) TISSUE TYPE: EMBRYO		÷
15	(1x) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1041393 (D) OTHER INFORMATION: /note= "MOP1 (CDNA)"		
20	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18: CTGCAGCAAG TGACCTCGGG TCGTGGACCG CTGCCCTGCC	60	
25	CTGCAGCAAG TGACCTCGGG TCGTGCCCC GGATCGCGCC GCG ATG CAC GTG CGC CGGCGCGGGC CCGGTGCCCC GGATCGCGCG TAGAGCCGGC GCG ATG CAC GTG CGC Het His Val Arg	115	
	TCG CTG CGC GCT GCG GCG CCA CAC AGC TTC GTG GCG CTC TGG GCG CCT Ser Leu Arg Ala Ala Pro His Ser Phe Val Ala Leu Trp Ala Pro 10 10 20	163	
30	CTG TTC TTG CTG CGC TCC GCC CTG GCC GAT TTC AGC CTG GAC AAC GAG Leu Phe Leu Leu Arg Ser Ala Leu Ala Asp Phe Ser Leu Asp Asn Glu 25	211	
35	GTG CAC TCC AGC TTC ATC CAC CGG CGC CTC CGC AGC CAG GAG CGG CGG Val His Ser Ser Phe Ile His Arg Arg Leu Arg Ser Gln Glu Arg Arg 40 50	259	
40	GAG ATG CAG CGG GAG ATC CTG TCC ATC TTA GGG TTG CCC CAT CGC CCG Glu Het Gln Arg Glu Ile Leu Ser Ile Leu Gly Leu Pro His Arg Pro 60 65	307	
45	CGC CCG CAC CTC CAG GGA AAG CAT AAT TCG GCG CCC ATG TTC ATG TTG Arg Pro His Leu Gln Gly Lys His Asn Ser Ala Pro Het Phe Het Leu 75	355	
5.0	GAC CTG TAC AAC GCC ATG GCG GTG GAG GAG AGC GGG CCG GAC GGA CAG Asp Leu Tyr Asn Ala Het Ala Val Glu Glu Ser Gly Pro Asp Gly Gln 90 95	403	÷

300

295

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	 (2)	INF	ORMA:	TION	FOR	SEQ	ID N	0:19	:							
5			(i)	SEC (A) (B)) Li	CE CH ENGTH PE: OPOLO	: 43 amin	o ar	STIC aino aid ar	S: acid	ls	•	•			
			(ii)	HO	LECU!	LE TY	PE:	prot	cein							
10		(ix)	(D	•	THER			TION:					L-PP'	•	
			(xi)	SE	QUEN	CE DI	ESCR:	IPTI(ON: 9	SEQ	ID N	0:19	:			
15	1				,				Ala A							
	Leu	Trp	Ala	Pro 20	Leu	Phe	Leu	Leu	Arg 5	Ser	Ala	Leu .	Ala .	Asp 30	Phe	Ser
20			35	Glu					Phe							
25		50							G1u							
	65	His	Arg			, 0			Gln							
30	Het	Phe	. Het	Leu	Asp 85	Leu	Tyr	Asn	Ala	Het 90	Ala	Val	Glu	Glu	Ser 95	Gly
	Pro	Asp	Gly	Gln 100	Gly	Phe	Ser	Tyr	Pro 105	Tyr	Lys	Ala	Val	Phe 110	Ser	Thr
35	Glr	ı Gly	, Pro	Pro	Leu	Ala	Ser	Leu 120	Gln	Asp	Ser	His	Phe 125	Leu	Thr	Asp
40		13	p Me1	t Val			100									Glu
40	Pho	e Ph	e Hi			170	,									160
45	Ly	s Il			TO.	,										e Tyr 5
				יטסנ												e Thi
50) Va	1 Ty	r Gl	n Va	l Le	u Gli	n Gli	ц Ні 20	s Sei	r Gl	y Ar	g Gl	2.0	r Asi	p Le	u Phe

		210					213					220				,
5	225					230						Val				
•	Asn	Leu	G1y	Leu	Gln 245	Leu	Ser	Val	Glu	Thr 250	Leu	Asp	Gly	Gln	Ser 255	Ile
10	Asn	Pro	Lys	Leu 260	Ala	Gly	Leu	Ile	Gly 265	Arg	His	Gly	PTO	Gln 270	Asn	Lys
	Gln	Pro	Phe 275			Ala	Phe	Phe 280	Lys	Ala	Thr	Glu	Val 285	His	Leu	Arg
15	Ser	Ile 290		Ser	Thr	Gly	Gly 295	Lys	Gln	Arg	Ser	Gln 300	Asn	AIg	Ser	Lys
20		Pro	Lys	Asn	Gln	Glu 310	Ala	Leu	Arg	Het	Ala 315	Ser	Val	Ala	Glu	Asn 320
20	305 Ser	Ser	Ser	Asp	Gln 325	Arg	Gln	Ala	Cys	Lys 330	Lys	His	Glu	Leu	Tyr 335	Val
25	Ser	Phe	Arg	Asp 340		Gly	Trp	Gľn	Asp 345	Trp	Ile	Ile	Ala	Pro 350	Glu	Gly
	Tyr	Ala	Ala 355		Tyr	Cys	Glu	Gly 360	Glu	Cys	Ala	Phe	Pro 365	Leu	Asn	Ser
30	Tyr	Het	Asn	Ala	Thr	Asn	His 375	Ala	Ile	Val	Gln	Thr 380	Leu	Val	His	Phe
25			Pro	Asp	Thr	Val	Pro	Lys	Pro	Cys	Cys 395	Ala	Pro	Thr	Gln 400	Leu
35	385 Asn	Ala	Ile	Ser	Val 405	Leu	Tyr	Phe	. Asp	Asp 410	Ser	: Ser	Asn	Val	Ile 415	Leu
40	Lys	Lys	Туг	Arg 420			; Val	. Val	Arg 425	Ala	. Cys	Gly	Cys	His 430		
	(2)			ATIC	N FO											
45		t)	(<i>E</i>	7) I	E CH ENGI	H: J	LIZO	Dast	, ,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	Lrs	•					
))	~\ \ \	TRAN	MEDI	NESS:	: 5ÎI	Ja Te							
50		(:	Li)H(LEC	JLE I	TYPE	cDl	AK	٠							

	(vi)ORIGINAL SOURCE: (A) ORGANISH: Homo sapiens (F) TISSUE TYPE: HIPPOCAMPUS	
5	(ix)FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 4901696 (D) OTHER INFORMATION: /note= "hOP2 (cDNA)"	
10	(xi)SEQUENCE DESCRIPTION: SEQ ID NO:20:	60
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	CCCCTGGAGG GCTCCCTATG AGTGGCGGAG ACGGCCCAGG AGGCGCTGGA GCAACAGCTC	120
15	CCACACCGCA CCAAGCGGTG GCTGCAGGAG CTCGCCCATC GCCCCTGCGC TGCTCGGACC	180
	GCGGCCACAG CCGGACTGGC GGGTACGGCG GCGACAGAGG CATTGGCCGA GAGTCCCAGT	240
	CCGCAGAGTA GCCCCGGCCT CGAGGCGGTG GCGTCCCGGT CCTCTCCGTC CAGGAGCCAG	300
20	GACAGGTGTC GCGCGGCGGG GCTCCAGGGA CCGCGCCTGA GGCCGGCTGC CCGCCCGTCC	360
	CGCCCCGCCC CGCCCGCCGA GCCCAGCCTC CTTGCCGTCG GGGCGTCCCC	420
25	AGGCCCTGGG TCGGCCGCGG AGCCGATGCG CGCCCGCTGA GCGCCCCAGC TGAGCGCCCC	480
	CGGCCTGCC ATG ACC GCG CTC CCC GGC CCG CTC TGG CTC CTG GGC CTG Met Thr Ala Leu Pro Gly Pro Leu Trp Leu Leu Gly Leu	528
30	1 5	576
	GCG CTA TGC GCG CTG GGC GGG GGC CCC GGC CTG CGA CCC CCG CCC Ala Leu Cys Ala Leu Gly Gly Gly Pro Gly Leu Arg Pro Pro Pro 15	_
35	GGC TGT CCC CAG CGA CGT CTG GGC GCG CGC GAG CGC CGG GAC GTG CAG Gly Cys Pro Gln Arg Arg Leu Gly Ala Arg Glu Arg Arg Asp Val Gln 30 45	624
40	THE SEC CTC CTC CTC CTC CCC CCC CCC CCC CCC C	672
45	GCG CCA CCC GCC GCC TCC CGG CTG CCC GCG TCC GCG CCG C	720
50	CTG GAC CTG TAC CAC GCC ATG GCC GGC GAC GAC GAC GAC GAC GCG GCG Leu Asp Leu Tyr His Ala Het Ala Gly Asp Asp Asp Glu Asp Gly Ala 80	768

										- 1	20		•					
	wo 9	93/04	692													PCT/U	JS92/07358	
	Pro	Ala 95	GAG Glu	Arg	Arg	ren	100	Arg	VIG	rsh		105					816	
5	AAC Asn 110	ATG Het	GTG Val	GAG Glu	CGA Arg	GAC Asp 115	CGT Arg	GCC Ala	CTG Leu	GGC Gly	CAC His 120	CAG Gln	GAG Glu	CCC Pro	CAT His	TGG Trp 125	864	s
10	AAG Lys	GAG Glu	TTC Phe	CGC Arg	TTT Phe 130	GAC Asp	CTG Leu	ACC Thr	CAG Gln	ATC Ile 135	CCG Pro	GCT Ala	GGG Gly	GAG Glu	GCG Ala 140	GTC Val	912	Š
15	ACA Thr	GCT Ala	GCG Ala	GAG Glu 145	TIC Phe	CGG Arg	ATT Ile	TAL	AAG Lys 150	GTG Val	CCC Pro	AGC Ser	ATC Ile	CAC His 155	CTG Leu	CTC Leu	960	
	AAC Asn	AGG Arg	ACC Thr 160	CTC Leu	CAC His	GTC Val	AGC Ser	ATG Met 165	TTC Phe	CAG Gln	GTG Val	GTC Val	CAG Gln 170	GAG Glu	CAG Gln	TCC Ser	1008	
20	AAC Asn	AGG Arg 175	GAG Glu	TCT Ser	GAC Asp	TTG Leu	TTC Phe 180	TTT Phe	TTG Leu	GAT Asp	CTT Leu	CAG Gln 185	ACG Thr	CTC Leu	CGA Arg	GCT Ala	1056	
25	GGA Gly 190	GAC Asp	GAG Glu	GGC Gly	TGG Trp	CTG Leu 195	GTG Val	CTG Leu	GAT Asp	GTC Val	ACA Thr 200	GCA Ala	GCC Ala	AGT Ser	GAC Asp	TGC Cys 205	1104	
30	TGG Trp	TTG Leu	CTG Leu	AAG Lys	CGT Arg 210	CAC His	AAG Lys	GAC Asp	CTG Leu	GGA Gly 215	CTC	ÇGC Arg	CTC Leu	TAT Tyr	GIG Val 220	GAG Glu	1152	
35	ACT Thr	GAG Glu	GAC Asp	GGG Gly 225	CAC His	AGC Ser	GTG Val	GAT Asp	CCT Pro 230	GTA	CTG Leu	GCC Ala	GGC Gly	CTG Leu 235		GGT Gly	1200	
	CAA Gln	CGG Arg	GCC Ala 240	Pro	CGC Arg	TCC Ser	CAA Gln	CAG Gln 245	CCT Pro	TTC Phe	GTG Val	GTC Val	ACT Thr 250	TTC Phe	TTC Phe	AGG Arg	1248	
40	GCC Ala	AGT Ser 255	CCG Pro	AGT Ser	CCC	ATC Ile	CGC Arg 260	ACC Thr	CCT Pro	CGG Arg	GCA Ala	GTG Val 265	AGG Arg	CCA Pro	CTG Leu	AGG Arg	1296	
45	AGG Arg 270	AGG Arg	CAG Gln	CCG Pro	AAG Lys	AAA Lys 275	AGC Ser	AAC Asn	GAG Glu	CTG Leu	CCG Pro 280		GCC Ala	AAC Asn	CGA Arg	CTC Leu 285	1344	
50			ATC Ile	TTT Phe	GAT Asp 290	ASP	GTC Val	CAC His	GGC Gly	TCC Ser 295	****	GGC Gly	CGG Arg	CAG Gln	GTC Val 300	TGC Cys	1392	£

WO 93/04692 CGT CGG CAC GAG CTC TAC GTC AGC TTC CAG GAC CTC GGC TGG CTG GAC 1440 Arg Arg His Glu Leu Tyr Val Ser Phe Gln Asp Leu Gly Trp Leu Asp 305 TGG GTC ATC GCT CCC CAA GGC TAC TCG GCC TAT TAC TGT GAG GGG GAG 1488 Trp Val Ile Ala Pro Gin Gly Tyr Ser Ala Tyr Tyr Cys Glu Gly Glu 320 TGC TCC TTC CCA CTG GAC TCC TGC ATG AAT GCC ACC AAC CAC GCC ATC 1536 10 Cys Ser Phe Pro Leu Asp Ser Cys Het Asn Ala Thr Asn His Ala Ile 335 CTG CAG TCC CTG GTG CAC CTG ATG AAG CCA AAC GCA GTC CCC AAG GCG 1584 Leu Gln Ser Leu Val His Leu Het Lys Pro Asn Ala Val Pro Lys Ala 15 350 1632 TGC TGT GCA CCC ACC AAG CTG AGC GCC ACC TCT GTG CTC TAC TAT GAC Cys Cys Ala Pro Thr Lys Leu Ser Ala Thr Ser Val Leu Tyr Tyr Asp 370 AGC AGC AAC AAC GTC ATC CTG CGC AAA CAC CGC AAC ATG GTG GTC AAG 1680 Ser Ser Asn Asn Val Ile Leu Arg Lys His Arg Asn Met Val Val Lys 385 1723 25 GCC TGC GGC TGC CAC T GAGTCAGCCC GCCCAGCCCT ACTGCAG Ala Cys Gly Cys His 400 (2) INFORMATION FOR SEQ ID NO:21: 30 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 402 amino acids TYPE: amino acid (B) (D) TOPOLOGY: linear 35 (ii) MOLECULE TYPE: protein (ix) FEATURE: (A)OTHER INFORMATION: /product= "hOP2-PF" (xi)SEQUENCE DESCRIPTION: SEQ ID NO:21: 40 Met Thr Ala Leu Pro Gly Pro Leu Trp Leu Leu Gly Leu Ala Leu Cys 1 Ala Leu Gly Gly Gly Pro Gly Leu Arg Pro Pro Pro Gly Cys Pro 45 Gln Arg Arg Leu Gly Ala Arg Glu Arg Arg Asp Val Gln Arg Glu Ile 50

		รก			Gly		3.7			-									
5	65				Leu	7.0													æ
	Tyr	•			Ala 85														
10	Arg	Arg	Leu	Gly 100	Arg	Ala	Asp	Leu	Val 105	Het	Ser	Phe	Val	Asn 110	Het	Val			3
	Glu	Arg	Asp	Arg	Àla	Leu	Gly	His 120	Gln	Glu	Pro	His	Trp 125	Lys	Glu	Phe			
15		130			Thr		100											٠	
20	145	Phe			Tyr	130													
20	Геп	His	Val	Ser	Het 165	Phe	Gln	Val	Val	Gln 170	Glu	Gln	Ser	Asn	Arg 175	Glu		٠	
25	Ser	Asp	Leu	Phe 180	Phe	Leu	Asp	Leu	GIn 185	Thr	Leu	Arg	Ala	Gly 190	Asp	Glu		•	
	Gly	Trp	Leu 195	Val	Leu	Asp	Val	Thr 200	Ala	Ala	Ser	Asp	Cys 205	Trp	Leu	Leu			
30		210	His	Lys	: Asp		213	,									,		
35	225					200	,									Ala 240			
J J	Pro	Arg	g Ser	Glr	1 Gln 245	Pro	Phe	· Val	. Val	250	r Phe	e Phe	Arg	g Ala	255	Pro			
40				200	נ											g Gln			
			27:)					•							, Ile			٠
45		291	n'					-								g His			<u> </u>
50		ı Lei	u Ty	r Va	1 Se	r Pho 310	e Gl: O	n Asj	p. Le	u Gl	y Tr 31	p Le 5	u As	p Tr	p Va	1 Ile 320			₹

					Tyr 325												
5	Pro	Leu	Asp	Ser 340	Cys	Het	Asn	Ala	Thr 345	Asn	His	Ala	Ile	Leu 350	Gln	Ser	
			355		Het			300									
10		370			Ser		313										
	Asn 385	Val	Ile	Leu	Arg	Lys 390	His	Arg	Asn	Het	Val 395	Val	Lys	Ala	Cys	Gly 400	
15	Cys	His															
	(2)	IN	FORM		N FO												
20			ť)	´ (B)	LENG TYPE STRA	TH: : nu NDEC	1926 tcleż NESS	bas c ac	e pa :id :ngle	irs						
25			(ii	(D 10LEC	TOPO				•							
30			(⊽:	•	ORIGI (A) (F)	ORGA	NIS	RCE: 4: HI TYPE	JRIDA EM	AE BRYO							
35			(i:	•	FEATU (A) (B) (D)	NAMI	ጥቸርነ	Y: C N: 9 NFOR	ว 1	289 ON:	 /not	e= "	mOP2	cDN	A"		•
			(x	i)	SEQU	ENCE	DES	CRIP	TION	: SE	Q ID	NO:	22:			a. 66	50
			GC	CAGG	CACA	GGT	GCGC	CGT	CTGG	TCCT	CC C	CGTC	TGGC	G TC	AGCC	GAGC	104
40	CC	GACC	AGCT	ACC	AGTG	GAT	GCGC	GCCG	GC 1	GAAA	.GTCC	G AG	ATG Met		Met.	CGT Arg	204
45	CC	C G0	G CC y Pr	A CI	C TG	Ъ ге	A TI u Le 0	G GG u G]	C CI	TT GC	T CI	IG TO eu Cy L5	GC GC 75 A]	G CI La Le	G GG u Gl	A GGC y Gly 20	152
50	G() G]	-	C GC is Gl	T CO	CO AI	T CC	C CC	CG CA	AC AC		GT CO ys P: 30	CC CA	AG CO	GT CC	C CI	CG GGA eu Gly 15	. 200

	GCG Ala	CGC Arg	GAG Glu	CGC Arg 40	CGC Arg	GAC Asp	ATG Met	CAG Gln	CGT Arg 45	GAA Glu	ATC Ile	CTG Leu	GCG Ala	GTG Val 50	CTC Leu	GGG Gly	٠	248
5		Pro	Gly 55	Arg	Pro	Arg	Pro	60	VITO	GIII	110		65	-				296
10	Pro	Ala 70	Ser	Ala	Pro	Leu	75	net	rea	ASP.	Den	80						344
15	Asp 85	Asp	Asp	Asp	Gly	90 GTA	PTO	Pro	GIII	ATG	95	Ter	برحي		GCC Ala	100		392
	Leu	Val	Met	Ser	Phe 105	Val	ASN	net	var	110	wrg	vah			CTG Leu 115			440
20	Tyr	Gln	Glu	Pro 120	His	Trp	Lys	Gin	125	птэ	Inc	мор		130	CAG Gln	•		488
25		Ala	Gly 135	Glu	Ala	Val	Thr	140	ALA	GTIT	File	urg	145	-)-	-,-			536
30	Pro	Ser 150	Thr	His	Pro	Leu	155	Inr	Int	Leu	urs	160	DC.					584
35	Val 165	Val	Gln	Glu	His	Ser 170	ASN	Arg	GTIT	Ser	175	neu.			TTG Leu	180		632
	CTT Leu	CAG Gln	ACG Thr	CTC	CGA Arg 185	TCT Ser	GCG	GAC Asp	GAG Glu	GGC Gly 190	TLD	CTG	GTG Val	CTG Leu	GAC Asp 195	ATC Ile		680
40	ACA Thr	GCA Ala	Ala	Set	ÁSD	Arg	Trp	ren	CTG Leu 205	AAC Asn	CAT His	CAC His	AAG Lys	GAC Asp 210	CTG Leu	GGA Gly		728
45	CTC	CGC Arg	CTC Leu 215	TAT Tyr	GTG Val	GAA Glu	ACC Thr	GCG Ala 220	GAT Asp	GGG Gly	CAC His	AGC Ser	ATG Het 225	GAT Asp	CCT Pro	GGC Gly		776
50	CTG Leu	GCT Ala 230	GGT Gly	CTG Leu	CTT Leu	GGA Gly	CGA Arg 235	CAA Gln	GCA Ala	CCA Pro	CGC Arg	TCC Ser 240	AGA Arg	CAG Gln	CCT	TTC Phe		824

1669

CTCAGCCCAC AATGGCAAAT TCTGGATGGT CTAAGAAGGC CGTGGAATTC TAAACTAGAT

50

	Lys	Glu	Pro	Ser 150	Thr	His	Pro	Leu	Asn 155	Thr	Thr	Leu	His	Ile 160	Ser	Het
5	Phe	Glu	Val 165	Val	Gln	Glu	His	Ser 170	Asn	Arg	Glu	Ser	Asp 175	Leu	Phe	Phe
		180				Leu	107					-				
10	195					Ser 200										
					212	Tyr										
15	Pro	Gly	Leu	Ala 230	Gly	Leu	Leu	Gly	Arg 235	Gln	Ala	Pro	Arg	Ser 240	Arg	Gln
20	Pro	Phe	Het 245	Val	Thr	Phe	Phe	Arg 250	Ala	Ser	Gln	Ser	Pro 255	Val	Arg	Ala
20	Pro	Arg 260		Ala	Arg	Pro	Leu 265	Lys	Arg	Arg	Gln	Pro 270	Lys	Lys	Thr	Asn
25	Glu 275	Leu		His	Pro	Asn 280	Lys	Leu	Pro	Gly	Ile 285	Phe	Asp	Asp	Gly	His 290
	Gly	Ser	Arg	g Gly	7 Arg	Glu	. Val	. Cys	Arg	Arg 300	His	Glu	Leu	Tyr	Val 305	Ser
30	Phe	Arg	g As	Let 310	ı G13	7 Trp	Leu	ı Asp	Trp	Val	. Ile	Ala	Pro	Glr 320	Gly	Tyr
35	Sei	: Ala	1 Ty:	r Ty:	r Cys	s Glu	ı Gly	7 Gli 330	ı Cys	. Ala	. Phe	e Pro	335	ı Ası	Set	Cys
J.J	Het	: Ası 340	n Al	a Th	r As	n Hi	s Ala 34!	a Ile 5	e Lei	ı Glı	ı Sei	T Let 350	va:	L His	s Let	1 Met.
40	Ly:	s Pr		p Va	l Va	1 Pro	o Ly :	s Al	а Су	s Cy	s Ala 36	a Pro	o Th	r Ly	s Le	370
	Ala	a Th	r Se	r Va	1 Le 37	и Ту 5	r Ty	r As	p Se	r Se 38	r As O	n As	n Va	1 11	e Le	u Arg 5
45	Ly	s Hi	s AI	g As	n Me	t Va	l Va	l Ly	s Al 39	а Су 5	s Gl	у Су	s Hi	S		

	(2) INFORM	ATION FOR SEQ ID NO:24:	
5	(1) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1368 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(ii	.) HOLECULE TYPE: cDNA	ž
10	(ix	(A) NAME/KEY: CDS (B) LOCATION: 11368 (D) OTHER INFORMATION:/STANDARD NAME="60A"	
15	(2	PUBLICATION INFORMATION: (A) AUTHORS: WHARTON, KRISTI A.; THOMSEN, GERALD H.; GELBERT, WILLIAM H. (B) TITLE: DROSOPHILA 60A GENE (B) TITLE: DROSOPHILA 60A GENE	
20		(B) TITLE: DROSUPHILA GOA GENERAL CONTINUES OF THE CONTIN	
25	ſx	SEQUENCE DESCRIPTION: SEQ ID NO:24:	n
	ATG TCG GG Het Ser Gl	A CTG CGA AAC ACC TCG GAG GCC GTT GCA GTG CTC GCC TCC y Leu Arg Asn Thr Ser Glu Ala Val Ala Val Leu Ala Ser 10	
30	Leu Gly Le	C GGA ATG GTT CTG CTC ATG TTC GTG GCG ACC ACG CCG CCG 9 Su Gly Het Val Leu Het Phe Val Ala Thr Thr Pro Pro 20 25	
35	Ala Val G	AG GCC ACC CAG TCG GGG ATT TAC ATA GAC AAC GGC AAG GAC LU Ala Thr Gln Ser Gly Ile Tyr Ile Asp Asn Gly Lys Asp 40 40	
40	Gln Inr 1.	TC ATG CAC AGA GTG CTG AGC GAG GAC GAC AAG CTG GAC GTC 19 10 ATG CAC AGA GTG CTG AGC GAG GAC GAC AAG CTG GAC GTC 19 10 ATG CAC AGA GTG CTG AGC GAG GAC AAG CTG GAC GTC 19 10 ATG CAC AGA GTG CTG AGC GAG GAC AAG CTG GAC GTC 19 10 ATG CAC AGA GTG CTG AGC GAG GAC AAG CTG GAC GTC 19 10 ATG CAC AGA GTG CTG AGC GAG GAC AAG CTG GAC GTC 19 10 ATG CAC AGA GTG CTG AGC GAC GAC AAG CTG GAC GTC 19 10 ATG CAC AGA GTG CTG AGC GAG GAC GAC AAG CTG GAC GTC 19 10 ATG CAC AGA GTG CTG AGC GAG GAC GAC AAG CTG GAC GTC 19 10 ATG CAC AGA GTG CTG AGC GAG GAC GAC AAG CTG GAC GTC 19 10 ATG CAC AGA GTG CTG AGC GAC GAC AAG CTG GAC GTC 19 10 ATG CAC AGA GTG CTG AGC GAC AAG CTG GAC GTC 19 10 ATG CAC AGA GTG CTG AGC GAC GAC AAG CTG GAC GTC 19 10 ATG CAC AGA GTG CTG AGC GAC AAG CTG GAC GTC 19 10 ATG CAC AGA GTG CTG AGC GAC AAG CTG GAC AAG CTG GAC GTC 19 10 ATG CAC AGA GTG CTG AGC GAC AAG CTG AAG CTG AAG CTG AAG AAG CTG AAG AAG CTG AAG AAG CTG AAG AAG AAG CTG AAG AAG AAG AAG CTG AAG AAG AAG AAG AAG AAG AAG AAG AAG A	
45	TCG TAC G	AG ATC CTC GAG TTC CTG GGC ATC GCC GAA CGG CCG ACC GAG CGG CCG ACC GAG GAG	
50	CTG AGC A Leu Ser S	GC CAC CAG TTG TCG CTG AGG AAG TCG GCT CCC AAG TTC CTG 2 GET His Gln Leu Ser Leu Arg Lys Ser Ala Pro Lys Phe Leu 95 96	88

1	Leu .	Asp	Val	100	UTP	CGC Arg			105					TIO				336
	Asp	Glu	ASP	GAC Asp	vah	TAC Tyr		120	-				125					384
10	GAC Asp	CTC Leu 130		GAG Glu	GAT Asp	GAG Glu	GGC Gly 135	GAG Glu	CAG Gln	CAG Gln	AAG Lys	AAC Asn 140	TTC	ATO	AC Th	C C	AC Asp	432
	Leu		AAG Lys	CGG Arg	GCC Ala	ATC Ile 150	GAC Asp	GAG Glu	AGC Ser	GAC Asp	ATC Ile 155	ATC	ATG Het	ACC Th	TI Pb	C (CTG Leu 160	480
15	AAC Asn	AAG Lys	CGC	CAC	CAC His	AAT Asn	GTG Val	GAC Asp	GAA Glu	CTG Leu 170	CGI	CAC His	GAC Glu	CAC Hi:	G G(s G)	C Ly 75	CGT Arg	528
20	CGC Arg	CTG Leu	TGG Trp	TT0	GAC Asp	GTC Val	TCC	AAC Asr	GTG Val 185	CCC Pro	AA(GA(AS)	AA(TA Ty 19	C C T L	TG eu	GTG Val	576
25	ATG Net	GCC	GAC Glu 195	CT(ATC	TAT	CA(Gl)	G AAC n Asi	GCC A Ala	AA(C GA	G GG u G1 20	C AA y Ly 5	G T s T	GG rp	CTG Leu	624
30	ACC Thi	GC(C AA(G GAG	G TTO	ACC Th: 21		C ACC	G GT. r Va	A TA 1 Ty	C GC r Al 22	C AT a Il	T GO e Gl	C A y T	.cc :hr	GGC Gly	672
25	Thi	CT Le		C CA y Gl	G CA n Hi	C ACC s Th		G GA t Gl	G CC	G CI o Le	G TC u Se 23	C TC r Se	G GI	G A	AC A	CC	ACC Thr 240	720
35			C TA p Ty	C GI	G GG 1 G1 24	y II	G TT p Le	G GA	G CI lu Le	C AA	C GI	G AC	CC GA	AG G Lu G	GC (CTG Leu 255	CAC	768
40	GA G1	G TO	G CI	eu va			G AA	G G/	AC AA	T CA	T G(GC A' Ly I	TC T	AC A yr I 2	TT le 70	GGA Gly	GCA Ala	816
45	CA Hi	.C GC	La Va	C A	•	GA CC	C GA	AC CO	GC GA rg G:	AG G' Lu V	IG A al L	AG C ys L	TG G eu A 2	AC G sp A 85	AC Sp	ATI Ile	GGA Gly	864
50	CI Le	u I	_	75 AC C is A	GC A.	AG G	37 **	AC G sp A 95	AC G. sp G	AG T lu P	TC C he G	AG C	CC Tro P	TC A	ATG let	AT(C GGC e Gly	912

- 130 - PCT/US92/07358														1592/07358			
W	O 93	/0469	92						•							-C170	372/0/000
	TTC Phe	TTC Phe	CGC Arg	GIY	PLU	310	CTG Leu		- •		315					320	960
5	•	AGG Arg	AGC Ser	AAG Lys	CGA Arg 325	AGC Ser	GCC Ala	AGC Ser	CAT His	CCA Pro 330	CGC Arg	AAG Lys	CGC Arg	AAG Lys	AAG Lys 335	TCG Ser	1008
10	GTG Val	TCG Ser	CCC Pro	AAC Asn 340		GTG Val	CCG Pro	CTG Leu	CTG Leu 345	_	CCG Pro	ATG Het	GAG Glu	AGC Ser 350	ACG Thr	CGC Arg	1056
15	AGC Ser	TGC Cys	CAG Gln 355	_	CAG Gln	ACC Thr	CTG Leu	TAC Tyr 360	ATA Ile	GAC Asp	TTC Phe	AAG Lys	GAT Asp 365	CTG Leu	GGC Gly	TGG Trp	1104
15	CAT His	GAC Asp 370	TGG Trp	ATC Ile	ATC Ile	GCA Ala	CCA Pro 375	GAG Glu	GGC Gly	TAT Tyr	GGC Gly	GCC Ala 380	TTC Phe	TAC Tyr	TGC Cys	AGC Ser	1152
20	Gly	GAG Glu		AAT Asn	TTC Phe	CCG Pro 390	Tien	AAT Asn	GCG Ala	CAC His	ATG Het 395	AAC	GCC	ACG Thr	AAC Asn	CAT His 400	1200
25	385 GCG Ala		GTC Val	ÇAG Gln	ACC Thr 405	CIG		CAC His	CTG	CTG Leu 410		CCC	AAG Lys	AAG Lys	GTG Val 415	CCC	1248
30	AAG Lys	CCC	TGC Cys	TGC Cys 420	GCT Ala		ACC	AGG Afg	CTG Leu 425	GGA Gly	GCA Ala	CTA Leu	CCC Pro	GTI Val 430	CTG Leu	TAC	1296
	CAC His	CTG	l Asn	GÁC Asp		AAT Asn	GTG Val	AAC Asn 440		AAA Lys	AAG Lys	TAI Tyi	AGA ATS	AAC ASI	ATG Het	ATT Ile	1344
35	GIG Val	Lys	: Sei	TGC	GGG Gly	TGC Cys	CAT His	,	L								1368
40	(2)	450 IN					EQ II) NO:									
45			(:	1	(A) I	ENG	CHAICH: 4 ami LOGY:	ino a	amino acid	o ac.	: ids						
			(i:	_,			TYPI										
50	•		(x:	i)	SEQUI	ENCE	DES	CRIP'	TION	: SE	Q ID	NO:	25:				

	Ket					_															
_	1 Leu	Gly	L	eu (Gly 20	Het	Val	Leu	L	eu 1	let 25	Phe	۷a	al A	la :	Thr	Thi	P:	ro :	Pr	0
5				3.5	Ala		Gln			. •											
10		51	1				Arg		•												
		Ту	r G				Glu 70														
15	Leu					0.															
20					TOO		Arg														
20				113			ту														
25		- 13	:u	G1u			p Gl		_												
	14'	1 A	5 p				a Il 15	•													
30	Ası	n L				TO															
35	•				10	U	p Va														
J	Мe			14	٦.		g I														
40		- 7	าก	As	n AI		Lu Pl	_													
	2.2	ır I	Leu	G1;																	
4	5 G1	Ly A					ly T 45														
5	G: 0	lu '	Irp) Le	u V	al L 60	ys S	er :	Lys	. As	р А 2	sn 1 65	His	Gl	y I]	le T	Уľ	11e 270	G1	.у	Ala

			210		Arg											
5		290			Lys		433									
	305				Pro	210					-		•			
10	His				Arg 325					•••						
				340	Asn				373	•						
15			355		Gln			500								
20		370			Ile		313									
	385				Phe	330			•		_					
25	Ala	Ile			405											Pro
				420	ļ			•								Tyr
30	His	Leu	Asn 435	Asp	Glu	Asn	Val	Asn 440	Leu	Lys	Lys	Туг	AI9	Asn	. Het	Ile
35	Val	Lys 450	Ser	Cys	Gly	Cys	His 455						•			
	(2)	INF	ORMA	TION	FOR	SEC	ID	NO:2	6:							
40 ⁻			(i	(EQUE (A) I (B) I (D) I	ENGT	CHAR H: a ami .OGY:	mino no a	aci cid	CICS:	•					
			(ii	i) 1	OLEC	ULE	TYPE	2: p1	cote	Ln:						
45			(iii	L) (ORIGI (A) (NAL RGAI	SOUI HSI:	RCE: Hor	no Sa	apie	ns					
50			(i)	-,	FEAT((A) 1 (B) 1 (D) (IAME.					note	="BH	P3"			

	(x1) SEQUENCE DESCRIPTION: SEQ ID NO:26:	
5	(i)SEQUENCE CHARACTERISTICS: (A) LENGTH: 104 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
10	(ii)MOLECULE TYPE: protein (ix)FEATURE: (A) NAME/KEY: Protein (B) LOCATION: 1104 (D) OTHER INFORMATION: /note="BMP3"	
15	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:	
	Cys Ala Arg Arg Tyr Leu Lys Val Asp Phe Ala Asp Ile Gly Trp Ser	
20	Glu Trp Ile Ile Ser Pro Lys Ser Phe Asp Ala Tyr Try Cys Ser Gly 20 25 30	
25	Ala Cys Gln Phe Pro Met Pro Lys Ser Leu Lys Pro Ser Asn His Ala 35	
2.3	Thr Ile Gln Ser Ile Val Ala Arg Ala Val Gly Val Val Pro Gly Ile 50 55	
30	Pro Glu Pro Cys Cys Val Pro Glu Lys Het Ser Ser Leu Ser Ile Leu 65 70 75 80	
	Phe Phe Asp Glu Asn Lys Asn Val Val Leu Lys Val Tyr Pro Asn Her 85 90	t
35	Thr Val Glu Ser Cys Ala Cys Arg 100	
40	(2) INFORMATION FOR SEQ ID NO:27:	
45	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 102 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	•
	(ii) HOLECULE TYPE: protein	
50	(vi) ORIGINAL SOURCE: (A) ORGANISH: HOMO SAPIENS	

			(ix)	(A		HE/F	EY: ION: INFO			/nc	te=	"BME	·5"	•	œ	
5			(xi)	SE	OUE	ICE I	ESCE	tPT]	ON:	SEQ	ID i	10:27	7:			
	•		Lys	His	Glu 5	Leu	Tyr	Val	Ser	Phe 10	Arg	Asp	Leu			
10				2.0			Glu									
15			3.5	Phe			Asn									
15		50	Gln				His 55									
20	25					, •	Lys									
	Asp	Asp	Ser	Ser	Asn 85	. Val	Ile	Leu	Lys	Lys 90	Tyr	Arg	Asn	Het	Val 95	Val
25	Arg	Ser	Cys	Gly 100	Cys	His	;									
20	(2)	INF	ORHA				[D						·			
30	٠		(i	, (A) I	ENG	CHAICH: I	102 a	mino icid	açı	i ids		•			
35				((D) :	[OPO]	LOGY:	111	near							
			(ii	- ,							•					-
40			(⊅:	i) ((A)	ORGA	NISH SOUI	: HO	HO S	APIE	NS			•		
45			(i :	,	(B) (D)	NAHE LOCA OTHE	/KEY TION R IN	: 1. FORM	ATIO	N: /						
			(x	i)	SEQU	ENCE	DES	CRIP	TION	: SE	Q II	NO:	28:		-	_ CT=
EΩ	Cy 1	s Ar	g Ly	s Hi	s G1 5	u Le	u Ty	r Va	1 Se	r Ph	e Gl	n As	p Le	u Gl	y 11 - 15	p Gln

PCT/US92/07358 WO 93/04692 Asp Trp Ile Ile Ala Pro Lys Gly Tyr Ala Ala Asn Tyr Cys Asp Gly 25 Glu Cys Ser Phe Pro Leu Asn Ala His Met Asn Ala Thr Asn His Ala Ile Val Gln Thr Leu Val His Leu Met Asn Pro Glu Tyr Val Pro Lys 10 Pro Cys Cys Ala Pro Thr Lys Leu Asn Ala Ile Ser Val Leu Tyr Phe Asp Asp Asn Ser Asn Val Ile Leu Lys Lys Tyr Arg Trp Het Val Val 15 Arg Ala Cys Gly Cys His (2) INFORMATION FOR SEQ ID NO:29: 20 SEQUENCE CHARACTERISTICS: (A) LENGTH: 102 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear HOLECULE TYPE: protein 25 (ii) FEATURE: (ix) (A) NAME/KEY: Protein (B) LOCATION: 1..102 (D) OTHER INFORMATION: /label= OPX /note= "WHEREIN XAA AT EACH POS'N IS INDEPENDENTLY 30 SELECTED FROM THE RESIDUES OCCURRING AT THE CORRESPONDING POS'N IN THE C-TERMINAL SEQUENCE OF HOUSE OR HUMAN OP1 OR OP2 (SEE SEQ. ID NOS. 5,6,7 and 8 or 16,18,20 and 22.)" 35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

Cys Xaa Xaa His Glu Leu Tyr Val Xaa Phe Xaa Asp Leu Gly Trp Xaa

Asp Trp Naa Ile Ala Pro Naa Gly Tyr Naa Ala Tyr Tyr Cys Glu Gly
25 30

Glu Cys Xaa Phe Pro Leu Xaa Ser Xaa Het Asn Ala Thr Asn His Ala

Ile Xaa Gln Xaa Leu Val His Xaa Xaa Xaa Pro Xaa Xaa Val Pro Lys 50

```
Yaa Cys Cys Ala Pro Thr Xaa Leu Xaa Ala Xaa Ser Val Leu Tyr Xaa
   Asp Xaa Ser Xaa Asn Val Xaa Leu Xaa Lys Xaa Arg Asn Het Val Val
5
   Kaa Ala Cys Gly Cys His
               100
        INFORMATION FOR SEQ ID NO:30:
10 (2)
        (i) SEQUENCE CHARACTERISTICS:
            (A) LENGTH: 97 amino acids
            (B) TYPE: amino acids (C) TOPOLOGY: linear
         (ii) MOLECULE TYPE: protein
15
         (ix) FEATURE:
             (A) NAME: Generic Sequence 5
                 OTHER INFORMATION: wherein each Xaa is independently
                  selected from a group of one or more specified amino acids as
             (D)
                  defined in the specification.
20
         (xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:
         Leu Xaa Xaa Xaa Phe
25
         Xaa Xaa Xaa Gly Trp Xaa Xaa Trp Xaa
                         10
         Xaa Xaa Pro Xaa Xaa Xaa Ala
                          20
         Xaa Tyr Cys Xaa Gly Xaa Cys Xaa
 30
                             30
               25
          Xaa Pro Xaa Xaa Xaa Xaa
                          35
          Xaa Xaa Xaa Asn His Ala Xaa Xaa
 35
                                  45
               40
          Xaa Xaa Xaa Xaa Xaa Xaa Xaa
                          50
          Xaa Xaa Xaa Xaa Xaa Xaa Cys
               55
          Cys Xaa Pro Xaa Xaa Xaa Xaa Xaa
 40
                 65
          Xaa Xaa Xaa Leu Xaa Xaa Xaa
                             75
           Xaa Xaa Xaa Xaa Val Xaa Leu Xaa
  45
                      80
           Xaa Xaa Xaa Xaa Het Xaa Val Xaa
           85
           Xaa Cys Xaa Cys Xaa
                95
  50
```

(ii)

```
INFORMATION FOR SEQ ID NO:31:
        (1) SEQUENCE CHARACTERISTICS:
                 LENGTH: 102 amino acids
                 TYPE: amino acids
5
                 TOPOLOGY: linear
            (C)
        (ii) MOLECULE TYPE: protein
        (ix) FEATURE:
            (A) NAME: Generic Sequence 6
            (D) OTHER INFORMATION: wherein each Xaa is independently
                 selected from a group of one or more specified amino acids as
10
                 defined in the specification.
       (xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:
15
       Cys Xaa Xaa Xaa Leu Xaa Xaa Phe
       Xaa Xaa Xaa Gly Trp Xaa Xaa Trp Xaa
       Xaa Xaa Pro Xaa Xaa Xaa Ala
20
                             25
        20
        Xaa Tyr Cys Xaa Gly Xaa Cys Xaa
                 30
        Xaa Pro Xaa Xaa Xaa Xaa Xaa
                         40
25
        Xaa Xaa Xaa Asn His Ala Xaa Xaa
                 45
        Xaa Xaa Xaa Xaa Xaa Xaa Xaa
                         55
        Xaa Xaa Xaa Xaa Xaa Xaa Cys
30
             60
        Cys Xaa Pro Xaa Xaa Xaa Xaa Xaa
                     70
        Xaa Xaa Xaa Leu Xaa Xaa Xaa
                             80
         75
35
        Xaa Xaa Xaa Val Xaa Leu Xaa
                     85
        Xaa Xaa Xaa Xaa Het Xaa Val Xaa
                             95
         90
        Xaa Cys Xaa Cys Xaa
40
                100
           INFORMATION FOR SEQ ID NO:32:
    (2)
                 SEQUENCE CHARACTERISTICS:
            (i)
                 LENGTH: 1238 base pairs, 372 amino acids
 45
            (A)
                 TYPE: nucleic acid, amino acid
            (B)
                 STRANDEDNESS: single
            (C)
                 TOPOLOGY: linear
            (D)
 50
                 MOLECULE TYPE: cDNA
```

	(111) ORIGINAL SOURCE: (A) ORGANISH: human (F) TISSUE TYPE: BRAIN														
5	(iv) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: (D) OTHER INFORMATION: /product= "GDF-1"														
10	/note= "GDF-1 CDNA"														
15	(D) VOLUME: 88 (E) RELEVANT RESIDUES: 1-1238 (F) PAGES: 4250-4254 (G) DATE: Hay-1991 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32: CCCGACACCG GCCCCGCCCT CAGCCCACTG GTCCCGGGCC GCCGCGGACC CTGCGCACTC														
20	(xi) SEQUENCE DESCRIPTION: SEQ ID NO.32.	60													
	GGGGACACCG GCCCCGCCCT CAGCCCACTG GTCCCGGGGCC GCCGCGGACC CTGCGCACTC	113													
25	GGGGACACCG GCCCCGCCCT CAGCCCACTG GTCCCGGGCC GCCGCGGACC CTGCGCACTC TCTGGTCATC GCCTGGGAGG AAG ATG CCA CCG CCG CAG CAA GGT CCC TGC GGC Het Pro Pro Pro Gln Gln Gly Pro Cys Gly 10 10														
	CAC CAC CTC CTC CTC CTG GCC CTG CTG CCC TCG CTG CCC His His Leu Leu Leu Leu Ala Leu Leu Pro Ser Leu Pro 15 20 25	158													
30	CTG ACC CGC GCC CCC GTG CCC CCA GGC CCA GCC GCC GCC CTG CTC Leu Thr Arg Ala Pro Val Pro Pro Gly Pro Ala Ala Ala Leu Leu 30 35	203													
35	CAG GCT CTA GGA CTG CGC GAT GAG CCC CAG GGT GCC CCC AGG CTC GIn Ala Leu Gly Leu Arg Asp Glu Pro Gln Gly Ala Pro Arg Leu 45 50	· 248													
40	CGG CCG GTT CCC CCG GTC ATG TGG CGC CTG TTT CGA CGC CGG GAC ATG Pro Val Pro Val Het Trp Arg Leu Phe Arg Arg Arg Ap 65	293													
45	CCC CAG GAG ACC AGG TCT GGC TCG CGG CGG ACG TCC CCA GGG GTC Pro Gln Glu Thr Arg Ser Gly Ser Arg Arg Thr Ser Pro Gly Val 85	338													
50	ACC CTG CAA CCG TGC CAC GTG GAG GAG CTG GGG GTC GCC GGA AAC Thr Leu Gln Pro Cyc His Val Glu Glu Leu Gly Val Ala Gly Asn 90 90 95	383													

	- 100
	ATC GTG CGC CAC ATC CCG GAC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CAC ATC CCG GAC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CAC ATC CCG GAC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CAC ATC CCG GAC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CAC ATC CCG GAC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CAC ATC CCG GAC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CAC ATC CCG GAC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CAC ATC CCG GAC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CAC ATC CCG GAC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CAC ATC CGG GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CCC ACC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CCC ACC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CCC ACC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CCC ACC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CCC ACC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CCC ACC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CCC ACC CGC GGT GCG CCC ACC CGG GCC TCG ATC GTG CGC CCC ACC CGC GGT GCG CCC ACC CGC GCC TCG GCC CCC ACC CGC GCC ACC CGC GCC TCG GCC CCC ACC CCC ACC CCC ACC CGC GCC TCG GCC CCC ACC ACC CCC ACC ACC CCC ACC A
5	GAG CCT GTC TCG GCC GCG GGG CAT TGC CCT GAG TGG ACA GTC GTC Glu Pro Val Ser Ala Ala Gly His Cys Pro Glu Trp Thr Val Val 120 125 473
10	TTC GAC CTG TCG GCT GTG GAA CCC GCT GAG CGC CCG AGC CGG GCC TTC GAC CTG TCG GCT GTG GAA CCC GCT GAG CGC CCG AGC CGG GCC Phe Asp Leu Ser Ala Val Glu Pro Ala Glu Arg Pro Ser Arg Ala 145
	CGC CTG GAG CTG CGT TTC GCG GCG GCG GCG GCG GCA GCC CCG GAG Arg Leu Glu Leu Arg Phe Ala Ala Ala Ala Ala Ala Ala Pro Glu 150 150 150
15	GGC GGC TGG GAG CTG AGC GTG GCG CAA GCG GGC CAG GGC GGC 608 Gly Gly Trp Glu Leu Ser Val Ala Gln Ala Gly Gln Gly Ala Gly 175 165
20	GCG GAC CCC GGG CCG GTG CTC CGC CAG TTG GTG CCC GCC CTG 653 Ala Asp Pro Gly Pro Val Leu Leu Arg Gln Leu Val Pro Ala Leu 190 180
25	GGG CCG CCA GTG CGC GCG GAG CTG CTG GGC GCC GCT TGG GCT CGC 698 Gly Pro Pro Val Arg Ala Glu Leu Leu Gly Ala Ala Trp Ala Arg
	195
30	AAC GCC TCA TGG CCG CGC AGC CTC CGC CTG GCG CTG GCG CTA CGC ASn Ala Ser Trp Pro Arg Ser Leu Arg Leu Ala Leu Ala Leu Arg 210 220
35	CCC CGG GCC CCT GCC GCC TGC GCG CGC CTG GCC GAG GCC TCG CTG Pro Arg Ala Pro Ala Ala Cys Ala Arg Leu Ala Glu Ala Ser Leu 235 225
	CTG CTG GTG ACC CTC GAC CCG CGC CTG TGC CAC CCC CTG GCC CGG 833 Leu Leu Val Thr Leu Asp Pro Arg Leu Cys His Pro Leu Ala Arg 250 240
40	CCG CGG CGC GAC GCC GAA CCC GTG TTG GGC GGC GGC CCC GGG GGC Pro Arg Arg Asp Ala Glu Pro Val Leu Gly Gly Pro Gly Gly 265
45	GCT TGT CGC GCG CGG CGG CTG TAC GTG AGC TTC CGC CAG GTG GGC 923 Ala Cys Arg Ala Arg Arg Leu Tyr Val Ser Phe Arg Glu Val Gly 270 280
50	TGG CAC CGC TGG GTC ATC GCG CCG CGC CCC TTC CTG GCC AAC TAC 968 Trp His Arg Trp Val Ile Arg Pro Arg Gly Phe Leu Ala Asn Tyr 290 295

WO 93/04692 TGC CAG GGT CAG TGC GCG CTG CCC GTC GCG CTG TCG GGG TCC GGG Cys Gln Gly Gln Cys Ala Leu Pro Val Ala Leu Ser Gly Ser Gly 300 GGG CCG CCG GCG CTC AAC CAC GCT GTG CTG CGC GCG CTC ATG CAC 1058 Gly Pro Pro Ala Leu Asn His Ala Val Leu Arg Ala Leu Met His 5 315 GCG GCC GCC GGA GCC GCC GAC CTG CCC TGC TGC GTG CCC GCG 1103 Ala Ala Ala Pro Gly Ala Ala Asp Leu Pro Cys Cys Val Pro Ala 10 330 CGC CTG TCG CCC ATC TCC GTG CTC TTC TTT GAC AAC AGC GAC AAC 1148 Arg Leu Ser Pro Ile Ser Val Leu Phe Phe Asp Asn Ser Asp Asn 345 GTG GTG CTG CGG CAG TAT GAG GAC ATG GTG GTG GAC GAG TGC GGC 1193 15 Val Val Leu Arg Gln Tyr Glu Asp Het Val Val Asp Glu Cys Gly 360 TGC CGC TAACCCGGGG CGGGCAGGGA CCCGGGCCCA ACAATAAATG CCGCGTGG 1238 20 Cys Arg 372 INFORMATION FOR SEQ ID NO:33: 25 (2) SEQUENCE CHARACTERISTICS: LENGTH: 372 amino acids (A) TYPE: amino acid (B) STRANDEDNESS: single (C) TOPOLOGY: linear 30 (ii) HOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO 35 (iv) ANTI-SENSE: NO ORIGINAL SOURCE: (vi) (A) ORGANISH: human TISSUE TYPE: BRAIN 40 (F) (ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: (D) OTHER INFORMATION: /function= 45 /product= "GDF-1" (x1) SEQUENCE DESCRIPTION: SEQ ID NO:33: Het Pro Pro Pro Glm Glm Gly Pro Cys Gly 50

	His				13											
5					Pro '											
•					Leu 45											
10					Pro 60											
					Arg 75											
15					Cyc 90					-						
20					Ile 105											
					Ala 120											
25					123										Ala 145	
					130										160	
30 .					1.00						-				175	
35	Ala	. As	Pro	Gly	7 Pro 180	Va]	Lei	ı Lei	ı Ar	g Glr 185	ı Let	ı Va	l Pro	o Ala	190	
					193)					•				a Arg 205	
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10	Gly	Pro	Pro	Ala	Leu 315	Asn	His	Ala	Val	Leu 320	Arg	.Ala	Leu	Ket	His 325
	Ala	Ala	Ala	Pro	Gly 330	Ala	Ala	Asp	Leu	Pro 335	Cys	Cys	Val	Pro	Ala 340
15	Arg	Leu	Ser	Pro	Ile 345	Ser	Val	Leu	Phe	Phe 350	Asp	Asn	Ser	Asp	Asn 355
20	Val	Val	Leu	Arg	Gln 360	Tyr	Glu	Asp	Ket	Val 365	Val	Asp	Glu	Cys	Gly 370
	Cys	Arg 372												•	

What is claimed is:

 A method for alleviating the tissue destructive effects associated with the inflammatory response to tissue injury in a mammal, the method comprising the step of:

providing to the injured tissue a therapeutically effective concentration of a morphogen sufficient to substantially inhibit or reduce the tissue damage resulting from said inflammatory response.

- 2. The method of claim 1 where said step of providing a therapeutically effective morphogen concentration to said injured tissue comprises the step of administering a therapeutically effective concentration of a morphogen to said mammal.
- 3. The method of claim 1 where said step of providing a therapeutically effective morphogen concentration to said injured tissue comprises the step of administering to said mammal an agent that stimulates in vivo a therapeutically effective concentration of an endogenous morphogen.
- 4. The method of claim 1 wherein said step of providing a therapeutically effective concentration of a morphogen is conducted prior to reduction or interruption of blood flow to the tissue.

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- 5. The method of claim 1 wherein said step of providing a therapeutically effective concentration of a morphogen is conducted after reduction or interruption of blood flow to the tissue and before reperfusion.
- 6. The method of claim 1 wherein said step of administering a therapeutically effective amount of a morphogen is conducted following ischemiareperfusion injury.
- 7. The method of claim 1 wherein said said step of administering a therapeutically effective amount of a morphogen is conducted following hyperoxia injury.
- 8. The method of claim 1 wherein said morphogen is provided to said tissue prior to said tissue injury.
- 9. The method of claim 1 wherein said step of providing a therapeutically effective concentration of a morphogen is conducted prior to ischemia-reperfusion injury.
- 10. The method of claim 1 wherein said tissue damage results from an abnormal immune response in said mammal.
- II. The method of claim 1 wherein said tissue damage is associated with an inflammatory disease.
- 12. The method of claim 11 wherein said inflammatory disease is an autoimmune disease.

- 13. The method of claim 11 wherein said inflammatory disease comprises arthritis, psoriasis, dermatitis or diabetes.
- 14. The method of claim 13 wherein said arthritis is rhematoid, degenerative or psoriatic arthritis.
- 15. The method of claim 11 wherein said inflammatory disease comprises an airway inflammation in a mammal.
- 16. The method of claim 15 wherein said airway inflammation comprises chronic bronchitis, emphysema, idiopathic pulmonary fibrosis or asthma.
- 17. The method of claim 11 wherein inflammatory disease comprises a generalized acute inflammatory response.
- 18. The method of claim 17 wherein said inflammatory disease comprises adult respiratory distress syndrome.
- 19. The method of claim 1 wherein said tissue damage is to a transplanted organ or tissue.
- 20. A method for reducing tissue damage associated with ischemia-reperfusion injury in a human, the method comprising the step of:

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providing to the injured tissue a therapeutic concentration of a morphogen sufficient to alleviate the damage associated with said injury.

- 21. A method for reducing the tissue damage associated with hyperoxia injury in a human, the method comprising the step of:
 - providing to the injured tissue a therapeutic concentration of a morphogen sufficient to alleviate the damage associated with said injury.
- 22. The method of claim 20 or 21 wherein said step of providing a therapeutically effective morphogen concentration to said injured tissue comprises the step of administering a therapeutically effective concentration of a morphogen to said mammal.
- 23. The method of claim 20 or 21 wherein said step of providing a therapeutically effective morphogen concentration to said injured tissue comprises the step of administering to said mammal an agent that stimulates in vivo a therapeutically effective concentration of an endogenous morphogen.
- 24. The method of claim 1, 20 or 21 wherein said tissue is lung tissue, cardiac tissue, hepatic tissue or renal tissue.
- 25. The method of claim 6, 9 or 20 wherein said ischemic-reperfusion injury results from cardiac arrest, preliminary occlusion, arterial occlusion, coronary occlusion or occlusive stroke.

- 26. The method of claim 1, 20 or 21 wherein said morphogen comprises an amino acid sequence sharing at least 70% homology with one of the sequences selected from the group consisting of: OP-1, OP-2, CBMP2, Vgl(fx), Vgr(fx), DPP(fx), GDF-1(fx) and 60A(fx).
- 27. The method of claim 26 wherein said morphogen comprises an amino acid sequence sharing a last 80% homology with one of the sequences selected from the group consisting of: OP-1, OP-2, CBMP2, BMP3(fx), BMP5(fx), BMP6(fx), Vg1(fx), Vgr(fx), DPP(fx), GDF-1(fx) and 60A(fx).
- 28. The method of claim 1, 20 or 21 wherein said morphogen comprises an amino acid sequence having greater than 60% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).
- 29. The method of claim 28 wherein said morphogen comprises an amino acid sequence having greater than 65% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).
- 30. The method of claim 29 wherein said morphogen comprises an amino acid sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1), including allelic and species variants thereof.
- 31. The method of claim 1, 20 or 21 wherein said morphogen comprises an amino acid sequence defined by Generic Sequences 1, 2, 3, 4, 5 or 6 (Seq. ID Nos. 1, 2, 3, 4, 30 or 31).

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- 32. The method of claim 1, 20 or 21 wherein said morphogen comprises an amino acid sequence defined by OPX (Seq. ID No. 29).
- 33. A method for reducing the ischemic-reperfusion injury associated with the interruption of blood flow to an organ in a clinical procedure, the method comprising the step of providing a therapeutic concentration of a morphogen to said organ prior to the interruption of blood flow.
- 34. A method for reducing the tissue injury associated with the reduction or interruption of blood flow to an organ or tissue in a clinical procedure, the method comprising the step of providing a therapeutic concentration of a morphogen to said organ or tissue after the reduction or interruption of blood flow to said organ or tissue.
- 35. The method of claim 33 or 34 wherein said clinical procedure is a carotid enterectomy, a coronary artery bypass, a tissue grafting procedure, an organ transplant, or a fibrinolytic therapy.
- 36. The method of claim 1, 33 or 34 wherein said morphogen is administered parenterally.
- 37. The method of claim 1, 33 or 34 wherein said morphogen is administered prophylactically.

- 38. A pharmaceutical composition for use in alleviating the injury associated with tissue exposure to toxic oxygen concentrations comprising a therapeutically effective amount of a morphogen in admixture with a free oxygen radical inhibiting agent or an anticoagulent.
- 39. A pharmaceutical composition for topical administration comprising a therapeutically effective concentration of a morphogen in admixture with a dermatologically acceptable carrier.
- 40. A pharmaceutical composition for topical administration to a tissue comprising a therapeutically effective concentration of a morphogen dispersed in a biocompatible, non-irritating tissue surface adhesive.
- 41. The composition of claim 40 wherein said adhesieve comprises hydroxypropylcellulose.
- 42. The composition of claim 38, 39 or 40 wherein said morphogen comprises an amino acid sequence sharing at least 70% homology with one of the sequences selected from the group consisting of: OP-1, OP-2, CBMP2, Vg1(fx), Vgr(fx), DPP(fx), GDF-1(fx) and 60A(fx).
- 43. The composition of claim 42 wherein said morphogen comprises an amino acid sequence sharing a last 80% homology with one of the sequences selected from the group consisting of: OP-1, OP-2, CBMP2, BMP3(fx), BMP5(fx), BMP6(fx), Vg1(fx), Vgr(fx), DPP(fx), GDF-1(fx) and 60A(fx).

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- 44. The composition of claim 38, 39 or 40 wherein said morphogen comprises an amino acid sequence having greater than 60% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).
- 45. The composition of claim 44 wherein said morphogen comprises an amino acid sequence having greater than 65% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).
- 46. The method of claim 45 wherein said morphogen comprises an amino acid sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1), including allelic and species variants thereof.
- 47. The composition of claim 38, 39 or 40 wherein said morphogen comprises an amino acid sequence defined by Generic Sequences 1, 2, 3, 4, 5 or 6 (Seq. ID Nos. 1, 2, 3, 4, 30 or 31).
- 48. The composition of claim 38, 39 or 40 wherein said morphogen comprises an amino acid sequence defined by OPX (Seq. ID No. 29).
- 49. A method of enhancing the viability of an organ or tissue to be transplanted in a mammal, the method of comprising the step of:

providing a therapeutically effective concentration of a morphogen to said tissue or organ to be transplanted.

- 50. The method of claim 49 wherein said therapeutically effective concentration is sufficient to substantially inhibit reperfusion injury to said tissue or organ.
- 51. The method of claim 49 wherein said morphogen is provided to said tissue or organ prior to reperfusion injury.
- 52. The method of claim 49 wherein said morphogen is provided to said tissue or organ prior to removal of said tissue or organ from the donor.
- 53. The method of claim 49 wherein said organ is placed in an organ preservation solution containing said morphogen or a morphogenstimulating agent after removal of said organ from the donor and prior to transplantation in the recipient.
- 54. The method of claim 49 wherein said organ is selected from the group consisting of lung, heart, kidney, liver or pancreas.
- 55. The method of claim 49 wherein said living tissue comprises skin, bone marrow or gastrointestinal mucosa tissue.

56. A method for protecting a living tissue or transplant organ from the tissue destructive effects associated with the inflammatory response in a mammal, the method comprising the step of:

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providing to said tissue or organ a therapeutically effective concentration of a morphogen.

57. A method of protecting a living tissue or transplanted organ from ischemia-reperfusion injury in a mammal, the method comprising the step of:

providing to said tissue or organ a therapeutically effective concentration of a morphogen, said concentration being sufficient to substantially inhibit or reduce the tissue damage associated with ischemia-reperfusion injury.

- 58. The method of claim 49, 56 or 57 wherein said step of providing a therapeutically effective morphogen concentration to said injured tissue comprises the step of administering a therapeutically effective concentration of a morphogen to said mammal.
- 59. The method of claim 49, 56 or 57 wherein said step of providing a therapeutically effective morphogen concentration to said injured tissue comprises the step of administering to said mammal an agent that stimulates in vivo a therapeutically effective concentration of an endogenous morphogen.

- 60. A composition useful as a living cell or living tissue preservation solution comprising:
 - a fluid formulation having as osmotic pressure substantially equivalent to the osmotic pressure of living mammalian cells in admixture with
 - a therapeutically effective concentration of a morphogen or morphogen-stimulating agent, said concentration being sufficient to protect living cell or tissue from the tissue destructive effects associated with the inflammatory response in a mammal when exposed to said cells or tissue.
- 61. The preservation solution of claim 60 wherein said therapeutically effective concentration is sufficient to substantially inhibit or reduce the tissue damage associated with ischemia-reperfusion injury.
- 62. The preservation solution of claim 60 wherein said formulation further comprises a sugar.
- 63. The preservation solution of claim 60 wherein said formulation further comprises an anticoagulant or a free oxygen radical inhibiting agent.
- 64. The invention of claim 49, 56, 57 or 60 wherein said morphogen comprises an amino acid sequence having greater than 60% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).

- 65. A composition useful in a treatment method to alleviate tissue damage associated with the inflammatory response in a mammal, the composition comprising a therapeutically effective concentration of a morphogen or morphogenstimulating agent.
- 66. The composition of claim 65 wherein said tissue damage is associated with ischemia-reperfusion injury or hyperoxia injury.
- 67. The composition of claim 65 wherein said tissue damage is to lung, cardiac, renalor hepatic tissue.
- 68. The composition of claim 65 wherein said tissue damage is to a transplanted organ or tissue.

AMENDED CLAIMS

[received by the International Bureau on 10 February 1993 (10.02.93); original claims 46 and 49 amended; remaining claims unchanged (1 page)]

- 44. The composition of claim 38, 39 or 40 wherein said morphogen comprises an amino acid sequence having greater than 60% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).
- 45. The composition of claim 44 wherein said morphogen comprises an amino acid sequence having greater than 65% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).
- 46. The composition of claim 45 wherein said morphogen comprises an amino acid sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1), including allelic and species variants thereof.
- 47. The composition of claim 38, 39 or 40 wherein said morphogen comprises an amino acid sequence defined by Generic Sequences 1, 2, 3, 4, 5 or 6 (Seq. ID Nos. 1, 2, 3, 4, 30 or 31).
- 48. The composition of claim 38, 39 or 40 wherein said morphogen comprises an amino acid sequence defined by OPX (Seq. ID No. 29).
- 49. A method of enhancing the viability of an organ or tissue to be transplanted in a mammal, the method comprising the step of:

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providing a therapeutically effective concentration of a morphogen to said tissue or organ to be transplanted.

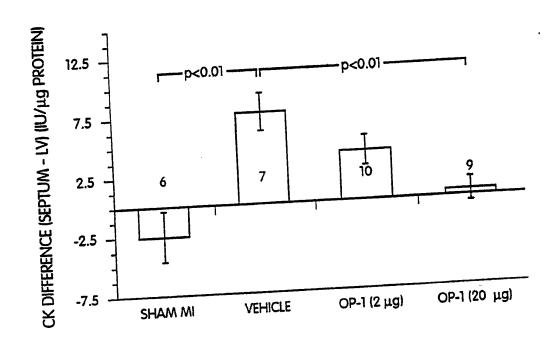


Fig. 1

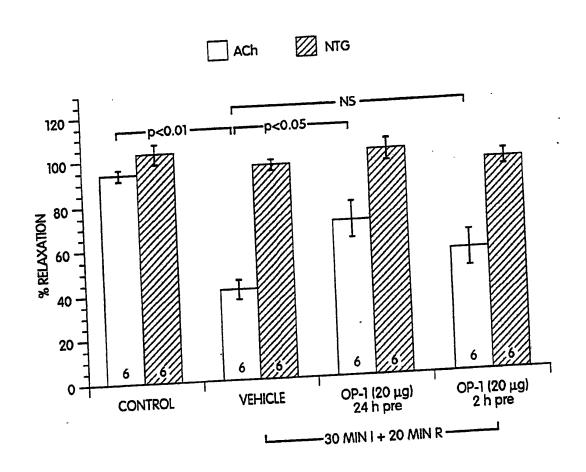


Fig 2

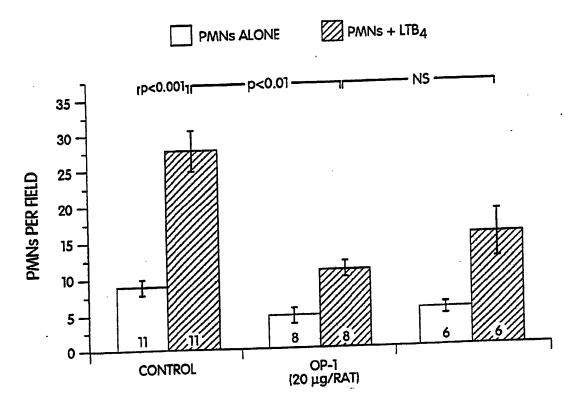
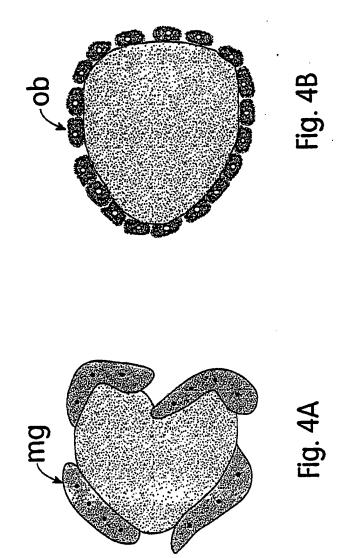
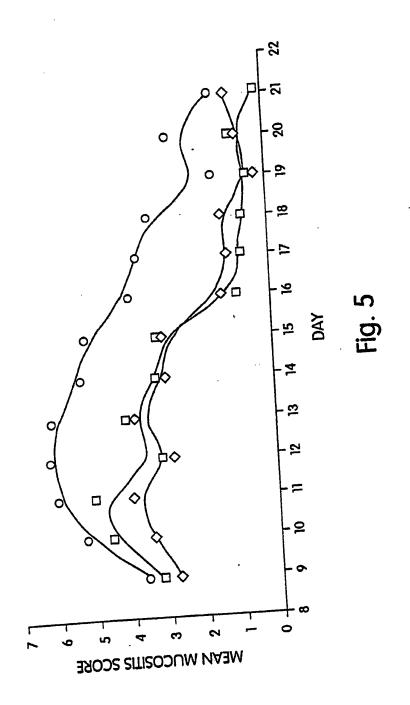


Fig. 3



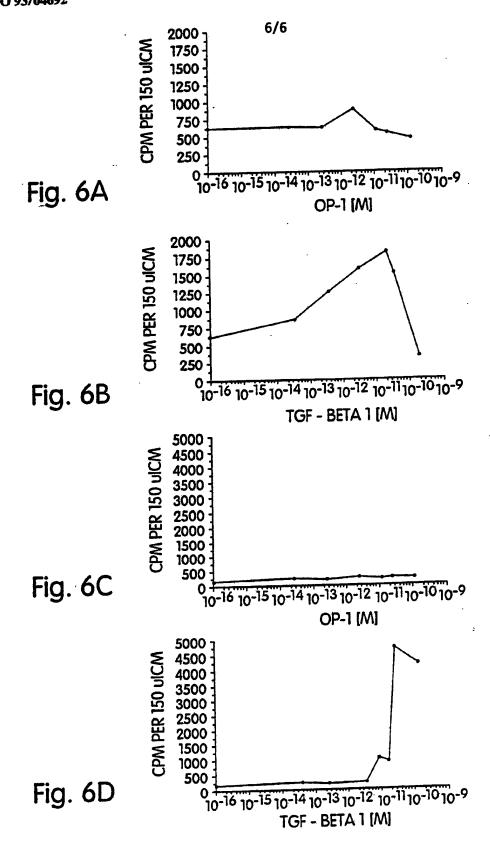
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III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)				
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see the whole document				
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	22-37, 56-59, 64-67			
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Form PCT/ISA/210 (noire sheet) (James y 1985)

INTERNATIONAL SEARCH REPORT

It ational application No. PCT/US 92/07358

•	INTERNATIONAL SEARCH REPORT	
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FURTHER INFORMATION CONTINUED FROM PCT/ISA/210

Remark: Although claims 1-37,49-52 (partially, when the method is carried out in vivo), 54-57 (partially, when the method is carried out in vivo), 58 - 59.64 (partially, according to the method of claims 49,56 or 57) are directed to a method of treatment of the human or animal body the search has been carried out and based on the alleged effects of the composition.

OBSCURITIES, INCONSISTENCIES, CONTRADICTIONS, LACK OF CONCISENESS; LACK OF READY COMPREHENSIBILITY)

(ART. 6 PCT)

REASON:

- Claim 46 has been understood as being dependant of claim 45.
 Therefore claim 46 should read: "The composition of claim 45, wherein said morphogen comprises an amino acid sequence defined by residues 43-139 of Seq. ID No. 5 (hOPI), including allelic and species variants thereof.
- 2. In view of the extremely large number of compounds used in the methods and compositions of claims 26-29, 31 (in as far as seq. ID 1 to 4 and 30-31), 42-45,47 (in as far as seq. ID 1 to 4 and 30-31), 64, the search division considers that it is not economically reasonable to draw up a search report for the methods using, or the compositions comprising all the compounds defined in the claims. The search has therefore been limited, on the basis of the examples and claims, to the methods using, or the compositions comprising the seq. ID no. 5 to 29, 32 and 33 (Art. 17 (2) (a)(ii) and (b) PCT.
- 3. The term "morphogen" is not concise.

Therefor, and for the same reaons as given in paragraph 2 above, it has been understood as being one of the proteins defines in seq. ID 5 to 29, 32 or 33.

(Art. 6 PCT and Art. 17/2)(a)(ii) and (b) PCT)

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ANNEX TO THE INTERNATIONAL SEARCH REPORT ON INTERNATIONAL PATENT APPLICATION NO. US 9207358 64364

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report. The members are as contained in the European Patent Office EDP file on The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information. 19/11/92

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